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(54) Title: TRANSPORTERS AND ION CHANNELS

(57) Abstract: The invention provides human transporters and ion channels (TRICH) and polynucleotides which identify and encode TRICH. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with aberrant expression of TRICH.



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TRANSPORTERS AND ION CHANNELS

TECHNICAL FIELD

This invention relates to nucleic acid and amino acid sequences of transporters and ion channels and to the use of these sequences in the diagnosis, treatment, and prevention of transport, neurological, muscle, immunological, and cell proliferative disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of transporters and ion channels.

BACKGROUND OF THE INVENTION

Eukaryotic cells are surrounded and subdivided into functionally distinct organelles by hydrophobic lipid bilayer membranes which are highly impermeable to most polar molecules. Cells and organelles require transport proteins to import and export essential nutrients and metal ions including K⁺, NH₄⁺, P_i, SO₄²⁻, sugars, and vitamins, as well as various metabolic waste products. Transport proteins also play roles in antibiotic resistance, toxin secretion, ion balance, synaptic neurotransmission, kidney function, intestinal absorption, tumor growth, and other diverse cell functions (Griffith, J. and C. Sansom (1998) The Transporter Facts Book, Academic Press, San Diego CA, pp. 3-29). Transport can occur by a passive concentration-dependent mechanism, or can be linked to an energy source such as ATP hydrolysis or an ion gradient. Proteins that function in transport include carrier proteins, which bind to a specific solute and undergo a conformational change that translocates the bound solute across the membrane, and channel proteins, which form hydrophilic pores that allow specific solutes to diffuse through the membrane down an electrochemical solute gradient.

Carrier proteins which transport a single solute from one side of the membrane to the other are called uniporters. In contrast, coupled transporters link the transfer of one solute with simultaneous or sequential transfer of a second solute, either in the same direction (symport) or in the opposite direction (antiport). For example, intestinal and kidney epithelium contains a variety of symporter systems driven by the sodium gradient that exists across the plasma membrane. Sodium moves into the cell down its electrochemical gradient and brings the solute into the cell with it. The sodium gradient that provides the driving force for solute uptake is maintained by the ubiquitous Na⁺/K⁺ ATPase system. Sodium-coupled transporters include the mammalian glucose transporter (SGLT1), iodide transporter (NIS), and multivitamin transporter (SMVT). All three transporters have twelve putative transmembrane segments, extracellular glycosylation sites, and cytoplasmically-oriented N- and C-termini. NIS plays a crucial role in the evaluation, diagnosis, and treatment of various thyroid pathologies because it is the molecular basis for radioiodide thyroid-imaging

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techniques and for specific targeting of radioisotopes to the thyroid gland (Levy, O. et al. (1997) Proc. Natl. Acad. Sci. USA 94:5568-5573). SMVT is expressed in the intestinal mucosa, kidney, and placenta, and is implicated in the transport of the water-soluble vitamins, e.g., biotin and pantothenate (Prasad, P.D. et al. (1998) J. Biol. Chem. 273:7501-7506).

One of the largest families of transporters is the major facilitator superfamily (MFS), also called the uniporter-symporter-antiporter family. MFS transporters are single polypeptide carriers that transport small solutes in response to ion gradients. Members of the MFS are found in all classes of living organisms, and include transporters for sugars, oligosaccharides, phosphates, nitrates, nucleosides, monocarboxylates, and drugs. MFS transporters found in eukaryotes all have a structure comprising 12 transmembrane segments (Pao, S.S. et al. (1998) Microbiol. Molec. Biol. Rev. 62:1-34). The largest family of MFS transporters is the sugar transporter family, which includes the seven glucose transporters (GLUT1-GLUT7) found in humans that are required for the transport of glucose and other hexose sugars. These glucose transport proteins have unique tissue distributions and physiological functions. GLUT1 provides many cell types with their basal glucose requirements and transports glucose across epithelial and endothelial barrier tissues; GLUT2 facilitates glucose uptake or efflux from the liver; GLUT3 regulates glucose supply to neurons; GLUT4 is responsible for insulin-regulated glucose disposal; and GLUT5 regulates fructose uptake into skeletal muscle. Defects in glucose transporters are involved in a recently identified neurological syndrome causing infantile seizures and developmental delay, as well as glycogen storage disease, Fanconi-Bickel syndrome, and non-insulin-dependent diabetes mellitus (Mueckler, M. (1994) Eur. J. Biochem. 219:713-725; Longo, N. and L.J. Elsas (1998) Adv. Pediatr. 45:293-313).

Monocarboxylate anion transporters are proton-coupled symporters with a broad substrate specificity that includes L-lactate, pyruvate, and the ketone bodies acetate, acetoacetate, and beta-hydroxybutyrate. At least seven isoforms have been identified to date. The isoforms are predicted to have twelve transmembrane (TM) helical domains with a large intracellular loop between TM6 and TM7, and play a critical role in maintaining intracellular pH by removing the protons that are produced stoichiometrically with lactate during glycolysis. The best characterized H⁺-monocarboxylate transporter is that of the erythrocyte membrane, which transports L-lactate and a wide range of other aliphatic monocarboxylates. Other cells possess H⁺-linked monocarboxylate transporters with differing substrate and inhibitor selectivities. In particular, cardiac muscle and tumor cells have transporters that differ in their K_m values for certain substrates, including stereoselectivity for L- over D-lactate, and in their sensitivity to inhibitors. There are Na⁺-monocarboxylate cotransporters on the luminal surface of intestinal and kidney epithelia, which allow the uptake of lactate, pyruvate, and ketone bodies in these tissues. In addition, there are specific and selective transporters for organic cations and organic anions

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in organs including the kidney, intestine and liver. Organic anion transporters are selective for hydrophobic, charged molecules with electron-attracting side groups. Organic cation transporters, such as the ammonium transporter, mediate the secretion of a variety of drugs and endogenous metabolites, and contribute to the maintenance of intercellular pH (Poole, R.C. and A.P. Halestrap (1993) Am. J. Physiol. 264:C761-C782; Price, N.T. et al. (1998) Biochem. J. 329:321-328; and Martinelle, K. and I. Haggstrom (1993) J. Biotechnol. 30:339-350).

ATP-binding cassette (ABC) transporters are members of a superfamily of membrane proteins that transport substances ranging from small molecules such as ions, sugars, amino acids, peptides, and phospholipids, to lipopeptides, large proteins, and complex hydrophobic drugs. ABC transporters consist of four modules: two nucleotide-binding domains (NBD), which hydrolyze ATP to supply the energy required for transport, and two membrane-spanning domains (MSD), each containing six putative transmembrane segments. These four modules may be encoded by a single gene, as is the case for the cystic fibrosis transmembrane regulator (CFTR), or by separate genes. When encoded by separate genes, each gene product contains a single NBD and MSD. These "half-molecules" form homo- and heterodimers, such as Tap1 and Tap2, the endoplasmic reticulum-based major histocompatibility (MHC) peptide transport system. Several genetic diseases are attributed to defects in ABC transporters, such as the following diseases and their corresponding proteins: cystic fibrosis (CFTR, an ion channel), adrenoleukodystrophy (adrenoleukodystrophy protein, ALDP), Zellweger syndrome (peroxisomal membrane protein-70, PMP70), and hyperinsulinemic hypoglycemia (sulfonylurea receptor, SUR). Overexpression of the multidrug resistance (MDR) protein, another ABC transporter, in human cancer cells makes the cells resistant to a variety of cytotoxic drugs used in chemotherapy (Taglicht, D. and S. Michaelis (1998) Meth. Enzymol. 292:130-162).

A number of metal ions such as iron, zinc, copper, cobalt, manganese, molybdenum, selenium, nickel, and chromium are important as cofactors for a number of enzymes. For example, copper is involved in hemoglobin synthesis, connective tissue metabolism, and bone development, by acting as a cofactor in oxidoreductases such as superoxide dismutase, ferroxidase (ceruloplasmin), and lysyl oxidase. Copper and other metal ions must be provided in the diet, and are absorbed by transporters in the gastrointestinal tract. Plasma proteins transport the metal ions to the liver and other target organs, where specific transporters move the ions into cells and cellular organelles as needed. Imbalances in metal ion metabolism have been associated with a number of disease states (Danks, D.M. (1986) J. Med. Genet. 23:99-106).

Transport of fatty acids across the plasma membrane can occur by diffusion, a high capacity, low affinity process. However, under normal physiological conditions a significant fraction of fatty acid transport appears to occur via a high affinity, low capacity protein-mediated transport process.

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Fatty acid transport protein (FATP), an integral membrane protein with four transmembrane segments, is expressed in tissues exhibiting high levels of plasma membrane fatty acid flux, such as muscle, heart, and adipose. Expression of FATP is upregulated in 3T3-L1 cells during adipose conversion, and expression in COS7 fibroblasts elevates uptake of long-chain fatty acids (Hui, T.Y. et al. (1998) J. Biol. Chem. 273:27420-27429).

Mitochondrial carrier proteins are transmembrane-spanning proteins which transport ions and charged metabolites between the cytosol and the mitochondrial matrix. Examples include the ADP, ATP carrier protein; the 2-oxoglutarate/malate carrier; the phosphate carrier protein; the pyruvate carrier; the dicarboxylate carrier which transports malate, succinate, fumarate, and phosphate; the tricarboxylate carrier which transports citrate and malate; and the Grave's disease carrier protein, a protein recognized by IgG in patients with active Grave's disease, an autoimmune disorder resulting in hyperthyroidism. Proteins in this family consist of three tandem repeats of an approximately 100 amino acid domain, each of which contains two transmembrane regions (Stryer, L. (1995) <u>Biochemistry</u>, W.H. Freeman and Company, New York NY, p. 551; PROSITE PDOC00189 Mitochondrial energy transfer proteins signature; Online Mendelian Inheritance in Man (OMIM) *275000 Graves Disease).

This class of transporters also includes the mitochondrial uncoupling proteins, which create proton leaks across the inner mitochondrial membrane, thus uncoupling oxidative phosphorylation from ATP synthesis. The result is energy dissipation in the form of heat. Mitochondrial uncoupling proteins have been implicated as modulators of thermoregulation and metabolic rate, and have been proposed as potential targets for drugs against metabolic diseases such as obesity (Ricquier, D. et al. (1999) J. Int. Med. 245:637-642).

Ion Channels

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The electrical potential of a cell is generated and maintained by controlling the movement of ions across the plasma membrane. The movement of ions requires ion channels, which form ion-selective pores within the membrane. There are two basic types of ion channels, ion transporters and gated ion channels. Ion transporters utilize the energy obtained from ATP hydrolysis to actively transport an ion against the ion's concentration gradient. Gated ion channels allow passive flow of an ion down the ion's electrochemical gradient under restricted conditions. Together, these types of ion channels generate, maintain, and utilize an electrochemical gradient that is used in 1) electrical impulse conduction down the axon of a nerve cell, 2) transport of molecules into cells against concentration gradients, 3) initiation of muscle contraction, and 4) endocrine cell secretion.

Ion Transporters

Ion transporters generate and maintain the resting electrical potential of a cell. Utilizing the energy derived from ATP hydrolysis, they transport ions against the ion's concentration gradient.

These transmembrane ATPases are divided into three families. The phosphorylated (P) class ion transporters, including Na⁺-K⁺ ATPase, Ca²⁺-ATPase, and H⁺-ATPase, are activated by a phosphorylation event. P-class ion transporters are responsible for maintaining resting potential distributions such that cytosolic concentrations of Na⁺ and Ca²⁺ are low and cytosolic concentration of K⁺ is high. The vacuolar (V) class of ion transporters includes H⁺ pumps on intracellular organelles, such as lysosomes and Golgi. V-class ion transporters are responsible for generating the low pH within the lumen of these organelles that is required for function. The coupling factor (F) class consists of H⁺ pumps in the mitochondria. F-class ion transporters utilize a proton gradient to generate ATP from ADP and inorganic phosphate (P_i).

The P-ATPases are hexamers of a 100 kD subunit with ten transmembrane domains and several large cytoplasmic regions that may play a role in ion binding (Scarborough, G.A. (1999) Curr. Opin. Cell Biol. 11:517-522). The V-ATPases are composed of two functional domains: the V_1 domain, a peripheral complex responsible for ATP hydrolysis; and the V_0 domain, an integral complex responsible for proton translocation across the membrane. The F-ATPases are structurally and evolutionarily related to the V-ATPases. The F-ATPase F_0 domain contains 12 copies of the c subunit, a highly hydrophobic protein composed of two transmembrane domains and containing a single buried carboxyl group in TM2 that is essential for proton transport. The V-ATPase V_0 domain contains three types of homologous c subunits with four or five transmembrane domains and the essential carboxyl group in TM4 or TM3. Both types of complex also contain a single a subunit that may be involved in regulating the pH dependence of activity (Forgac, M. (1999) J. Biol. Chem. 274:12951-12954).

The resting potential of the cell is utilized in many processes involving carrier proteins and gated ion channels. Carrier proteins utilize the resting potential to transport molecules into and out of the cell. Amino acid and glucose transport into many cells is linked to sodium ion co-transport (symport) so that the movement of Na⁺ down an electrochemical gradient drives transport of the other molecule up a concentration gradient. Similarly, cardiac muscle links transfer of Ca²⁺ out of the cell with transport of Na⁺ into the cell (antiport).

Gated Ion Channels

Gated ion channels control ion flow by regulating the opening and closing of pores. The ability to control ion flux through various gating mechanisms allows ion channels to mediate such diverse signaling and homeostatic functions as neuronal and endocrine signaling, muscle contraction, fertilization, and regulation of ion and pH balance. Gated ion channels are categorized according to the manner of regulating the gating function. Mechanically-gated channels open their pores in response to mechanical stress; voltage-gated channels (e.g., Na⁺, K⁺, Ca²⁺, and Cl⁻ channels) open their pores in response to changes in membrane potential; and ligand-gated channels (e.g.,

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acetylcholine-, serotonin-, and glutamate-gated cation channels, and GABA- and glycine-gated chloride channels) open their pores in the presence of a specific ion, nucleotide, or neurotransmitter. The gating properties of a particular ion channel (i.e., its threshold for and duration of opening and closing) are sometimes modulated by association with auxiliary channel proteins and/or post translational modifications, such as phosphorylation.

Mechanically-gated or mechanosensitive ion channels act as transducers for the senses of touch, hearing, and balance, and also play important roles in cell volume regulation, smooth muscle contraction, and cardiac rhythm generation. A stretch-inactivated channel (SIC) was recently cloned from rat kidney. The SIC channel belongs to a group of channels which are activated by pressure or stress on the cell membrane and conduct both Ca²⁺ and Na⁺ (Suzuki, M. et al. (1999) J. Biol. Chem. 274:6330-6335).

The pore-forming subunits of the voltage-gated cation channels form a superfamily of ion channel proteins. The characteristic domain of these channel proteins comprises six transmembrane domains (S1-S6), a pore-forming region (P) located between S5 and S6, and intracellular amino and carboxy termini. In the Na⁺ and Ca²⁺ subfamilies, this domain is repeated four times, while in the K⁺ channel subfamily, each channel is formed from a tetramer of either identical or dissimilar subunits. The P region contains information specifying the ion selectivity for the channel. In the case of K⁺ channels, a GYG tripeptide is involved in this selectivity (Ishii, T.M. et al. (1997) Proc. Natl. Acad. Sci. USA 94:11651-11656).

Voltage-gated Na⁺ and K⁺ channels are necessary for the function of electrically excitable cells, such as nerve and muscle cells. Action potentials, which lead to neurotransmitter release and muscle contraction, arise from large, transient changes in the permeability of the membrane to Na⁺ and K⁺ ions. Depolarization of the membrane beyond the threshold level opens voltage-gated Na⁺ channels. Sodium ions flow into the cell, further depolarizing the membrane and opening more voltage-gated Na⁺ channels, which propagates the depolarization down the length of the cell. Depolarization also opens voltage-gated potassium channels. Consequently, potassium ions flow outward, which leads to repolarization of the membrane. Voltage-gated channels utilize charged residues in the fourth transmembrane segment (S4) to sense voltage change. The open state lasts only about 1 millisecond, at which time the channel spontaneously converts into an inactive state that cannot be opened irrespective of the membrane potential. Inactivation is mediated by the channel's N-terminus, which acts as a plug that closes the pore. The transition from an inactive to a closed state requires a return to resting potential.

Voltage-gated Na⁺ channels are heterotrimeric complexes composed of a 260 kDa pore-forming α subunit that associates with two smaller auxiliary subunits, β1 and β2. The β2 subunit is a integral

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membrane glycoprotein that contains an extracellular Ig domain, and its association with α and β 1 subunits correlates with increased functional expression of the channel, a change in its gating properties, as well as an increase in whole cell capacitance due to an increase in membrane surface area (Isom, L.L. et al. (1995) Cell 83:433-442).

Non voltage-gated Na⁺ channels include the members of the amiloride-sensitive Na⁺ channel/degenerin (NaC/DEG) family. Channel subunits of this family are thought to consist of two transmembrane domains flanking a long extracellular loop, with the amino and carboxyl termini located within the cell. The NaC/DEG family includes the epithelial Na⁺ channel (ENaC) involved in Na⁺ reabsorption in epithelia including the airway, distal colon, cortical collecting duct of the kidney, and exocrine duct glands. Mutations in ENaC result in pseudohypoaldosteronism type 1 and Liddle's syndrome (pseudohyperaldosteronism). The NaC/DEG family also includes the recently characterized H⁺-gated cation channels or acid-sensing ion channels (ASIC). ASIC subunits are expressed in the brain and form heteromultimeric Na⁺-permeable channels. These channels require acid pH fluctuations for activation. ASIC subunits show homology to the degenerins, a family of mechanically-gated channels originally isolated from C. elegans. Mutations in the degenerins cause neurodegeneration. ASIC subunits may also have a role in neuronal function, or in pain perception, since tissue acidosis causes pain (Waldmann, R. and M. Lazdunski (1998) Curr. Opin. Neurobiol. 8:418-424; Eglen, R.M. et al. (1999) Trends Pharmacol. Sci. 20:337-342).

K⁺ channels are located in all cell types, and may be regulated by voltage, ATP concentration, or second messengers such as Ca²⁺ and cAMP. In non-excitable tissue, K⁺ channels are involved in protein synthesis, control of endocrine secretions, and the maintenance of osmotic equilibrium across membranes. In neurons and other excitable cells, in addition to regulating action potentials and repolarizing membranes, K⁺ channels are responsible for setting resting membrane potential. The cytosol contains non-diffusible anions and, to balance this net negative charge, the cell contains a Na⁺-K⁺ pump and ion channels that provide the redistribution of Na⁺, K⁺, and Cl⁻. The pump actively transports Na⁺ out of the cell and K⁺ into the cell in a 3:2 ratio. Ion channels in the plasma membrane allow K⁺ and Cl⁻ to flow by passive diffusion. Because of the high negative charge within the cytosol, Cl⁻ flows out of the cell. The flow of K⁺ is balanced by an electromotive force pulling K⁺ into the cell, and a K⁺ concentration gradient pushing K⁺ out of the cell. Thus, the resting membrane potential is primarily regulated by K⁺ flow (Salkoff, L. and T. Jegla (1995) Neuron 15:489-492).

Potassium channel subunits of the <u>Shaker</u>-like superfamily all have the characteristic six transmembrane/1 pore domain structure. Four subunits combine as homo- or heterotetramers to form functional K channels. These pore-forming subunits also associate with various cytoplasmic β subunits that alter channel inactivation kinetics. The <u>Shaker</u>-like channel family includes the voltage-

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gated K⁺ channels as well as the delayed rectifier type channels such as the human ether-a-go-go related gene (HERG) associated with long QT, a cardiac dysrythmia syndrome (Curran, M.E. (1998) Curr. Opin. Biotechnol. 9:565-572; Kaczorowski, G.J. and M.L. Garcia (1999) Curr. Opin. Chem. Biol. 3:448-458).

A second superfamily of K⁺ channels is composed of the inward rectifying channels (Kir). Kir channels have the property of preferentially conducting K⁺ currents in the inward direction. These proteins consist of a single potassium selective pore domain and two transmembrane domains, which correspond to the fifth and sixth transmembrane domains of voltage-gated K⁺ channels. Kir subunits also associate as tetramers. The Kir family includes ROMK1, mutations in which lead to Bartter syndrome, a renal tubular disorder. Kir channels are also involved in regulation of cardiac pacemaker activity, seizures and epilepsy, and insulin regulation (Doupnik, C.A. et al. (1995) Curr. Opin. Neurobiol. 5:268-277; Curran, supra).

The recently recognized TWIK K⁺ channel family includes the mammalian TWIK-1, TREK-1 and TASK proteins. Members of this family possess an overall structure with four transmembrane domains and two P domains. These proteins are probably involved in controlling the resting potential in a large set of cell types (Duprat, F. et al. (1997) EMBO J 16:5464-5471).

The voltage-gated Ca $^{2+}$ channels have been classified into several subtypes based upon their electrophysiological and pharmacological characteristics. L-type Ca $^{2+}$ channels are predominantly expressed in heart and skeletal muscle where they play an essential role in excitation-contraction coupling. T-type channels are important for cardiac pacemaker activity, while N-type and P/Q-type channels are involved in the control of neurotransmitter release in the central and peripheral nervous system. The L-type and N-type voltage-gated Ca $^{2+}$ channels have been purified and, though their functions differ dramatically, they have similar subunit compositions. The channels are composed of three subunits. The α_1 subunit forms the membrane pore and voltage sensor, while the $\alpha_2\delta$ and β subunits modulate the voltage-dependence, gating properties, and the current amplitude of the channel. These subunits are encoded by at least six α_1 , one $\alpha_2\delta$, and four β genes. A fourth subunit, γ , has been identified in skeletal muscle (Walker, D. et al. (1998) J. Biol. Chem. 273:2361-2367; McCleskey, E.W. (1994) Curr. Opin. Neurobiol. 4:304-312).

The transient receptor family (Trp) of calcium ion channels are thought to mediate capacitative calcium entry (CCE). CCE is the Ca²⁺ influx into cells to resupply Ca²⁺ stores depleted by the action of inositol triphosphate (IP3) and other agents in response to numerous hormones and growth factors. Trp and Trp-like were first cloned from Drosophila and have similarity to voltage gated Ca²⁺ channels in the S3 through S6 regions. This suggests that Trp and/or related proteins may form mammalian CCC entry channels (Zhu, X. et al. (1996) Cell 85:661-671; Boulay, G. et al. (1997) J. Biol. Chem.

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272:29672-29680). Melastatin is a gene isolated in both the mouse and human, and whose expression in melanoma cells is inversely correlated with melanoma aggressiveness in vivo. The human cDNA transcript corresponds to a 1533-amino acid protein having homology to members of the Trp family. It has been proposed that the combined use of malastatin mRNA expression status and tumor thickness might allow for the determination of subgroups of patients at both low and high risk for developing metastatic disease (Duncan, L.M. et al (2001) J. Clin. Oncol. 19:568-576).

Chloride channels are necessary in endocrine secretion and in regulation of cytosolic and organelle pH. In secretory epithelial cells, C1 enters the cell across a basolateral membrane through an Na+, K+/C1 cotransporter, accumulating in the cell above its electrochemical equilibrium concentration. Secretion of C1 from the apical surface, in response to hormonal stimulation, leads to flow of Na+ and water into the secretory lumen. The cystic fibrosis transmembrane conductance regulator (CFTR) is a chloride channel encoded by the gene for cystic fibrosis, a common fatal genetic disorder in humans. CFTR is a member of the ABC transporter family, and is composed of two domains each consisting of six transmembrane domains followed by a nucleotide-binding site. Loss of CFTR function decreases transepithelial water secretion and, as a result, the layers of mucus that coat the respiratory tree, pancreatic ducts, and intestine are dehydrated and difficult to clear. The resulting blockage of these sites leads to pancreatic insufficiency, "meconium ileus", and devastating "chronic obstructive pulmonary disease" (Al-Awqati, Q. et al. (1992) J. Exp. Biol. 172:245-266).

The voltage-gated chloride channels (CLC) are characterized by 10-12 transmembrane domains, as well as two small globular domains known as CBS domains. The CLC subunits probably function as homotetramers. CLC proteins are involved in regulation of cell volume, membrane potential stabilization, signal transduction, and transepithelial transport. Mutations in CLC-1, expressed predominantly in skeletal muscle, are responsible for autosomal recessive generalized myotonia and autosomal dominant myotonia congenita, while mutations in the kidney channel CLC-5 lead to kidney stones (Jentsch, T.J. (1996) Curr. Opin. Neurobiol. 6:303-310).

Ligand-gated channels open their pores when an extracellular or intracellular mediator binds to the channel. Neurotransmitter-gated channels are channels that open when a neurotransmitter binds to their extracellular domain. These channels exist in the postsynaptic membrane of nerve or muscle cells. There are two types of neurotransmitter-gated channels. Sodium channels open in response to excitatory neurotransmitters, such as acetylcholine, glutamate, and serotonin. This opening causes an influx of Na^+ and produces the initial localized depolarization that activates the voltage-gated channels and starts the action potential. Chloride channels open in response to inhibitory neurotransmitters, such as γ -aminobutyric acid (GABA) and glycine, leading to hyperpolarization of the membrane and the subsequent generation of an action potential. Neurotransmitter-gated ion channels have four

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transmembrane domains and probably function as pentamers (Jentsch, <u>supra</u>). Amino acids in the second transmembrane domain appear to be important in determining channel permeation and selectivity (Sather, W.A. et al. (1994) Curr. Opin. Neurobiol. 4:313-323).

Ligand-gated channels can be regulated by intracellular second messengers. For example, calcium-activated K^+ channels are gated by internal calcium ions. In nerve cells, an influx of calcium during depolarization opens K^+ channels to modulate the magnitude of the action potential (Ishi et al., supra). The large conductance (BK) channel has been purified from brain and its subunit composition determined. The α subunit of the BK channel has seven rather than six transmembrane domains in contrast to voltage-gated K^+ channels. The extra transmembrane domain is located at the subunit N-terminus. A 28-amino-acid stretch in the C-terminal region of the subunit (the "calcium bowl" region) contains many negatively charged residues and is thought to be the region responsible for calcium binding. The β subunit consists of two transmembrane domains connected by a glycosylated extracellular loop, with intracellular N- and C-termini (Kaczorowski, supra; Vergara, C. et al. (1998) Curr. Opin. Neurobiol. 8:321-329).

Cyclic nucleotide-gated (CNG) channels are gated by cytosolic cyclic nucleotides. The best examples of these are the cAMP-gated Na $^+$ channels involved in olfaction and the cGMP-gated cation channels involved in vision. Both systems involve ligand-mediated activation of a G-protein coupled receptor which then alters the level of cyclic nucleotide within the cell. CNG channels also represent a major pathway for Ca $^{2+}$ entry into neurons, and play roles in neuronal development and plasticity. CNG channels are tetramers containing at least two types of subunits, an α subunit which can form functional homomeric channels, and a β subunit, which modulates the channel properties. All CNG subunits have six transmembrane domains and a pore forming region between the fifth and sixth transmembrane domains, similar to voltage-gated K^+ channels. A large C-terminal domain contains a cyclic nucleotide binding domain, while the N-terminal domain confers variation among channel subtypes (Zufall, F. et al. (1997) Curr. Opin. Neurobiol. 7:404-412).

The activity of other types of ion channel proteins may also be modulated by a variety of intracellular signalling proteins. Many channels have sites for phosphorylation by one or more protein kinases including protein kinase A, protein kinase C, tyrosine kinase, and casein kinase II, all of which regulate ion channel activity in cells. Kir channels are activated by the binding of the $G\beta\gamma$ subunits of heterotrimeric G-proteins (Reimann, F. and F.M. Ashcroft (1999) Curr. Opin. Cell. Biol. 11:503-508). Other proteins are involved in the localization of ion channels to specific sites in the cell membrane. Such proteins include the PDZ domain proteins known as MAGUKs (membrane-associated guanylate kinases) which regulate the clustering of ion channels at neuronal synapses (Craven, S.E. and D.S. Bredt (1998) Cell 93:495-498).

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Disease Correlation

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The etiology of numerous human diseases and disorders can be attributed to defects in the transport of molecules across membranes. Defects in the trafficking of membrane-bound transporters and ion channels are associated with several disorders, e.g., cystic fibrosis, glucose-galactose malabsorption syndrome, hypercholesterolemia, von Gierke disease, and certain forms of diabetes mellitus. Single-gene defect diseases resulting in an inability to transport small molecules across membranes include, e.g., cystinuria, iminoglycinuria, Hartup disease, and Fanconi disease (van't Hoff, W.G. (1996) Exp. Nephrol. 4:253-262; Talente, G.M. et al. (1994) Ann. Intern. Med. 120:218-226; and Chillon, M. et al. (1995) New Engl. J. Med. 332:1475-1480).

Human diseases caused by mutations in ion channel genes include disorders of skeletal muscle, cardiac muscle, and the central nervous system. Mutations in the pore-forming subunits of sodium and chloride channels cause myotonia, a muscle disorder in which relaxation after voluntary contraction is delayed. Sodium channel myotonias have been treated with channel blockers. Mutations in muscle sodium and calcium channels cause forms of periodic paralysis, while mutations in the sarcoplasmic calcium release channel, T-tubule calcium channel, and muscle sodium channel cause malignant hyperthermia. Cardiac arrythmia disorders such as the long QT syndromes and idiopathic ventricular fibrillation are caused by mutations in potassium and sodium channels (Cooper, E.C. and L.Y. Jan (1998) Proc. Natl. Acad. Sci. USA 96:4759-4766). All four known human idiopathic epilepsy genes code for ion channel proteins (Berkovic, S.F. and I.E. Scheffer (1999) Curr. Opin. Neurology 12:177-182). Other neurological disorders such as ataxias, hemiplegic migraine and hereditary deafness can also result from mutations in ion channel genes (Jen, J. (1999) Curr. Opin. Neurobiol. 9:274-280; Cooper, supra).

Ion channels have been the target for many drug therapies. Neurotransmitter-gated channels have been targeted in therapies for treatment of insomnia, anxiety, depression, and schizophrenia. Voltage-gated channels have been targeted in therapies for arrhythmia, ischemic stroke, head trauma, and neurodegenerative disease (Taylor, C.P. and L.S. Narasimhan (1997) Adv. Pharmacol. 39:47-98). Various classes of ion channels also play an important role in the perception of pain, and thus are potential targets for new analgesics. These include the vanilloid-gated ion channels, which are activated by the vanilloid capsaicin, as well as by noxious heat. Local anesthetics such as lidocaine and mexiletine which blockade voltage-gated Na⁺ channels have been useful in the treatment of neuropathic pain (Eglen, supra).

Ion channels in the immune system have recently been suggested as targets for immunomodulation. T-cell activation depends upon calcium signaling, and a diverse set of T-cell specific ion channels has been characterized that affect this signaling process. Channel blocking agents

can inhibit secretion of lymphokines, cell proliferation, and killing of target cells. A peptide antagonist of the T-cell potassium channel Kv1.3 was found to suppress delayed-type hypersensitivity and allogenic responses in pigs, validating the idea of channel blockers as safe and efficacious immunosuppressants (Cahalan, M.D. and K.G. Chandy (1997) Curr. Opin. Biotechnol. 8:749-756).

The discovery of new transporters and ion channels, and the polynucleotides encoding them, satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of transport, neurological, muscle, immunological, and cell proliferative disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of transporters and ion channels.

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SUMMARY OF THE INVENTION

The invention features purified polypeptides, transporters and ion channels, referred to collectively as "TRICH" and individually as "TRICH-1," "TRICH-2," "TRICH-3," "TRICH-4," "TRICH-5," "TRICH-6," "TRICH-6," "TRICH-7," "TRICH-8," "TRICH-9," "TRICH-10," "TRICH-11," "TRICH-11," "TRICH-12," "TRICH-13," "TRICH-14," "TRICH-15," "TRICH-16," "TRICH-17," "TRICH-18," "TRICH-19," "TRICH-20," "TRICH-21," "TRICH-22," "TRICH-23," "TRICH-24," "TRICH-25," "TRICH-26," "TRICH-27," "TRICH-28," "TRICH-29," "TRICH-30," "TRICH-31," and "TRICH-32." In one aspect, the invention provides an isolated polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. In one alternative, the invention provides an isolated polypeptide comprising the amino acid sequence of SEQ ID NO:1-32.

The invention further provides an isolated polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. In one alternative, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-32. In another alternative, the polynucleotide is selected from the group consisting of SEQ ID NO:33-64.

Additionally, the invention provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. In one alternative, the invention provides a cell transformed with the recombinant polynucleotide. In another alternative, the invention provides a transgenic organism comprising the recombinant polynucleotide.

The invention also provides a method for producing a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Additionally, the invention provides an isolated antibody which specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.

The invention further provides an isolated polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). In one alternative, the polynucleotide comprises at least 60 contiguous nucleotides.

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Additionally, the invention provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex, and optionally, if present, the amount thereof. In one alternative, the probe comprises at least 60 contiguous nucleotides.

The invention further provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

The invention further provides a composition comprising an effective amount of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and a pharmaceutically acceptable excipient. In one embodiment, the composition comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The invention additionally provides a method of treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

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The invention also provides a method for screening a compound for effectiveness as an agonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting agonist activity in the sample. In one alternative, the invention provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

Additionally, the invention provides a method for screening a compound for effectiveness as an antagonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. In one alternative, the invention provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with overexpression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

The invention further provides a method of screening for a compound that specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) combining the polypeptide with at least one test compound under

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suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

The invention further provides a method of screening for a compound that modulates the activity of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the presence of the test compound, wherein a change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

The invention further provides a method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence selected from the group consisting of SEQ ID NO:33-64, the method comprising a) exposing a sample comprising the target polynucleotide to a compound, and b) detecting altered expression of the target polynucleotide.

The invention further provides a method for assessing toxicity of a test compound, said method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, iii) a polynucleotide having a sequence complementary to i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, ii) a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, iii) a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, iii) a polynucleotide complementary to the polynucleotide of i), iv) a polynucleotide complementary to the

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polynucleotide of ii), and v) an RNA equivalent of i)-iv). Alternatively, the target polynucleotide comprises a fragment of a polynucleotide sequence selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the present invention.

Table 2 shows the GenBank identification number and annotation of the nearest GenBank homolog for polypeptides of the invention. The probability score for the match between each polypeptide and its GenBank homolog is also shown.

Table 3 shows structural features of polypeptide sequences of the invention, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used for analysis of the polypeptides.

Table 4 lists the cDNA and/or genomic DNA fragments which were used to assemble polynucleotide sequences of the invention, along with selected fragments of the polynucleotide sequences.

Table 5 shows the representative cDNA library for polynucleotides of the invention.

Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze the polynucleotides and polypeptides of the invention, along with applicable descriptions, references, and threshold parameters.

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DESCRIPTION OF THE INVENTION

Before the present proteins, nucleotide sequences, and methods are described, it is understood that this invention is not limited to the particular machines, materials and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a

reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

DEFINITIONS

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"TRICH" refers to the amino acid sequences of substantially purified TRICH obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of . TRICH. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of TRICH either by directly interacting with TRICH or by acting on components of the biological pathway in which TRICH participates.

An "allelic variant" is an alternative form of the gene encoding TRICH. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding TRICH include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as TRICH or a polypeptide with at least one functional characteristic of TRICH. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding TRICH, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding TRICH. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent TRICH. Deliberate amino acid substitutions may be made on the basis of similarity in polarity, charge,

solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of TRICH is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

The terms "amino acid" and "amino acid sequence" refer to an oligopeptide, peptide, polypeptide, or protein sequence, or a fragment of any of these, and to naturally occurring or synthetic molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid sequence.

Amplification is generally carried out using polymerase chain reaction (PCR) technologies well known in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of TRICH. Antagonists may include proteins such as antibodies, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of TRICH either by directly interacting with TRICH or by acting on components of the biological pathway in which TRICH participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')₂, and Fv fragments, which are capable of binding an epitopic determinant.

Antibodies that bind TRICH polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

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The term "antisense" refers to any composition capable of base-pairing with the "sense" (coding) strand of a specific nucleic acid sequence. Antisense compositions may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone linkages such as phosphorothioates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic TRICH, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid sequences that anneal by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

A "composition comprising a given polynucleotide sequence" and a "composition comprising a given amino acid sequence" refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotide sequences encoding TRICH or fragments of TRICH may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, Madison WI) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

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"Conservative amino acid substitutions" are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

| | Original Residue | Conservative Substitution |
|----|------------------|---------------------------|
| | Ala | Gly, Ser |
| | Arg | His, Lys |
| | Asn | Asp, Gln, His |
| 10 | Asp | Asn, Glu |
| | Cys | Ala, Ser |
| | Gln | Asn, Glu, His |
| | Glu . | Asp, Gln, His |
| | Gly | Ala |
| 15 | His | Asn, Arg, Gln, Glu |
| | Ile | Leu, Val |
| | Leu | Ile, Val |
| | Lys | Arg, Gln, Glu |
| | Met | Leu, Ile |
| 20 | Phe | His, Met, Leu, Trp, Tyr |
| | Ser | Cys, Thr |
| | Thr | Ser, Val |
| | Trp | Phe, Tyr |
| | Tyr | His, Phe, Trp |
| 25 | Va1 | Ile, Leu, Thr |

Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to a chemically modified polynucleotide or polypeptide. Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

"Differential expression" refers to increased or upregulated; or decreased, downregulated, or

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absent gene or protein expression, determined by comparing at least two different samples. Such comparisons may be carried out between, for example, a treated and an untreated sample, or a diseased and a normal sample.

A "fragment" is a unique portion of TRICH or the polynucleotide encoding TRICH which is identical in sequence to but shorter in length than the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from 5 to 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:33-64 comprises a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:33-64, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEQ ID NO:33-64 is useful, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:33-64 from related polynucleotide sequences. The precise length of a fragment of SEQ ID NO:33-64 and the region of SEQ ID NO:33-64 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-32 is encoded by a fragment of SEQ ID NO:33-64. A fragment of SEQ ID NO:1-32 comprises a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-32. For example, a fragment of SEQ ID NO:1-32 is useful as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-32. The precise length of a fragment of SEQ ID NO:1-32 and the region of SEQ ID NO:1-32 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A "full length" polynucleotide sequence is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A "full length" polynucleotide sequence encodes a "full length" polypeptide sequence.

"Homology" refers to sequence similarity or, interchangeably, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to

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the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore, achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989) CABIOS 5:151-153 and in Higgins, D.G. et al. (1992) CABIOS 8:189-191.

For pairwise alignments of polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polynucleotide sequences.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/b12.html. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62
Reward for match: 1
Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50

Expect: 10 Word Size: 11

Filter: on

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Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with polynucleotide alignments, the percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

Expect: 10

Word Size: 3

Filter: on

Percent identity may be measured over the length of an entire defined polypeptide sequence, for

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example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about 100 μg/ml sheared, denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5° C to 20° C lower than the thermal melting point ($T_{\rm m}$) for the specific sequence at a defined ionic strength and pH. The $T_{\rm m}$ is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating $T_{\rm m}$ and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, $2^{\rm nd}$ ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; specifically see volume 2, chapter 9.

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68° C in the presence of about $0.2 \times SSC$ and about 0.1% SDS, for 1 hour.

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Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about 100-200 µg/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acid sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., $C_0 t$ or $R_0 t$ analysis) or formed between one nucleic acid sequence present in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or nucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

An "immunogenic fragment" is a polypeptide or oligopeptide fragment of TRICH which is capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide fragment of TRICH which is useful in any of the antibody production methods disclosed herein or known in the art.

The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of TRICH. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of TRICH.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or

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synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an TRICH may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in the art. These processes may occur synthetically or biochemically. Biochemical modifications will vary by cell type depending on the enzymatic milieu of TRICH.

"Probe" refers to nucleic acid sequences encoding TRICH, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acid sequences. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

Methods for preparing and using probes and primers are described in the references, for example Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; Ausubel, F.M. et al. (1987) Current Protocols in Molecular Biology, Greene Publ. Assoc. & Wiley-Intersciences, New York NY; Innis, M. et al. (1990) PCR

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<u>Protocols. A Guide to Methods and Applications</u>, Academic Press, San Diego CA. PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

A "recombinant nucleic acid" is a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook, <u>supra</u>. The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

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Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent, chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA sequence, is composed of the same linear sequence of nucleotides as the reference DNA sequence with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The term "sample" is used in its broadest sense. A sample suspected of containing TRICH, nucleic acids encoding TRICH, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least 60% free, preferably at least 75% free, and most preferably at least 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers,

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microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" refers to the collective pattern of gene expression by a particular cell type or tissue under given conditions at a given time.

"Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection, electroporation, heat shock, lipofection, and particle bombardment. The term "transformed cells" includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

A "transgenic organism," as used herein, is any organism, including but not limited to animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. The term genetic manipulation does not include classical cross-breeding, or in vitro fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection, transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook et al. (1989), supra.

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant

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identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternative splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotide sequences that vary from one species to another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length of one of the polypeptides.

THE INVENTION

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The invention is based on the discovery of new human transporters and ion channels (TRICH), the polynucleotides encoding TRICH, and the use of these compositions for the diagnosis, treatment, or prevention of transport, neurological, muscle, immunological, and cell proliferative disorders.

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide sequence is denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown.

Table 2 shows sequences with homology to the polypeptides of the invention as identified by BLAST analysis against the GenBank protein (genpept) database. Columns 1 and 2 show the polypeptide sequence identification number (Polypeptide SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for polypeptides of the invention. Column 3 shows the GenBank identification number (Genbank ID NO:) of the nearest GenBank homolog. Column 4 shows the probability score for the match between each polypeptide and its GenBank

homolog. Column 5 shows the annotation of the GenBank homolog along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

Table 3 shows various structural features of the polypeptides of the invention. Columns 1 and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites, and column 5 shows potential glycosylation sites, as determined by the MOTIFS program of the GCG sequence analysis software package (Genetics Computer Group, Madison WI). Column 6 shows amino acid residues comprising signature sequences, domains, and motifs. Column 7 shows analytical methods for protein structure/function analysis and in some cases, searchable databases to which the analytical methods were applied.

Together, Tables 2 and 3 summarize the properties of polypeptides of the invention, and these properties establish that the claimed polypeptides are transporters and ion channels. For example, SEQ ID NO:5 is 83% identical to rat GABA receptor rho-3 subunit precursor (GenBank ID g1060975) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.7e-206, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:5 also contains a neurotransmitter-gated ion channel domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:5 is a neurotransmitter-gated ion channel. In an alternate example, SEQ ID NO:16 is 57% identical to human Na+/glucose cotransporter (GenBank ID g338055) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 2.4e-181, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:16 also contains a sodium:solute symporter family domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:16 is a Na+/glucose cotransporter. In an alternate example, SEQ ID NO:27 is 53% identical to human ATP-binding cassette transporter-1 (ABC-1) (GenBank ID g4128033) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:27 also contains an ABC transporter domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from

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BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:27 is an ABC transporter. In an alternate example, SEQ ID NO:12 is 45% identical to rat thyroid sodium/iodide symporter NIS (GenBank ID g1399954) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 3.0e-143, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:12 also contains a sodium:solute symporter family domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:12 is a sodium:solute symporter. SEQ ID NO:1-4, SEQ ID NO:6-11, SEQ ID NO:13-15, SEQ ID NO:17-26, and SEQ ID NO:28-32 were analyzed and annotated in a similar manner. The algorithms and parameters for the analysis of SEQ ID NO:1-32 are described in Table 7.

As shown in Table 4, the full length polynucleotide sequences of the present invention were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Columns 1 and 2 list the polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and the corresponding Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) for each polynucleotide of the invention. Column 3 shows the length of each polynucleotide sequence in basepairs. Column 4 lists fragments of the polynucleotide sequences which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:33-64 or that distinguish between SEQ ID NO:33-64 and related polynucleotide sequences. Column 5 shows identification numbers corresponding to cDNA sequences, coding sequences (exons) predicted from genomic DNA, and/or sequence assemblages comprised of both cDNA and genomic DNA. These sequences were used to assemble the full length polynucleotide sequences of the invention. Columns 6 and 7 of Table 4 show the nucleotide start (5') and stop (3') positions of the cDNA and/or genomic sequences in column 5 relative to their respective full length sequences.

The identification numbers in Column 5 of Table 4 may refer specifically, for example, to Incyte cDNAs along with their corresponding cDNA libraries. For example, 6724643H1 is the identification number of an Incyte cDNA sequence, and LUNLTMT01 is the cDNA library from which it is derived. Incyte cDNAs for which cDNA libraries are not indicated were derived from pooled cDNA libraries (e.g., 71495515V1). Alternatively, the identification numbers in column 5 may refer to GenBank cDNAs or ESTs (e.g., g5746200) which contributed to the assembly of the full length polynucleotide sequences. In addition, the identification numbers in column 5 may identify sequences derived from the ENSEMBL (The Sanger Centre, Cambridge, UK) database (*i.e.*, those sequences

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including the designation "ENST"). Alternatively, the identification numbers in column 5 may be derived from the NCBI RefSeq Nucleotide Sequence Records Database (i.e., those sequences including the designation "NM" or "NT") or the NCBI RefSeq Protein Sequence Records (i.e., those sequences including the designation "NP"). Alternatively, the identification numbers in column 5 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. For example, FL_XXXXXX_N₁_N₂_YYYYY_N₃_N₄ represents a "stitched" sequence in which XXXXXX is the identification number of the cluster of sequences to which the algorithm was applied, and YYYYY is the number of the prediction generated by the algorithm, and $N_{L2.3...}$, if present, represent specific exons that may have been manually edited during analysis (See Example V). Alternatively, the identification numbers in column 5 may refer to assemblages of exons brought together by an "exon-stretching" algorithm. For example, FLXXXXXX_gAAAAA_gBBBBB_1_N is the identification number of a "stretched" sequence, with XXXXXX being the Incyte project identification number, gAAAAA being the GenBank identification number of the human genomic sequence to which the "exon-stretching" algorithm was applied, gBBBBB being the GenBank identification number or NCBI RefSeq identification number of the nearest GenBank protein homolog, and N referring to specific exons (See Example V). In instances where a RefSeq sequence was used as a protein homolog for the "exon-stretching" algorithm, a RefSeq identifier (denoted by "NM," "NP," or "NT") may be used in place of the GenBank identifier (i.e., gBBBBB).

Alternatively, a prefix identifies component sequences that were hand-edited, predicted from genomic DNA sequences, or derived from a combination of sequence analysis methods. The following Table lists examples of component sequence prefixes and corresponding sequence analysis methods associated with the prefixes (see Example IV and Example V).

| Prefix | Type of analysis and/or examples of programs | |
|-----------|--|--|
| GNN, GFG, | Exon prediction from genomic sequences using, for example, | |
| ENST | GENSCAN (Stanford University, CA, USA) or FGENES | |
| | (Computer Genomics Group, The Sanger Centre, Cambridge, UK). | |
| GBI | Hand-edited analysis of genomic sequences. | |
| FL | Stitched or stretched genomic sequences (see Example V). | |

In some cases, Incyte cDNA coverage redundant with the sequence coverage shown in column 5 was obtained to confirm the final consensus polynucleotide sequence, but the relevant Incyte cDNA identification numbers are not shown.

Table 5 shows the representative cDNA libraries for those full length polynucleotide sequences which were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte

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cDNA library which is most frequently represented by the Incyte cDNA sequences which were used to assemble and confirm the above polynucleotide sequences. The tissues and vectors which were used to construct the cDNA libraries shown in Table 5 are described in Table 6.

The invention also encompasses TRICH variants. A preferred TRICH variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid sequence identity to the TRICH amino acid sequence, and which contains at least one functional or structural characteristic of TRICH.

The invention also encompasses polynucleotides which encode TRICH. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:33-64, which encodes TRICH. The polynucleotide sequences of SEQ ID NO:33-64, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The invention also encompasses a variant of a polynucleotide sequence encoding TRICH. In particular, such a variant polynucleotide sequence will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding TRICH. A particular aspect of the invention encompasses a variant of a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:33-64 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:33-64. Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of TRICH.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding TRICH, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring TRICH, and all such variations are to be considered as being specifically disclosed.

Although nucleotide sequences which encode TRICH and its variants are generally capable of hybridizing to the nucleotide sequence of the naturally occurring TRICH under appropriately selected conditions of stringency, it may be advantageous to produce nucleotide sequences encoding TRICH or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide

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occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding TRICH and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

The invention also encompasses production of DNA sequences which encode TRICH and TRICH derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding TRICH or any fragment thereof.

Also encompassed by the invention are polynucleotide sequences that are capable of hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID NO:33-64 and fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol. 152:507-511.) Hybridization conditions, including annealing and wash conditions, are described in "Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied Biosystems), thermostable T7 polymerase (Amersham Pharmacia Biotech, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Life Technologies, Gaithersburg MD). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Applied Biosystems), the MEGABACE 1000 DNA sequencing system (Molecular Dynamics, Sunnyvale CA), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art. (See, e.g., Ausubel, F.M. (1997) Short Protocols in Molecular Biology, John Wiley & Sons, New York NY, unit 7.7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853.)

The nucleic acid sequences encoding TRICH may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic

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DNA within a cloning vector. (See, e.g., Sarkar, G. (1993) PCR Methods Applic. 2:318-322.) Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences. (See, e.g., Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186.) A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA. (See, e.g., Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to 72°C.

When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode TRICH may be cloned in recombinant DNA molecules that direct expression of TRICH, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of

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the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence may be produced and used to express TRICH.

The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter TRICH-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create-new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent Number 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of TRICH, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then subjected to selection or screening procedures that identify those gene variants with the desired properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

In another embodiment, sequences encoding TRICH may be synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; and Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232.) Alternatively, TRICH itself or a fragment thereof may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques. (See, e.g., Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH Freeman, New York NY, pp. 55-60; and Roberge, J.Y. et al. (1995) Science 269:202-204.) Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems). Additionally, the amino acid sequence of TRICH, or any part thereof, may be altered during direct synthesis and/or combined with sequences

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from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing. (See, e.g., Creighton, supra, pp. 28-53.)

In order to express a biologically active TRICH, the nucleotide sequences encoding TRICH or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotide sequences encoding TRICH. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of sequences encoding TRICH. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where sequences encoding TRICH and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used. (See, e.g., Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162.)

Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding TRICH and appropriate transcriptional and translational control elements. These methods include <u>in vitro</u> recombinant DNA techniques, synthetic techniques, and <u>in vivo</u> genetic recombination. (See, e.g., Sambrook, J. et al. (1989) <u>Molecular Cloning, A Laboratory Manual</u>, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and 16-17; Ausubel, F.M. et al. (1995) <u>Current Protocols in Molecular Biology</u>, John Wiley & Sons, New York NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express sequences encoding TRICH. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or

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animal cell systems. (See, e.g., Sambrook, <u>supra</u>; Ausubel, <u>supra</u>; Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509; Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945; Takamatsu, N. (1987) EMBO J. 6:307-311; <u>The McGraw Hill Yearbook of Science and Technology</u> (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659; and Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.) Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. (See, e.g., Di Nicola, M. et al. (1998) Cancer Gen. Ther. 5(6):350-356; Yu, M. et al. (1993) Proc. Natl. Acad. Sci. USA 90(13):6340-6344; Buller, R.M. et al. (1985) Nature 317(6040):813-815; McGregor, D.P. et al. (1994) Mol. Immunol. 31(3):219-226; and Verma, I.M. and N. Somia (1997) Nature 389:239-242.) The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotide sequences encoding TRICH. For example, routine cloning, subcloning, and propagation of polynucleotide sequences encoding TRICH can be achieved using a multifunctional E. coli vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Life Technologies). Ligation of sequences encoding TRICH into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for <u>in vitro</u> transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence. (See, e.g., Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509.) When large quantities of TRICH are needed, e.g. for the production of antibodies, vectors which direct high level expression of TRICH may be used. For example, vectors containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of TRICH. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast <u>Saccharomyces cerevisiae</u> or <u>Pichia pastoris</u>. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign sequences into the host genome for stable propagation. (See, e.g., Ausubel, 1995, <u>supra;</u> Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; and Scorer, C.A. et al. (1994) Bio/Technology 12:181-184.)

Plant systems may also be used for expression of TRICH. Transcription of sequences encoding TRICH may be driven by viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311).

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Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See, e.g., Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; and Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105.) These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. (See, e.g., The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196.)

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, sequences encoding TRICH may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses TRICH in host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659.) In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.)

For long term production of recombinant proteins in mammalian systems, stable expression of TRICH in cell lines is preferred. For example, sequences encoding TRICH can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in tk and apr cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823.) Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, dlifr confers resistance to methotrexate; neo confers resistance to the aminoglycosides neomycin and G-418; and als and pat confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., Wigler, M. et al. (1980)

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Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14.) Additional selectable genes have been described, e.g., *trpB* and *hisD*, which alter cellular requirements for metabolites. (See, e.g., Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051.) Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), β glucuronidase and its substrate β-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system. (See, e.g., Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131.)

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding TRICH is inserted within a marker gene sequence, transformed cells containing sequences encoding TRICH can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding TRICH under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

In general, host cells that contain the nucleic acid sequence encoding TRICH and that express TRICH may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

Immunological methods for detecting and measuring the expression of TRICH using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on TRICH is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art. (See, e.g., Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; and Pound, J.D. (1998) Immunochemical Protocols, Humana Press, Totowa NJ.)

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding TRICH include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide.

Alternatively, the sequences encoding TRICH, or any fragments thereof, may be cloned into a vector for

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the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes <u>in vitro</u> by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Pharmacia Biotech, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with nucleotide sequences encoding TRICH may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode TRICH may be designed to contain signal sequences which direct secretion of TRICH through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding TRICH may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric TRICH protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of TRICH activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, *c-myc*, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, respectively. FLAG, *c-myc*, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize

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these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the TRICH encoding sequence and the heterologous protein sequence, so that TRICH may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel (1995, supra, ch. 10). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

In a further embodiment of the invention, synthesis of radiolabeled TRICH may be achieved <u>in vitro</u> using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, ³⁵S-methionine.

TRICH of the present invention or fragments thereof may be used to screen for compounds that specifically bind to TRICH. At least one and up to a plurality of test compounds may be screened for specific binding to TRICH. Examples of test compounds include antibodies, oligonucleotides, proteins (e.g., receptors), or small molecules.

In one embodiment, the compound thus identified is closely related to the natural ligand of TRICH, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner. (See, e.g., Coligan, J.E. et al. (1991) <u>Current Protocols in Immunology</u> 1(2): Chapter 5.) Similarly, the compound can be closely related to the natural receptor to which TRICH binds, or to at least a fragment of the receptor, e.g., the ligand binding site. In either case, the compound can be rationally designed using known techniques. In one embodiment, screening for these compounds involves producing appropriate cells which express TRICH, either as a secreted protein or on the cell membrane. Preferred cells include cells from mammals, yeast, <u>Drosophila</u>, or <u>E. coli</u>. Cells expressing TRICH or cell membrane fractions which contain TRICH are then contacted with a test compound and binding, stimulation, or inhibition of activity of either TRICH or the compound is analyzed.

An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with TRICH, either in solution or affixed to a solid support, and detecting the binding of TRICH to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

TRICH of the present invention or fragments thereof may be used to screen for compounds that modulate the activity of TRICH. Such compounds may include agonists, antagonists, or partial

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or inverse agonists. In one embodiment, an assay is performed under conditions permissive for TRICH activity, wherein TRICH is combined with at least one test compound, and the activity of TRICH in the presence of a test compound is compared with the activity of TRICH in the absence of the test compound. A change in the activity of TRICH in the presence of the test compound is indicative of a compound that modulates the activity of TRICH. Alternatively, a test compound is combined with an in vitro or cell-free system comprising TRICH under conditions suitable for TRICH activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of TRICH may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

In another embodiment, polynucleotides encoding TRICH or their mammalian homologs may be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES) cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease. (See, e.g., U.S. Patent Number 5,175,383 and U.S. Patent Number 5,767,337.) For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and grown in culture. The ES cells are transformed with a vector containing the gene of interest disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (neo; Capecchi, M.R. (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or developmental stage-specific manner (Marth, J.D. (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330). Transformed ES cells are identified and microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding TRICH may also be manipulated <u>in vitro</u> in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding TRICH can also be used to create "knockin" humanized animals (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a region of a polynucleotide encoding TRICH is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with

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potential pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a mammal inbred to overexpress TRICH, e.g., by secreting TRICH in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74).

THERAPEUTICS

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Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of TRICH and transporters and ion channels. In addition, the expression of TRICH is closely associated with adrenal, testicular, and prostate tumors, Crohn's disease, teratocarcinoma and dendritic cells, brain, lung, ileum, small intestine, uterine myometrial, colon, and pancreatic tissues. Therefore, TRICH appears to play a role in transport, neurological, muscle, immunological, and cell proliferative disorders. In the treatment of disorders associated with increased TRICH expression or activity, it is desirable to decrease the expression or activity of TRICH. In the treatment of disorders associated with decreased TRICH expression or activity, it is desirable to increase the expression or activity of TRICH.

Therefore, in one embodiment, TRICH or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH. Examples of such disorders include, but are not limited to, a transport disorder such as akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral neoplasms, prostate cancer, cardiac disorders associated with transport, e.g., angina, bradyarrythmia, tachyarrythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, polymyositis, neurological disorders associated with transport, e.g., Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia, and other disorders associated with transport, e.g., neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, hypercholesterolemia, adrenoleukodystrophy, Zellweger syndrome, Menkes disease, occipital horn syndrome, von Gierke disease, cystinuria, iminoglycinuria, Hartup disease, and Fanconi disease; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other

extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a muscle disorder such as cardiomyopathy, myocarditis, Duchenne's muscular dystrophy, Becker's muscular dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, pheochromocytoma, and myopathies including encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, ophthalmoplegia, and acid maltase deficiency (AMD, also known as Pompe's disease); an immunological disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal

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circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus.

In another embodiment, a vector capable of expressing TRICH or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those described above.

In a further embodiment, a composition comprising a substantially purified TRICH in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of TRICH may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those listed above.

In a further embodiment, an antagonist of TRICH may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of TRICH. Examples of such disorders include, but are not limited to, those transport, neurological, muscle, immunological, and cell proliferative disorders described above. In one aspect, an antibody which specifically binds TRICH may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express TRICH.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding TRICH may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of TRICH including, but not limited to, those described above.

In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary sequences, or vectors of the invention may be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

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An antagonist of TRICH may be produced using methods which are generally known in the art. In particular, purified TRICH may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind TRICH. Antibodies to TRICH may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are generally preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, humans, and others may be immunized by injection with TRICH or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially preferable.

It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to TRICH have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein. Short stretches of TRICH amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to TRICH may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. (See, e.g., Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; and Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120.)

In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; and Takeda, S. et al. (1985) Nature 314:452-454.) Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce TRICH-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be

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generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137.)

Antibodies may also be produced by inducing <u>in vivo</u> production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299.)

Antibody fragments which contain specific binding sites for TRICH may also be generated. For example, such fragments include, but are not limited to, $F(ab')_2$ fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the $F(ab')_2$ fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. et al. (1989) Science 246:1275-1281.)

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between TRICH and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering TRICH epitopes is generally used, but a competitive binding assay may also be employed (Pound, supra).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for TRICH. Affinity is expressed as an association constant, K_a , which is defined as the molar concentration of TRICH-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K_a determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple TRICH epitopes, represents the average affinity, or avidity, of the antibodies for TRICH. The K_a determined for a preparation of monoclonal antibodies, which are monospecific for a particular TRICH epitope, represents a true measure of affinity. High-affinity antibody preparations with K_a ranging from about 10^9 to 10^{12} L/mole are preferred for use in immunoassays in which the TRICH-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K_a ranging from about 10^6 to 10^7 L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of TRICH, preferably in active form, from the antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

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The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of TRICH-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available. (See, e.g., Catty, supra, and Coligan et al. supra.)

In another embodiment of the invention, the polynucleotides encoding TRICH, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding TRICH. Such technology is well known in the art, and antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding TRICH. (See, e.g., Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press Inc., Totawa NJ.)

In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence complementary to at least a portion of the cellular sequence encoding the target protein. (See, e.g., Slater, J.E. et al. (1998) J. Allergy Clin. Immunol. 102(3):469-475; and Scanlon, K.J. et al. (1995) 9(13):1288-1296.) Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and adeno-associated virus vectors. (See, e.g., Miller, A.D. (1990) Blood 76:271; Ausubel, supra; Uckert, W. and W. Walther (1994) Pharmacol. Ther. 63(3):323-347.) Other gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art. (See, e.g., Rossi, J.J. (1995) Br. Med. Bull. 51(1):217-225; Boado, R.J. et al. (1998) J. Pharm. Sci. 87(11):1308-1315; and Morris, M.C. et al. (1997) Nucleic Acids Res. 25(14):2730-2736.)

In another embodiment of the invention, polynucleotides encoding TRICH may be used for somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by X-linked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase (ADA) deficiency (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), thalassamias, familial

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hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 389:239-242)), (ii) express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA. 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida albicans and Paracoccidioides brasiliensis; and protozoan parasites such as Plasmodium falciparum and Trypanosoma cruzi). In the case where a genetic deficiency in TRICH expression or regulation causes disease, the expression of TRICH from an appropriate population of transduced cells may alleviate the clinical manifestations caused by the genetic deficiency.

In a further embodiment of the invention, diseases or disorders caused by deficiencies in TRICH are treated by constructing mammalian expression vectors encoding TRICH and introducing these vectors by mechanical means into TRICH-deficient cells. Mechanical transfer technologies for use with cells <u>in vivo</u> or <u>ex vitro</u> include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev. Biochem. 62:191-217; Ivics, Z. (1997) Cell 91:501-510; Boulay, J-L. and H. Récipon (1998) Curr. Opin. Biotechnol. 9:445-450).

Expression vectors that may be effective for the expression of TRICH include, but are not limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). TRICH may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β-actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc. Natl. Acad. Sci. USA 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen)); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi, F.M.V. and Blau, H.M. supra)), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding TRICH from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver

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polynucleotides to target cells in culture and require minimal effort to optimize experimental parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al. (1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these standardized mammalian transfection protocols.

In another embodiment of the invention, diseases or disorders caused by genetic defects with respect to TRICH expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding TRICH under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus cis-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et al. (1998) J. Virol. 72:9873-9880). U.S. Patent Number 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant") discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4⁺ T-cells), and the return of transduced cells to a patient are procedures well known to persons skilled in the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

In the alternative, an adenovirus-based gene therapy delivery system is used to deliver polynucleotides encoding TRICH to cells which have one or more genetic abnormalities with respect to the expression of TRICH. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S. Patent Number 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999) Annu.

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Rev. Nutr. 19:511-544 and Verma, I.M. and N. Somia (1997) Nature 18:389:239-242, both incorporated by reference herein.

In another alternative, a herpes-based, gene therapy delivery system is used to deliver polynucleotides encoding TRICH to target cells which have one or more genetic abnormalities with respect to the expression of TRICH. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing TRICH to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent Number 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent Number 5,804,413 teaches the use of recombinant HSV d92 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the appropriate promoter for purposes including human gene therapy. Also taught by this patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999) J. Virol. 73:519-532 and Xu, H. et al. (1994) Dev. Biol. 163:152-161, hereby incorporated by reference. The manipulation of cloned herpesvirus sequences, the generation of recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to those of ordinary skill in the art.

In another alternative, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding TRICH to target cells. The biology of the prototypic alphavirus, Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA, resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for TRICH into the alphavirus genome in place of the capsid-coding region results in the production of a large number of TRICH-coding RNAs and the synthesis of high levels of TRICH in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy application

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(Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of TRICH into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177.) A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences encoding TRICH.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by <u>in vitro</u> and <u>in vivo</u> transcription of DNA sequences encoding TRICH. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

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RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding TRICH.

Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased TRICH expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding TRICH may be therapeutically useful, and in the treatment of disorders associated with decreased TRICH expression or activity, a compound which specifically promotes expression of the polynucleotide encoding TRICH may be therapeutically useful.

At least one, and up to a plurality, of test compounds may be screened for effectiveness in altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective in altering polynucleotide expression; selection from an existing, commercially-available or proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding TRICH is exposed to at least one test compound thus obtained. The sample may comprise, for example, an intact or permeabilized cell, or an in vitro cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding TRICH are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding TRICH. The amount of hybridization may be quantified, thus forming the basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of

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the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a Schizosaccharomyces pombe gene expression system (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

Many methods for introducing vectors into cells or tissues are available and equally suitable for use <u>in vivo</u>, <u>in vitro</u>, and <u>ex vivo</u>. For <u>ex vivo</u> therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466.)

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable excipient. Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may consist of TRICH, antibodies to TRICH, and mimetics, agonists, antagonists, or inhibitors of TRICH.

The compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

Compositions for pulmonary administration may be prepared in liquid or dry powder form. These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S.

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Patent No. 5,997,848). Pulmonary delivery has the advantage of administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising TRICH or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, TRICH or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example TRICH or fragments thereof, antibodies of TRICH, and agonists, antagonists or inhibitors of TRICH, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED_{50} (the dose therapeutically effective in 50% of the population) or LD_{50} (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the LD_{50}/ED_{50} ratio. Compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED_{50} with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy.

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Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about $0.1 \mu g$ to $100,000 \mu g$, up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

DIAGNOSTICS

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In another embodiment, antibodies which specifically bind TRICH may be used for the diagnosis of disorders characterized by expression of TRICH, or in assays to monitor patients being treated with TRICH or agonists, antagonists, or inhibitors of TRICH. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for TRICH include methods which utilize the antibody and a label to detect TRICH in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring TRICH, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of TRICH expression. Normal or standard values for TRICH expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibodies to TRICH under conditions suitable for complex formation. The amount of standard complex formation may be quantitated by various methods, such as photometric means. Quantities of TRICH expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values.

Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, the polynucleotides encoding TRICH may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of TRICH may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of TRICH, and to monitor regulation of TRICH levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding TRICH or closely related molecules may be used to identify nucleic acid sequences which encode TRICH. The specificity of the probe, whether it is made

from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding TRICH, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the TRICH encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:33-64 or from genomic sequences including promoters, enhancers, and introns of the TRICH gene.

Means for producing specific hybridization probes for DNAs encoding TRICH include the cloning of polynucleotide sequences encoding TRICH or TRICH derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes <u>in vitro</u> by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as ³²P or ³⁵S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotide sequences encoding TRICH may be used for the diagnosis of disorders associated with expression of TRICH. Examples of such disorders include, but are not limited to, a transport disorder such as akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral neoplasms, prostate cancer, cardiac disorders associated with transport, e.g., angina, bradyarrythmia, tachyarrythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, polymyositis, neurological disorders associated with transport, e.g., Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia, and other disorders associated with transport, e.g., neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, hypercholesterolemia, adrenoleukodystrophy, Zellweger syndrome, Menkes disease, occipital horn syndrome, von Gierke disease, cystinuria, iminoglycinuria, Hartup disease, and

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Fanconi disease; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a muscle disorder such as cardiomyopathy, myocarditis, Duchenne's muscular dystrophy, Becker's muscular dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, pheochromocytoma, and myopathies including encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, ophthalmoplegia, and acid maltase deficiency (AMD, also known as Pompe's disease); an immunological disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic

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anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus. The polynucleotide sequences encoding TRICH may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered TRICH expression. Such qualitative or quantitative methods are well known in the art.

In a particular aspect, the nucleotide sequences encoding TRICH may be useful in assays that detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding TRICH may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of nucleotide sequences encoding TRICH in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of TRICH, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding TRICH, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

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Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding TRICH may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced in vitro. Oligomers will preferably contain a fragment of a polynucleotide encoding TRICH, or a fragment of a polynucleotide complementary to the polynucleotide encoding TRICH, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

In a particular aspect, oligonucleotide primers derived from the polynucleotide sequences encoding TRICH may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from the polynucleotide sequences encoding TRICH are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSCCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in high-throughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed in silico SNP (isSNP), are capable of identifying polymorphisms by comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computerbased methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the

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alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

Methods which may also be used to quantify the expression of TRICH include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229-236.) The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotide sequences described herein may be used as elements on a microarray. The microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described below. The microarray may also be used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of disease as a function of gene expression, and to develop and monitor the activities of therapeutic agents in the treatment of disease. In particular, this information may be used to develop a pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

In another embodiment, TRICH, fragments of TRICH, or antibodies specific for TRICH may be used as elements on a microarray. The microarray may be used to monitor or measure protein-protein interactions, drug-target interactions, and gene expression profiles, as described above.

A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time. (See Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent Number 5,840,484, expressly incorporated by reference herein.) Thus a transcript image may be generated by hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

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Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression <u>in vivo</u>, as in the case of a tissue or biopsy sample, or <u>in vitro</u>, as in the case of a cell line.

Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with in vitro model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000) Toxicol. Lett. 112-113:467-471, expressly incorporated by reference herein). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity, (See, for example, Press Release 00-02 from the National Institute of Environmental Health Sciences, released February 29, 2000, available at http://www.niehs.nih.gov/oc/news/toxchip.htm.) Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

In one embodiment, the toxicity of a test compound is assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

Another particular embodiment relates to the use of the polypeptide sequences of the present invention to analyze the proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are

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analyzed by quantifying the number of expressed proteins and their relative abundance under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, supra). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots from different samples, for example, from biological samples either treated or untreated with a test compound or therapeutic agent, are compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous amino acid residues, to the polypeptide sequences of the present invention. In some cases, further sequence data may be obtained for definitive protein identification.

A proteomic profile may also be generated using antibodies specific for TRICH to quantify the levels of TRICH expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by exposing the microarray to the sample and detecting the levels of protein bound to each array element (Lucking, A. et al. (1999) Anal. Biochem. 270:103-111; Mendoze, L.G. et al. (1999) Biotechniques 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiol- or amino-reactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

Toxicant signatures at the proteome level are also useful for toxicological screening, and should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) Electrophoresis 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference

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in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art. (See, e.g., Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; and Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662.) Various types of microarrays are well known and thoroughly described in <u>DNA Microarrays: A Practical Approach</u>, M. Schena, ed. (1999) Oxford University Press, London, hereby expressly incorporated by reference.

In another embodiment of the invention, nucleic acid sequences encoding TRICH may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; and Trask, B.J. (1991) Trends Genet. 7:149-154.) Once mapped, the nucleic acid sequences of the invention may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism (RFLP). (See, for example, Lander, E.S. and D. Botstein (1986) Proc. Natl. Acad. Sci. USA 83:7353-7357.)

Fluorescent <u>in situ</u> hybridization (FISH) may be correlated with other physical and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, <u>supra</u>, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding TRICH on a physical

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map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation. (See, e.g., Gatti, R.A. et al. (1988) Nature 336:577-580.) The nucleotide sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, TRICH, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between TRICH and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with TRICH, or fragments thereof, and washed. Bound TRICH is then detected by methods well known in the art. Purified TRICH can also be coated directly onto plates for use in the aforementioned drug screening techniques.

Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding TRICH specifically compete with a test compound for binding TRICH. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with TRICH.

In additional embodiments, the nucleotide sequences which encode TRICH may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

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solid support.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications and publications, mentioned above and below including U.S. Ser. No. 60/216,547, U.S. Ser. No. 60/218,232, U.S. Ser. No. 60/220,112, and U.S. Ser. No. 60/221,839 are expressly incorporated by reference herein., are expressly incorporated by reference herein.

10 EXAMPLES

I. Construction of cDNA Libraries

Incyte cDNAs were derived from cDNA libraries described in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA) and shown in Table 4, column 5. Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A)+ RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Life Technologies), using the recommended procedures or similar methods known in the art. (See, e.g., Ausubel, 1997, supra, units 5.1-6.6.) Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Pharmacia Biotech) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g.,

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PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Life Technologies), PCDNA2.1 plasmid (Invitrogen, Carlsbad CA), PBK-CMV plasmid (Stratagene), or pINCY (Incyte Genomics, Palo Alto CA), or derivatives thereof. Recombinant plasmids were transformed into competent <u>E. coli</u> cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5α, DH10B, or ElectroMAX DH10B from Life Technologies.

II. Isolation of cDNA Clones

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Plasmids obtained as described in Example I were recovered from host cells by <u>in vivo</u> excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

20 III. Sequencing and Analysis

Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows. Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Pharmacia Biotech or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Molecular Dynamics); the ABI PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (reviewed in Ausubel, 1997, supra, unit 7.7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VIII.

The polynucleotide sequences derived from Incyte cDNAs were validated by removing vector, linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The Incyte cDNA sequences or translations thereof were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM, and hidden Markov model (HMM)-based protein family databases such as PFAM. (HMM is a probabilistic approach which analyzes consensus primary structures of gene families. See, for example, Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries were performed using programs based on BLAST, FASTA, BLIMPS, and HMMER. The Incyte cDNA sequences were assembled to produce full length polynucleotide sequences. Alternatively, GenBank cDNAs, GenBank ESTs, stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) were used to extend Incyte cDNA assemblages to full length. Assembly was performed using programs based on Phred, Phrap, and Consed, and cDNA assemblages were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length polypeptide sequences. Alternatively, a polypeptide of the invention may begin at any of the methionine residues of the full length translated polypeptide. Full length polypeptide sequences were subsequently analyzed by querying against databases such as the GenBank protein databases (genpept), SwissProt, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, and hidden Markov model (HMM)-based protein family databases such as PFAM. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (Hitachi Software Engineering, South San Francisco CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent

Table 7 summarizes the tools, programs, and algorithms used for the analysis and assembly of Incyte cDNA and full length sequences and provides applicable descriptions, references, and threshold parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score or the lower the probability value, the greater the identity between two sequences).

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identity between aligned sequences.

The programs described above for the assembly and analysis of full length polynucleotide and polypeptide sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:33-64. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 4.

5 IV. Identification and Editing of Coding Sequences from Genomic DNA

Putative transporters and ion channels were initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a general-purpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (See Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94, and Burge, C. and S. Karlin (1998) Curr. Opin. Struct. Biol. 8:346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a FASTA database of polynucleotide and polypeptide sequences. The maximum range of sequence for Genscan to analyze at once was set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode transporters and ion channels, the encoded polypeptides were analyzed by querying against PFAM models for transporters and ion channels. Potential transporters and ion channels were also identified by homology to Incyte cDNA sequences that had been annotated as transporters and ion channels. These selected Genscan-predicted sequences were then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscan-predicted sequences were then edited by comparison to the top BLAST hit from genpept to correct errors in the sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis was also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage was available, this information was used to correct or confirm the Genscan predicted sequence. Full length polynucleotide sequences were obtained by assembling Genscan-predicted coding sequences with Incyte cDNA sequences and/or public cDNA sequences using the assembly process described in Example III. Alternatively, full length polynucleotide sequences were derived entirely from edited or unedited Genscan-predicted coding sequences.

V. Assembly of Genomic Sequence Data with cDNA Sequence Data "Stitched" Sequences

Partial cDNA sequences were extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example III were mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster was analyzed using an algorithm based on graph theory and dynamic programming to integrate cDNA and genomic information, generating possible splice variants that were subsequently confirmed, edited, or extended to create a full length sequence.

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Sequence intervals in which the entire length of the interval was present on more than one sequence in the cluster were identified, and intervals thus identified were considered to be equivalent by transitivity. For example, if an interval was present on a cDNA and two genomic sequences, then all three intervals were considered to be equivalent. This process allows unrelated but consecutive genomic sequences to be brought together, bridged by cDNA sequence. Intervals thus identified were then "stitched" together by the stitching algorithm in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence to genomic sequence) were given preference over linkages which change parent type (cDNA to genomic sequence). The resultant stitched sequences were translated and compared by BLAST analysis to the genpept and gbpri public databases. Incorrect exons predicted by Genscan were corrected by comparison to the top BLAST hit from genpept. Sequences were further extended with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

"Stretched" Sequences

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Partial DNA sequences were extended to full length with an algorithm based on BLAST analysis. First, partial cDNAs assembled as described in Example III were queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases using the BLAST program. The nearest GenBank protein homolog was then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein was generated by using the resultant high-scoring segment pairs (HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both were used as probes to search for homologous genomic sequences from the public human genome databases. Partial DNA sequences were therefore "stretched" or extended by the addition of homologous genomic sequences. The resultant stretched sequences were examined to determine whether it contained a complete gene.

VI. Chromosomal Mapping of TRICH Encoding Polynucleotides

The sequences which were used to assemble SEQ ID NO:33-64 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:33-64 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences

had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, of human chromosomes. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's p-arm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (http.//www.ncbi.nlm.nih.gov/genemap/), can be employed to determine if previously identified disease genes map within or in proximity to the intervals indicated above.

VII. Analysis of Polynucleotide Expression

Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound. (See, e.g., Sambrook, <u>supra</u>, ch. 7; Ausubel (1995) <u>supra</u>, ch. 4 and 16.)

Analogous computer techniques applying BLAST were used to search for identical or related molecules in cDNA databases such as GenBank or LIFESEQ (Incyte Genomics). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer. search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

BLAST Score x Percent Identity

5 x minimum {length(Seq. 1), length(Seq. 2)}

The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For

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example, a product score of 100 is produced only for 100% identity over the entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

Alternatively, polynucleotide sequences encoding TRICH are analyzed with respect to the tissue sources from which they were derived. For example, some full length sequences are assembled, at least in part, with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived from a cDNA library constructed from a human tissue. Each human tissue is classified into one of the following organ/tissue categories: cardiovascular system; connective tissue; digestive system; embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells; hemic and immune system; liver; musculoskeletal system; nervous system; pancreas; respiratory system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number of libraries in each category is counted and divided by the total number of libraries across all categories. Similarly, each human tissue is classified into one of the following disease/condition categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled, and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding TRICH. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA).

VIII. Extension of TRICH Encoding Polynucleotides

Full length polynucleotide sequences were also produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer was synthesized to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg^{2+} , $(NH_4)_2SO_4$, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Pharmacia Biotech), ELONGASE enzyme

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(Life Technologies), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

The concentration of DNA in each well was determined by dispensing 100 μ l PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 μ l of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II (Labsysteins Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5 μ l to 10 μ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1% agarose gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Pharmacia Biotech). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Pharmacia Biotech), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent <u>E. coli</u> cells. Transformed cells were selected on antibiotic-containing media, and individual colonies were picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase (Amersham Pharmacia Biotech) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries were reamplified using the same conditions as described above. Samples were diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Pharmacia Biotech) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

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In like manner, full length polynucleotide sequences are verified using the above procedure or are used to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

IX. Labeling and Use of Individual Hybridization Probes

Hybridization probes derived from SEQ ID NO:33-64 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μ Ci of [γ - 32 P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Pharmacia Biotech). An aliquot containing 10^7 counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

X. Microarrays

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The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing, See, e.g., Baldeschweiler, <u>supra.</u>), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned technologies should be uniform and solid with a non-porous surface (Schena (1999), <u>supra</u>). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers. Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements. (See, e.g., Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645; Marshall, A. and J. Hodgson (1998) Nat. Biotechnol. 16:27-31.)

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection. After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorbtion and mass spectrometry may be used for detection of hybridization. The degree of complementarity and the relative abundance of each polynucleotide which hybridizes to an element on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

<u>Tissue or Cell Sample Preparation</u>

Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and poly(A)⁺ RNA is purified using the oligo-(dT) cellulose method. Each poly(A)⁺ RNA sample is reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/μl oligo-(dT) primer (21mer), 1X first strand buffer, 0.03 units/μl RNase inhibitor, 500 μM dATP, 500 μM dGTP, 500 μM dTTP, 40 μM dCTP, 40 μM dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Pharmacia Biotech). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)⁺ RNA with GEMBRIGHT kits (Incyte). Specific control poly(A)⁺ RNAs are synthesized by in vitro transcription from non-coding yeast genomic DNA. After incubation at 37°C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85°C to the stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (CLONTECH Laboratories, Inc. (CLONTECH), Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 μl 5X SSC/0.2% SDS.

Microarray Preparation

Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 µg. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Pharmacia Biotech).

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Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in US Patent No. 5,807,522, incorporated herein by reference. 1 μ l of the array element DNA, at an average concentration of 100 ng/ μ l, is loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2% SDS and distilled water as before.

Hybridization

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Hybridization reactions contain 9 µl of sample mixture consisting of 0.2 µg each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140 µl of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X SSC), and dried.

25 Detection

Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially. Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate

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filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte).

XI. Complementary Polynucleotides

Sequences complementary to the TRICH-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring TRICH. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of TRICH. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the TRICH-encoding transcript.

XII. Expression of TRICH

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Expression and purification of TRICH is achieved using bacterial or virus-based expression systems. For expression of TRICH in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the trp-lac (tac) hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the *lac* operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express TRICH upon induction with isopropyl beta-Dthiogalactopyranoside (IPTG). Expression of TRICH in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding TRICH by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription. Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus. (See Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945.)

In most expression systems, TRICH is synthesized as a fusion protein with, e.g., glutathione Stransferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Pharmacia Biotech). Following purification, the GST moiety can be proteolytically cleaved from TRICH at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel (1995, supra, ch. 10 and 16). Purified TRICH obtained by these methods can be used directly in the assays shown in Examples XVI, XVII, and XVIII where applicable.

30 XIII. Functional Assays

TRICH function is assessed by expressing the sequences encoding TRICH at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include PCMV SPORT (Life Technologies) and PCR3.1 (Invitrogen, Carlsbad CA), both of which

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contain the cytomegalovirus promoter. 5-10 µg of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2 μ g of an additional plasmid containing sequences encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser opticsbased technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; downregulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994) Flow Cytometry, Oxford, New York NY.

The influence of TRICH on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding TRICH and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding TRICH and other genes of interest can be analyzed by northern analysis or microarray techniques.

XIV. Production of TRICH Specific Antibodies

TRICH substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols.

Alternatively, the TRICH amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art. (See, e.g., Ausubel, 1995, <u>supra</u>, ch. 11.)

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Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Applied Biosystems) using FMOC chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity. (See, e.g., Ausubel, 1995, supra.) Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-TRICH activity by, for example, binding the peptide or TRICH to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

XV. Purification of Naturally Occurring TRICH Using Specific Antibodies

Naturally occurring or recombinant TRICH is substantially purified by immunoaffinity chromatography using antibodies specific for TRICH. An immunoaffinity column is constructed by covalently coupling anti-TRICH antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing TRICH are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of TRICH (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/TRICH binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and TRICH is collected.

XVI. Identification of Molecules Which Interact with TRICH

Molecules which interact with TRICH may include transporter substrates, agonists or antagonists, modulatory proteins such as Gβγ proteins (Reimann, supra) or proteins involved in TRICH localization or clustering such as MAGUKs (Craven, supra). TRICH, or biologically active fragments thereof, are labeled with ¹²⁵I Bolton-Hunter reagent. (See, e.g., Bolton A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled TRICH, washed, and any wells with labeled TRICH complex are assayed. Data obtained using different concentrations of TRICH are used to calculate values for the number, affinity, and association of TRICH with the candidate molecules.

Alternatively, proteins that interact with TRICH are isolated using the yeast 2-hybrid system (Fields, S. and O. Song (1989) Nature 340:245-246). TRICH, or fragments thereof, are expressed as fusion proteins with the DNA binding domain of Gal4 or lexA, and potential interacting proteins are expressed as fusion proteins with an activation domain. Interactions between the TRICH fusion protein and the TRICH interacting proteins (fusion proteins with an activation domain) reconstitute a transactivation function that is observed by expression of a reporter gene. Yeast 2-hybrid systems are

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commercially available, and methods for use of the yeast 2-hybrid system with ion channel proteins are discussed in Niethammer, M. and M. Sheng (1998, Meth. Enzymol. 293:104-122).

TRICH may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

Potential TRICH agonists or antagonists may be tested for activation or inhibition of TRICH ion channel activity using the assays described in section XVIII.

XVII. Demonstration of TRICH Activity

Ion channel activity of TRICH is demonstrated using an electrophysiological assay for ion conductance. TRICH can be expressed by transforming a mammalian cell line such as COS7, HeLa or CHO with a eukaryotic expression vector encoding TRICH. Eukaryotic expression vectors are commercially available, and the techniques to introduce them into cells are well known to those skilled in the art. A second plasmid which expresses any one of a number of marker genes, such as β-galactosidase, is co-transformed into the cells to allow rapid identification of those cells which have taken up and expressed the foreign DNA. The cells are incubated for 48-72 hours after transformation under conditions appropriate for the cell line to allow expression and accumulation of TRICH and β-galactosidase.

Transformed cells expressing β-galactosidase are stained blue when a suitable colorimetric substrate is added to the culture media under conditions that are well known in the art. Stained cells are tested for differences in membrane conductance by electrophysiological techniques that are well known in the art. Untransformed cells, and/or cells transformed with either vector sequences alone or β-galactosidase sequences alone, are used as controls and tested in parallel. Cells expressing TRICH will have higher anion or cation conductance relative to control cells. The contribution of TRICH to conductance can be confirmed by incubating the cells using antibodies specific for TRICH. The antibodies will bind to the extracellular side of TRICH, thereby blocking the pore in the ion channel, and the associated conductance.

Alternatively, ion channel activity of TRICH is measured as current flow across a TRICH-containing Xenopus laevis oocyte membrane using the two-electrode voltage-clamp technique (Ishi et al., supra; Jegla, T. and L. Salkoff (1997) J. Neurosci. 17:32-44). TRICH is subcloned into an appropriate Xenopus oocyte expression vector, such as pBF, and 0.5-5 ng of mRNA is injected into mature stage IV oocytes. Injected oocytes are incubated at 18°C for 1-5 days. Inside-out macropatches are excised into an intracellular solution containing 116 mM K-gluconate, 4 mM KCl, and 10 mM Hepes (pH 7.2). The intracellular solution is supplemented with varying concentrations of the TRICH mediator, such as cAMP, cGMP, or Ca⁺² (in the form of CaCl₂), where appropriate.

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Electrode resistance is set at 2-5 M Ω and electrodes are filled with the intracellular solution lacking mediator. Experiments are performed at room temperature from a holding potential of 0 mV. Voltage ramps (2.5 s) from -100 to 100 mV are acquired at a sampling frequency of 500 Hz. Current measured is proportional to the activity of TRICH in the assay.

In particular, the activities of TRICH-1, TRICH-2, and TRICH-10, are measured as K⁺ conductance, the activities of TRICH-6 and TRICH-9 are measured as K⁺ conductance in the presence of membrane stretch or free fatty acids, the activities of TRICH-18, TRICH-25 and TRICH-31 are measured as voltage-gated K⁺ conductance, TRICH-5 activity is measured as Cl⁻ conductance in the presence of GABA, TRICH-11 activity is measured as cation conductance in the presence of heat, and the activity of TRICH-9, TRICH-28 is measured as Ca²⁺ conductance.

Transport activity of TRICH is assayed by measuring uptake of labeled substrates into Xenopus laevis oocytes. Oocytes at stages V and VI are injected with TRICH mRNA (10 ng per oocyte) and incubated for 3 days at 18°C in OR2 medium (82.5mM NaCl, 2.5 mM KCl, 1mM CaCl₂, 1mM MgCl₂, 1mM Na₂HPO₄, 5 mM Hepes, 3.8 mM NaOH, 50μg/ml gentamycin, pH 7.8) to allow expression of TRICH. Oocytes are then transferred to standard uptake medium (100mM NaCl, 2 mM KCl, 1mM CaCl₂, 1mM MgCl₂, 10 mM Hepes/Tris pH 7.5). Uptake of various substrates (e.g., amino acids, sugars, drugs, ions, and neurotransmitters) is initiated by adding labeled substrate (e.g. radiolabeled with ³H, fluorescently labeled with rhodamine, etc.) to the oocytes. After incubating for 30 minutes, uptake is terminated by washing the oocytes three times in Na⁺-free medium, measuring the incorporated label, and comparing with controls. TRICH activity is proportional to the level of internalized labeled substrate. In particular, test substrates include pigment precursors and related molecules for TRICH-3, aminophospholipids for TRICH-4, fructose and glucose for TRICH-13 and TRICH-15, amino acids for TRICH-8, Na⁺ and iodide for TRICH-12, Na⁺ and H⁺ for TRICH-13 and TRICH-21, Na⁺ and glucose for TRICH-16 and TRICH-19, and glucose for TRICH-23, TRICH-26, TRICH-29, TRICH-30, and TRICH-32.

ATPase activity associated with TRICH can be measured by hydrolysis of radiolabeled ATP- $[\gamma^{-32}P]$, separation of the hydrolysis products by chromatographic methods, and quantitation of the recovered ^{32}P using a scintillation counter. The reaction mixture contains ATP- $[\gamma^{-32}P]$ and varying amounts of TRICH in a suitable buffer incubated at 37 °C for a suitable period of time. The reaction is terminated by acid precipitation with trichloroacetic acid and then neutralized with base, and an aliquot of the reaction mixture is subjected to membrane or filter paper-based chromatography to separate the reaction products. The amount of ^{32}P liberated is counted in a scintillation counter. The amount of radioactivity recovered is proportional to the ATPase activity of TRICH in the assay.

XVIII. Identification of TRICH Agonists and Antagonists

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TRICH is expressed in a eukaryotic cell line such as CHO (Chinese Hamster Ovary) or HEK (Human Embryonic Kidney) 293. Ion channel activity of the transformed cells is measured in the presence and absence of candidate agonists or antagonists. Ion channel activity is assayed using patch clamp methods well known in the art or as described in Example XVII. Alternatively, ion channel activity is assayed using fluorescent techniques that measure ion flux across the cell membrane (Velicelebi, G. et al. (1999) Meth. Enzymol. 294:20-47; West, M.R. and C.R. Molloy (1996) Anal. Biochem. 241:51-58). These assays may be adapted for high-throughput screening using microplates. Changes in internal ion concentration are measured using fluorescent dyes such as the Ca²⁺ indicator Fluo-4 AM, sodium-sensitive dyes such as SBFI and sodium green, or the Cl⁻ indicator MQAE (all available from Molecular Probes) in combination with the FLIPR fluorimetric plate reading system (Molecular Devices). In a more generic version of this assay, changes in membrane potential caused by ionic flux across the plasma membrane are measured using oxonyl dyes such as DiBAC4 (Molecular Probes). DiBAC₄ equilibrates between the extracellular solution and cellular sites according to the cellular membrane potential. The dye's fluorescence intensity is 20-fold greater when bound to hydrophobic intracellular sites, allowing detection of DiBAC4 entry into the cell (Gonzalez, J.E. and P.A. Negulescu (1998) Curr. Opin. Biotechnol. 9:624-631). Candidate agonists or antagonists may be selected from known ion channel agonists or antagonists, peptide libraries, or combinatorial chemical libraries.

Various modifications and variations of the described methods and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following claims.

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Table 1

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|----------------|-------------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|-----------|------------|------------|------------|
| Incyte | Polynucleotide ID | 3474673CB1 | 4588877CB1 | 7472214CB1 | 7473053CB1 | 7473347CB1 | 7474240CB1 | 7475338CB1 | 7476747CB1 | 7477898CB1 | 7472728CB1 | 7474322CB1 | 5455621CB1 | 7477248CB1 | 2944004CB1 | 3046849CB1 | 4538363CB1 | 6427460CB1 | 7474127CB1 | 7476949CB1 | 7477249CB1 | 7477720CB1 | 7477852CB1 | 1471717CB1 | 3874406CB1 | 4599654CB1 | 5047435CB1 | 7475603CB1 | 7477845CB1 | 168827CB1 | 7472734CB1 | 7473473CB1 | 7477755CR1 |
| Polynucleotide | SEQ ID NO: | 33 | 34 | 35 | 36 | 37 | 38 | 3.9 | 40 | 41 | 42 | 43 | 77 | 45 | 46 | 47 | 48 | 49 | 50 | 51 | 52 | 53 | 54 | 55 | 56 | 57 | | | | | | 63 | 64 |
| Incyte | Polypeptide ID | 3474673CD1 | 4588877CD1 | 7472214CD1 | 7473053CD1 | 7473347CD1 | 7474240CD1 | 7475338CD1 | 7476747CD1 | 7477898CD1 | 7472728CD1 | 7474322CD1 | 5455621CD1 | 7477248CD1 | 2944004CD1 | 3046849CD1 | 4538363CD1 | 6427460CD1 | 7474127CD1 | 7476949CD1 | 7477249CD1 | 7477720CD1 | 7477852CD1 | 1471717CD1 | 3874406CD1 | 4599654CD1 | 5047435CD1 | 7475603CD1 | 7477845CD1 | 168827CD1 | 7472734CD1 | 7473473CD1 | 7477725CD1 |
| Polypeptide | SEQ ID NO: | | 2 | 3 | 4 | | 9 | 7 | 8 | 6 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | 20 | 21 | | 23 | 24 | 25 | 26 | 27 | 28 | 29 | 30 | 31 | 32 |
| Incyte | Project ID | 3474673 | 4588877 | 7472214 | 7473053 | 7473347 | 7474240 | 7475338 | 7476747 | 7477898 | 7472728 | 7474322 | 5455621 | 7477248 | 2944004 | 3046849 | 4538363 | 6427460 | 7474127 | 7476949 | 7477249 | 7477720 | 7477852 | 1471717 | 3874406 | 4599654 | 5047435 | 7475603 | 7477845 | 168827 | 7472734 | 7473473 | 7477725 |

Table 2

| Dolymentide | Thryte | ConBank ID | Drohability | Can Bank Homolog |
|-------------|-------------------|------------|-------------|--|
| SEQ ID NO: | Polypeptide ID | NO: | score | |
| | 3474673CD1 | g13507377 | 1.00E-151 | [fl][Homo sapiens] potassium channel TASK-4 (Decher, N. et al. (2001) FEBS Lett. 492 (1-2), 84-89) |
| | 4588877CD1 | g13926111 | 3.00E-96 | [fl][Homo sapiens] (AF358910) 2P domain potassium channel Talk-2 |
| | 7472214CD1 | g1107730 | 1.70E-243 | [Mus musculus] ABC8 (Savary, S. et al. (1996) Mamm. Genome 7 (9), 673-676) |
| | | g11342541 | 0 | [f1][Homo sapiens] putative white family ATP-binding cassette transporter |
| | 7473053CD1 | g3850108 | 9.00E-209 | [Schizosaccharomyces pombe] putative calcium- transporting atpase |
| | | g3628757 | 0 | [Homo sapiens] FIC1 (Bull,L.N. et al. (1998) Nat. Genet. 18 (3), 219-224) |
| | 7473347CD1 | g1060975 | 1.70E-206 | [Rattus norvegicus] GABA receptor rho-3 subunit |
| | | | | precursor (Ogurusu,T. et al. (1996) Biochim. Biophys. Acta 1305 (1-2), 15-18) |
| | 7474240CD1 | g2745727 | 0 | [Rattus norvegicus] potassium channel (Shi, W. et al. (1997) J. Neurosci. 17 (24), 9423-9432) |
| | 7475338CD1 | g183298 | 2.10E-158 | GLUTS protein al. (1990) J. Biol. Chem. 265 |
| | 7477898CD1 | g2745729 | 0 | [Rattus norvegicus] potassium channel (Shi, W. et al. (1997) J. Neurosci. 17 (24), 9423-9432) |
| 10 | 7472728CD1 | g8452900 | 3.50E-261 | norvegicus] potassium channel TREK-2 . et al. (2000) J. Biol. Chem. 275 (2 |
| 11 | 7474322CD1 | g12003146 | 0 | [f1][Homo sapiens] capsaicin receptor |
| 12 | 5455621CD1 | g1399954 | 3.00E-143 | 3] thyroid a |
| 13 | 7477248CD1 | g2944233 | 3.10E-195 | apiens] sodium-hydrogen exchanger, M. et al. (1998) J. Biol. Chem. 2 |
| | 2944004CD1 | g3451312 | 1.40E-188 | [Schizosaccharomyces pombe] membrane atpase |
| 15 | 3046849CD1 | g12802047 | 0 | |

| Polypeptide | Incyte | GenBank ID | Probability | GenBank Homolog |
|-------------|----------------|------------|-------------|--|
| SEQ ID NO: | Polypeptide ID | NO: | score | |
| 16 | 4538363CD1 | g338055 | 7.40E-181 | [Homo sapiens] Na+/glucose cotransporter (Hediger, M.A. et al. (1989) Proc. Natl. Acad. Sci. U.S.A. 86 (15), 5748-5752) |
| 17 | 6427460CD1 | g6457274 | 0 | [Mus musculus] putative E1-E2 ATPase (Halleck, M.S. et al. (1999) Physiol. Genomics (Online) 1 (3), 139-150) |
| 18 | 7474127CD1 | g206044 | 0 | [Rattus norvegicus] potassium channel Kv3.2b (Wiedmann, R. et al. (1991) FEBS Lett. 288, 163-167) |
| 19 | 7476949CD1 | g9588428 | 0 | ter family member simil |
| | | 9338055 | 3.70E-202 | [Homo sapiens] Na+/glucose cotransporter (Hediger, M.A. et al. (1989) Proc. Natl. Acad. Sci. U.S.A. 86 (15), 5748-5752) |
| 20 | 7477249CD1 | g7715417 | 0 | [Oryctolagus cuniculus] RING-finger binding protein (Mansharamani, M. et al. (2001) J. Biol. Chem. 276 (5), 3641-3649) |
| 21 | 7477720CD1 | g205709 | 0 | [Rattus norvegicus] sodium-hydrogen exchange protein- isoform 4 (Orlowski, J. et al. (1992) J. Biol. Chem. 267, 9331- 9339) |
| 22 | 7477852CD1 | g8920219 | 0 | [f1][Homo sapiens] epithelial calcium channel (Muller, D. et al. (2000) Genomics 67 (1), 48-53) |
| 23 | 1471717CD1 | g529590 | 5.00E-36 | |
| 24 | 3874406CD1 | g1514530 | 1.90E-117 | [Homo sapiens] ABC-C transporter (Klugbauer, N. et al. (1996) FEBS Lett. 391 (1-2), 61- 65) |
| 25 | 4599654CDI | g3242244 | 0 | [Mus musculus] hyperpolarization-activated cation channel, HAC3 (Ludwig, A. et al. (1998) Nature 393 (6685), 587-591) |

Table 3

| Analytical | Methods and | Databases | HMMER | HMMER_PFAM | HMMER | BLAST_PRODOM | | HMMER | | HMMER_PFAM | BLIMPS_BLOCKS | PROFILESCAN | BLAST_PRODOM | | | BLAST_DOMO | BLAST_DOMO | | MOTIFS | MOTIFS | |
|----------------------|--------------------|------------|--|-------------------------------------|------------------------------------|--------------|---|------------------------|---------------------------------|-------------------------------------|--|--|---------------------------------|-----------------------------|---------------------|---|-------------------------|---------------------------------|-------------------------------------|--------------------------------|-----------|
| Signature Sequences, | Domains and Motifs | | Transmembrane domains: R130-M155, V245-L264 | TASK K+ channel domain: V14-S332 | Transmembrane domain: V139-L158 | M SUBUNIT | AT FULMILYE SUBFAMILY N MEMBER PD021430: A78-E162 | Transmembrane domains: | S430-M450, W564-D589, M618-V637 | ABC transporter domain: R95-G277 | ABC transporters family signature BL00211: I100-F111, I201-D232 | ABC transporters family signature: V181-D232 | PROTEIN TRANSMEMBRANE TRANSPORT | GLYCOPROTEIN INNER PUTATIVE | PD000633: T365-Y583 | <pre>do WHITE; FRUIT; FLY; SCARLET; DM05200 P45844 289-650: G277-L623</pre> | ABC TRANSPORTERS FAMILY | DM00000 F43044 /3-20/: 101-02/0 | ABC transporter motif: L201-L215 | ATP/GTP binding site (P-loop): | G102-S109 |
| Potential | | tion Sites | N65 N94 | | | | | N169 N422 | | | | | | | | | | | | | - |
| Potential | Phosphorylation | | 320 328 | S289.S51 T169 T67 | S101 S128 S159 S174 S175 S183 | S95 | | 8229 | S34 | S554 S57 S644 S69 S89 T138 | T157 T23 T472 T500 T591 | | | | | | | | | | |
| Amino | | es | 332 | | 226 | | | 646 | | | | | | | | | | | - | | |
| Incyte | lypeptide | A | 3474673CD1 | | 4588877CD1 | | | 7472214CD1 | | | | | | | | | | | | | |
| SEQ | Ωï | 0N | ٦. | | 2 | | | m | - | | | | | | | | | _ | | | |

| Analytical | Methods and | Databases | HMMER | HMMER_PFAM | | BLIMPS_BLOCKS | | PROFILESCAN | BLIMPS PRINTS | | | BLAST_PRODOM | | | BLAST_PRODOM | BLAST_DOMO | | MOTIFS | HMMER | | |
|----------------------|-----------------|------------|--|---|------|------------------------------------|--------------------------------------|-------------------------------------|-----------------------------------|------------|------|--------------------------------|---|--------------------------------|------------------------------------|-----------------------------------|---|----------------------------------|-----------------------|------|----------------------------|
| Signature Sequences, | | | Transmembrane domains: S77-V94, L276-W298, Y330-R350, L947- I971, Q991-I1009 | E1-E2 ATPase domains: E381-V403, O530-A562, Y633-G685, R788- | | E1-E2 ATPases phosphorylation site | ELO0154: G134-L151, V386-F404, D650- | E1-E2 ATPases phosphorylation site: | P-type cation-transporting ATPase | rfe 001 | I831 | ATPASE HYDROLASE TRANSMEMBRANE | PHOSPHORYLATION ATPRINDING PROTEIN PROBABLE CALCIUMTRANSPORTING CALCIUM | TRANSPORT PD004657: S846-P1093 | FIC1 PROTEIN PD180313: H1039-W1165 | do ATPASE; CALCIUM; TRANSPORTING; | DM02405 P32660 318-1225: W128-F418, E466-N910 | ATPase E1-E2 motif: D392-T398 | Transmembrane domain: | 4 | |
| Potential | Glycosyla- | tion Sites | N579 | | | | | | | | | | | | | | | | N126 N197 | | |
| Potential | Phosphorylation | 2 | S153 S259 S268 S391 S413 S452 S493 S545 S573 | S631 S739 | S117 | S1164 S1124 S1143 S1168 T267 | T370 T37 | T705 T980 | w | Y489 Y607 | | | | | | | | | S149 S175 S344 | S42, | T157 T355 T356 T366 T41 |
| Amino | | 20 | 1190 | | | | | | | | | | | | | | | | 467 | | |
| Incyte | lypeptide | | 7473053CD1 | | | | | | | | | | | | | | | | 7473347CD1 | | |
| SEQ | A | NO: | 4 | | | | | | | | | | | | | | | | ဌ | | |

Table 3 (cont.)

| 0 | Potential Phogshowy] ation | Potential | | Analytical |
|------|--|--------------------------|--|---|
| ques | rnospnorylation Sites | Glycosyla- tion Sites | | Methods and Databases |
| | | | | HMMER_PFAM |
| | | | Neurotransmitter-gated ion channels | BLIMPS_BLOCKS |
| | | | signature BL00236: V85-P122, I139-H148, D169- | |
| | | | Y207, Y254-A295 | |
| | | | Neurotransmitter-gated ion-channels | PROFILESCAN |
| | | | signature: L164-H218 | |
| | | | Neurotransmitter-gated ion-channels | BLIMPS_PRINTS |
| | | | signature | |
| | | | PR00252: T105-F121, K138-S149, C184- | |
| | | | 3198, S261-P273 | |
| | | | Samma-aminobutyric acid A (GABAA) | BLIMPS_PRINTS |
| | | | receptor signature | |
| | | | PR00253: F270-W290, V296-V317, V330- | |
| | | | V351, Y446-Y466 | |
| | | | CHANNEL IONIC TRANSMEMBRANE GLYCOPROTEIN | BLAST_PRODOM |
| | | | POSTSYNAPTIC MEMBRANE RECEPTOR PRECURSOR | |
| | | | SIGNAL PROTEIN | |
| | | | PD000153: E62-S427 | |
| | | | | BLAST_DOMO |
| | - | | DM00560 P50573 34-464: S37-V467 | |
| | | | | MOTIFS |
| | | <u></u> | notif: | |
| | | | 2012-120 | |
| | Incyte Amino Polypeptide Acid Residues | gnes | Potential Posphorylation Glycosyla- Sites tion Sites | Potential Signature Sequences, Phosphorylation Glycosyla- Phosphorylation tion Sites Neurotransmitter-gated ion-channel Comain: P58-Q362, H441-W463 Neurotransmitter-gated ion channels Signature Signature: L164-H218 Neurotransmitter-gated ion-channels Signature: Signature: Signature PR00252: T105-F121, K138-S149, C184-C198, S261-P273 Gamma-aminobutyric acid A (GABAA) Receptor signature PR00253: F270-W290, V296-V317, V330-V351, Y446-Y466 CHANNEL IONIC TRANSMEMBRANE GLYCOPROTEIN POSTSYNAPTIC MEMBRANE GLYCOPROTEIN POSTSYNAPTIC MEMBRANE RECEPTOR PRECURSOR SIGNAL PROTEIN PROUDSOR SIGNAL SATED ION-CHANNELS DM00560 P50573 34-464: S37-V467 Neurotransmitter-gated ion channels Neurotransmitter-gated ion channels Neurotransmitter-gated ion channels |

| Analytical | Methods and | Databases | HMMER | HMMER_PFAM | | HMMER_PFAM | | BLAST_PRODOM | BLAST_PRODOM | BLAST_DOMO | | BLAST_DOMO | BLAST_DOMO | SPSCAN | HMMER | HMMER_PFAM | PROFILESCAN | BLIMPS_PRINTS | | BLIMPS_PRINTS | |
|----------------------|--------------------|-----------|----------------------------------|------------|-------------------------|------------|------|--|---|--|---|---|--|---------------------------|--|--|---|-----------------------------|------------|-------------------------|---|
| Signature Sequences, | Domains and Motifs | | Transmembrane domain: V551-Y571 | embi | gated ion channel: | 3 | - 1 | POTASSIUM CHANNEL IONIC CHANNEL PD104127: S852-Y1028 | POTASSIUM CHANNEL IONIC CHANNEL PD104126: A1076-K1196 | CAMP RECEPTOR PROTEIN CYCLIC NUCLEOTIDE- | BINDING DOMAIN DM01165 138465 562-948: H564-A914 | do POTASSIUM; CHANNEL; KST1; AKT1; DM02383 138465 353-560: S353-A563 | do CHANNEL; POTASSIUM; EAG; DM05484 138465 1-351; M1-P351 | Signal peptide: M1-A35 | Transmembrane domains: C79-G96, M171-L188, Y322-V342, F448- I466 | Sugar (and other) transporter domain: A26-F481 | Sugar transport proteins signatures: A119-I185, V323-S379 | Sugar transporter signature | # 192 | e transporter signature | PR00172: L284-Y305, Q321-V342, L352- Q372, L383-T406, A416-F434, Y446-L466 |
| Potential | Glycosyla- | | | N600 N661 | N736 N881 N905 N1139 | | | | | | | | | N41 N57 | | | | | | | |
| Potential | Phosphorylation | ສ | S174 S187 S209 S211 S239 S269 | S275 S317 | 8354 8514 8609 8639 | S869 S879 | 2896 | S906 S922 S923 S939 S940 S963 | S974 S985 S1020 S1091 S1170 | ທ ້ | T582 | H H , | T1027 T1134 T1099 Y248 Y446 | | T235 T59 1 | | | | | | • |
| Amino | | S | 1196 | | | <u> </u> | | | | | | - | | 512 | | | | | | | |
| Incyte | lypeptide | | 7474240CD1 | | | - | | | | | | | | 7475338CD1 | | | | | | | |
| SEQ | a | NO: | ဖ | | | | | | | | | | | - | | | | | .—.· | | |

Table 3 (cont.)

| ~ | - | - | | | _ | - | | | | | | | | | _ | | | - | | |
|--|------------|---|--------------------------------------|---|---|--------------------------------------|------------------------------|---|----------------------------|------------------------------------|--|---------------------------------|--|---|---------------------------------|-----------------------------------|--|--|--|---|
| Analytical Methods and | Databases | BLAST_DOMO | MOTIFS | MOTIFS | HMMER | HMMER PFAM | | BLAST_PRODOM | | HMMER | HMMER_PFAM | | HMMER_PFAM | BLAST_PRODOM | BLAST PRODOM | | | BLAST_DOMO | | BLAST_DOMO |
| Signature Sequences, Domains and Motifs | L | SUGAR TRANSPORT PROTEINS DM00135 P22732 132-466: R138-T473 | Sugar transporter 1 motif: S338-A353 | Sugar transporter 2 motif: V140-R165 | Transmembrane domains: I242-F269, Y289-P308, I322-Y342 | Transmembrane amino acid transporter | protein domain: A102-G543 | ACID AMINO PROTEIN TRANSPORTER PERMEASE TRANSMEMBRANE INTERGENIC REGION PUTATIVE | PROLINE PD001875: W80-L380 | Transmembrane domain: L300-N318 | Transmembrane region cyclic nucleotide | gated ion channel: Y341-1580 | Cyclic nucleotide-binding domain: V608-A699 | POTASSIUM CHANNEL IONIC CHANNEL PD118772: E702-S955 | CHANNEL PROTEIN IONIC POTASSIUM | NONPHOTOTROPIC HYPOCOTYL PUTATIVE | SUBUNIT REPEAT EAG PD009483: M1-L86 | CAMP RECEPTOR PROTEIN CYCLIC NUCLEOTIDE-BLAST_DOMO | BINDING DOMAIN DM01165 138465 562-948; H413-F738, | do POTASSIUM; CHANNEL; KST1; AKT1; DM02383 138465 353-560: T201-A412 |
| | tion Sites | | | | N141 N205 N214 N256 | N562 | 9 LN | | | N218 N449 N510 N742 | | | | | | | | | | |
| Potential Phosphorylation | Sites | | | | S143 S365 S4 S456 S46 S51 S55 | T430 Y45 | | | | S140 S145 S26 S283 | \$45 | S670 S75 | S788 S864 S872 S879 S897 S929 | T170 T30 | T363 T377 T486 | T522 T678 | | | | |
| Amino Acid | Residues | | | | 268 | | | | | 958 | | | | - | | | | | | |
| Incyte Polypeptide | fl | | | | 7476747CD1 | | | | | 7477898CD1 | | | | | | | - | | | |
| ar Seg | NO: | _ | | | ∞ | | | | | 6 | | | | | | | | | | |

| Analytical | Methods and Databases | HMMER | HMMER_PFAM | BLAST_PRODOM | | HMMER | | BLAST_PRODOM | | HMMER | | | HMMER_PFAM | BLIMPS_BLOCKS | PROFILESCAN | | PROTEIN BLAST_PRODOM | | | BLAST_PRODOM | | BLAST_DOMO |
|----------------------|--------------------------|---|--------------------------------------|--|----|--|-------------|------------------------------|---------------------|------------------------|-------------------------------------|-----------|--|---|-------------|-------------------------|----------------------|---|--------------------|---------------------------------|--------------------------------|---|
| Signature Sequences, | DOMMAINS AND MOCILS | Transmembrane domains: A370-L388, I419-F437, V486-M503 | TASK K+ channel domain: M250-D646 | TWIKI RELATED POTASSIUM CHANNEL, SIBFAMILY K. MEMBER 2 TREKI K+ CHANNEL | 걸성 | Transmembrane domains: F62-Y87, F139-F163, F212-L230, I293- | 1312 | VANILLOID RECEPTOR SUBTYPE 1 | PD137334: C348-K470 | Transmembrane domains: | D10-F28, F81-Y104, F278-M297, L439- | 1306-N360 | Sodium:solute symporter family domain: F41-G445 | Sodium:solute symporter signature BL00456: T154-G208 | 15 | signature: N151-T198 | T PERMEASE | SODIUM SYMPORT PROLINE COTRANSPORTER SYMPOPHED CLYCOBROTHEN | PD000991: F41-C304 | SYMPORTER SODIUM IODIDE THYROID | PD024705: I446-L489, S490-G575 | SODIUM:SOLUTE SYMPORTER FAMILY DM00745 P31636 24-561: D10-N219, G220- Y459 |
| Potential | clycosyla- tion Sites | | | | | N236 N256 N321 N380 | | | | N25 | N480 N574 | | | | | | | | | | | |
| ntial | S | S283 S303 S512 S545 | S666 S718 T19 T223 | T51 | | S142 S245 S355 S408 | S415 | T15 7 | | S265 S313 | S490 S550 | 2 6 | | T567 T70 | - | | | | | | | |
| 0 | Residues (| 724 | <u> </u> | <u></u> | | 470 | <u> </u> | 92 | <u> </u> | 618 | <u> </u> | | . · <u>. <u>.</u> ·</u> | <u> </u> | | | | | | | | |
| | 7 | 7472728CD1 | | | | 7474322CD1 | | | | 5455621CD1 | | • | | | | | | | | | | |
| SEQ | NO T | 10 | | | | 11 | | | | 12 | | | | O rași șa | | | | | | | | |

| Analytical | Methods and | Databases | HMMER | domain: HMMER_PFAM | BLIMPS_PRINTS | BLIMPS_PRINTS | BLIMPS_PRODOM | BLAST_PRODOM | BLAST_PRODOM | BLAST_DOMO | HMMER . | HMMER_PFAM | BLIMPS_BLOCKS | | PROFILESCAN |
|----------------------|--------------------|------------|---|--|---|---|---------------|---|--|---|--|---|-------------------------|--------------------------|--|
| Signature Sequences, | Domains and Motifs | | Transmembrane domains: V22-F41, L159-M181, I391-A407 | Sodium/hydrogen exchanger family domain: L25-V491 | Na+/H+ exchanger isoform 6 signature PR01088: Y14-I38, W39-V57, Y58-V84, Q119-E132, A269-M288, T480-Q506, K515- | No. 2018 - 100 / P. 200 - E. 2018 No. 4 / H+ exchanger signature PR01084: I133-F144, G147-S161, I162-M170 C208-M218 | TRAIN 13 A | WA+/H+ PROTEIN TRANSMEMBRANE TRANSPORT ANTIPORTER SYMPORT SODIUM EXCHANGER GLYCOPROTEIN SODIUM/HYDROGEN PD000631: G20-G63, E132-R490 | SODIUMHYDROGEN EXCHANGER 6 MYELOBLAST KIAA0267 PD177855: G478-Y591 | do BETA; EXCHANGER; NA; DM02572 P48764 10-734: L124-L541 | Transmembrane domains: Y231-Y251, L415-L434, I933-I959, F966- L985, I1002-F1020, N1104-M1122 | E1-E2 ATPase domains: V274-V365, G490-D506, Q672-A785, L851- S899 | es phosphorylation site | N724-M764, V878-S901, A9 | EI-E2 ATPases phosphorylation site: I478-E526 |
| Potential | Glycosyla- | tion Sites | N352 N516 N96 | | | | | | | | N150 N23 N300 N312 N318 N704 | N1045 N1053 N1059 | N1073 N1247 | | |
| Potential | Phosphorylation | Sites | | T551 T73 T79 Y14 | | | | | | | 1 | S98 9 T353 T502 | T576 T74 T1212 T1061 |) | |
| Amino | Acid | Residues | 631 | | | | | | | | 1256 | | | | |
| Incyte | Polypeptide | ID | 7477248CD1 | | | | | | | | 2944004CD1 | | | | |
| SEQ | QI | NO: | 13 | | | | | | | | 14 | | | | |

| Analytical Methods and Databases | BLIMPS_PRINTS | BLAST_PRODOM | BLAST_DOMO | MOTIFS | SPSCAN | HMMER | signature: HMMER_PFAM | PROFILESCAN | BLIMPS_PRINTS | BLIMPS_PRINTS | BLAST_DOMO | MOTIFS |
|--|---|--|------------|----------------------------------|----------------------------------|---|---|---|--|---|---|---|
| Signature Sequences, Domains and Motifs | P-type cation-transporting ATPase superfamily signature PR00119: N318-T332, C496-L510, A740- D750, C881-L900 | ATPASE PROBABLE CALCIUMTRANSPORTING PROTEIN HYDROLASE CALCIUM TRANSPORT TRANSMEMBRANE PHOSPHORYLATION MAGNESIUM PD090368: Q995-Y1094, D1064-L1114 | | E1-E2 ATPase motif: D498-T504 | Signal peptide: M1-G27 | Transmembrane domains: M163-L181, T371-G389, M418-L440 | Sugar (and other) transporter signature: L18-L474 | Sugar transport proteins signature: A112-V178 | Sugar transporter signature PR00171: T28-I38, M128-M147, M376- L397, T399-C411 | Glucose transporter signature PR00172: Q314-1335, M376-T399, A409- L427 | SUGAR TRANSPORT PROTEINS DM00135 P22732 132-466: R131-T466 | Sugar transporter 2 motif: L133-R158 |
| Potential Glycosyla- tion Sites | | | | | N292 N34 N50 | | | | | | | |
| Potential Phosphorylation Sites | | | | | S100 S118 S215 S285 T466 T487 | | | | | | | |
| o | | | | | 499 | | | | | | | |
| Incyte Amin Polypeptide Acid ID | | | | | 3046849CD1 | | | | | | | |
| SEQ ID NO: | 년 디 | | | | 15 | | | | | | | · |

| Incyte Amino | _ | Potential | Potential | Signature Sequences, | Analytical |
|--------------|----------|-----------------|------------|---|---------------|
| eptide | | Phosphorylation | Glycosyla- | Glycosyla- Domains and Motifs | Methods and |
| | Residues | Sites | tion Sites | | Databases |
| 4538363CD1 | 965 | S17 S290 S39 S5 | N239 N386 | Transmembrane domains: | HMMER |
| | | T119 T211 | N4 N545 | S73-W95, I185-I212, L356-A376, L410- | |
| | | | 96N | V430, F473-F491, Y513-L533 | |
| | | | | Sodium:solute symporter family domain: Y50-G479 | HMMER_PFAM |
| | | | | Sodium: solute symporter signature | BLIMPS BLOCKS |
| | | | | BL00456: Y27-G81, A103-R132, L165- | İ |
| | | | | G219, P452-G461 | |
| | | | | Sodium: solute symporter family | PROFILESCAN |
| | | | | signatures: | |
| | | | | H162-I209, V412-D502 | |
| | | | | TRANSMEMBRANE TRANSPORT PERMEASE PROTEIN BLAST_PRODOM | SLAST_PRODOM |
| | | | | SODIUM SYMPORT PROLINE COTRANSPORTER | |
| | | | | SYMPORTER GLYCOPROTEIN | |
| | | | | PD000991: Y50-G479 | |
| | | | | NA+/GLUCOSE COTRANSPORTERRELATED PROTEIN BLAST_PRODOM | SLAST_PRODOM |
| | | | | PD134393: L551-A596 | |
| | | | | NA+/GLUCOSE COTRANSPORTERRELATED PROTEIN BLAST_PRODOM | SLAST_PRODOM |
| | | | | PD166538: M1-G49 | |
| | | | | SODIUM: SOLUTE SYMPORTER FAMILY | BLAST_DOMO |
| | | | | DM00745 P13866 24-561: S17-W548 | |
| | | | | Na solute symporter 2 motif: | MOTIFS |
| | | | | G461-V481 | |

| Analytical Methods and Databases | HMMER | HMMER_PFAM | BLIMPS_BLOCKS | | PROFILESCAN | BLIMPS_PRINTS | | BLAST_PRODOM | | | | BLAST_DOMO | MOTIFS | | HMMER | HMMER_PFAM | 1 | BLIMPS_PRINTS | |
|--|---|--|----------------------------------|--|---|-----------------------------------|--|--------------------------------|------------------------------------|--------------------------------------|-----------------------------------|--|---------------------|-----------|------------------------|---------------|----------------|-----------------------------|---|
| Signature Sequences, Domains and Motifs | Transmembrane domains: V299-Y316, F1004-L1022, I1030-W1049, A1075-L1092 | E1-E2 ATPase domains: E403-E425 I550-C698 | | BL00154: G149-F166, V408-F426, D663- L703 | E1-E2 ATPases phosphorylation site: L395-C442 | P-type cation-transporting ATPase | superfamily signature PR00119: F412-F426, A679-D689 | ATPASE HYDROLASE TRANSMEMBRANE | PHOSPHORYLATION ATPBINDING PROTEIN | PROBABLE CALCIUMTRANSPORTING CALCIUM | Transport PD004657: A857-V1108 | do ATPASE; CALCIUM; TRANSPORTING; DM02405 209891 206-1107: T105-Y436, | E1-E2 ATPase motif: | D414-T420 | Transmembrane domains: | rt protein do | | Potassium channel signature | K307, F310-V330, F352-S378, E381-E404, F421-M443, G450-F476 |
| Potential Glycosyla- tion Sites | | | | | | | | | | | | | | | N259 N266 | | | | |
| Potential Phosphorylation Sites | S169 S188 S287 S335 S507 S508 | 555 593 | T255 T259 T269 T333 T380 T413 | T659 T715 | T1103 T1017 T1105 Y885 Y1026 | | | | | | | | | | S336 | S564 S86 | T120 T146 T155 | | T57 |
| Amino Acid Residues | | | | | | - | | | | | - | | | | 638 | | | | |
| Incyte Polypeptide I | 427460CD1 | | | | | | | | | | | | | | 7474127CD1 | | | | |
| SEQ ID NO: | 17 | | | | | | | | | | | | | | 18 | | _ | | |

Table 3 (cont.)

| SEQ | Incyte | Amino | Potential | Potential | Signature Sequences, | Analytical |
|-------------|------------|----------|-----------------|-------------------|--|---------------|
| QI. | lypeptide | Acid | Phosphorylation | Glycosyla- | Domains and Motifs | Methods and |
| 02 | IΩ | Residues | Sites | tion Sites | | Databases |
| 18 | | | | | VOLTAGEGATED POTASSIUM CHANNEL PROTEIN KV3.2 KSHIIIA IONIC TRANSMEMBRANE ION | BLAST_PRODOM |
| | | | | | TRANSPORT GLICOPROTEIN MULTIGENE FAMILY ALTERNATIVE SPLICING PHOSPHORYLATION PD085814: K495-S538 | |
| | | | | | do CHANNEL; POTASSIUM; CDRK; FORM; DM00436 P22462 189-350: R189-R351 | BLAST_DOMO |
| | | | | | do CHANNEL; POTASSIUM; CDRK; SHAW; DM00490 P22462 34-151: L34-C152 | BLAST_DOMO |
| 13 | 7476949CD1 | 681 | S421 S56 | N113 N251 | Transmembrane domains: | HMMER |
| | | | | N256 N403 N603 | I38-157, S90-W112, I150-I167, L188- M207, L373-A393, V432-I448, Y530-L550 | |
| | | | | | Sodium:solute symporter family domain: Y67-G496 | HMMER_PFAM |
| | | | | | Sodium:solute symporter signature BL00456: Y44-G98, A120-R149, L182- G236, P469-A478 | BLIMPS_BLOCKS |
| | | | | | Sodium:solute symporter family | PROFILESCAN |
| | | | | | signatures: Q179-V226, D458-D519 | |
| | | | | | TRANSMEMBRANE TRANSPORT PERMEASE PROTEIN BLAST_PRODOM SODIUM SYMPORT PROLINE COTRANSPORTER | BLAST_PRODOM |
| | | | | | SYMPORTER GLYCOPROTEIN PD000991: Y67-G496 | |
| | | | | | SODIUM: SOLUTE SYMPORTER FAMILY DM00745 P13866 24-561: H34-W565 | BLAST_DOMO |
| | | | | | Na solute symporter 1 motif: G183-A208 | MOTIFS |

| Analytical | Methods and | Databases | HMMER | HMMER_PFAM | BLIMPS_BLOCKS | | BLIMPS_PRINTS | • | BLIMPS_PRINTS | BLAST_PRODOM | | | טאטת שפע זמ | | | MOTIFS | SPSCAN | | HMMER | HMMER_PFAM | BLIMPS_PRINTS |
|----------------------|--------------------|------------|---|---|------------------------------------|--|-----------------------------------|--|---|--------------------------------|--|-----------|-------------|---------|------|----------------------------------|-----------------|----------|---|---|---|
| Signature Sequences, | Domains and Motifs | | Transmembrane domains: F289-L307, F935-L953, W967-V996, F1008-D1028 | E1-E2 ATPase domains: T340-Q352, H502-V648 | E1-E2 ATPases phosphorylation site | signature BL00154: G143-L160, V335-F353, K529- C539, D616-H656 | P-type cation-transporting ATPase | superfamily signature PR00119: F339-F353, A632-D642 | H+-transporting ATPase signatur PR00120: T547-A565 | ATPASE HYDROLASE TRANSMEMBRANE | PHOSPHORYLATION ATPBINDING PROTEIN PROBABLE CALCIUMTRANSPORTING CALCIUM | TRANSPORT | 17 | <u></u> | 1851 | E1-E2 ATPase motif: D341-T347 | Signal peptide: | M1-A26 | Transmembrane domains: I155-Y178, I271-T292, | Sodium/hydrogen exchanger family domain: V73-K482 | Na+/H+ exchanger signature PR01084: I158-A166, G200-A210, I129- I140, G143-S157 |
| Potential | Glycosyla- | tion Sites | N331 N383 N395 N411 N720 N932 | | | | | | | | | | | | | | N297 N31 | N342 N35 | | | |
| Potential | Phosphorylation | ຜ | S115 S163 S276 S280 S332 S333 S404 S454 S46 | S462 S671 | | T345 T T570 T T840 T | T1034 T1036 Y322 | | | | | | | | | | S299 S360 | S488 | S58 S585 S591 S620 S638 S679 | | |
| | | Residues | 1096 | | | | | | | | | | | | | : | 707 | | | | |
| 1 | lypeptide | ID | 7477249CD1 | | | | | | | | | | | | | | 7477720CD1 | | | | |
| SEQ | <u>n</u> | NO: | 20 | | | | | | | | | | | | | - | 21 | _ | | | |

Table 3 (cont.)

| Analytical | Methods and | Databases | BLIMPS_PRINTS | | BLIMPS_PRODOM | | | BIAST PRODOM | | | BLAST_DOMO | | HMMER | WARD DRAW | TIMILE K - F F AM | BLAST_PRODOM | MOTIFS | HMMER | | | HMMER_PFAM | | BLAST_DOMO | | BLAST_DOMO |
|----------------------|--------------------|--------------|-----------------------------------|--|--------------------------|-------------------------------------|--|--------------|-------------------------------------|---|------------|-------------------------------------|--|------------------|--------------------------------|--|---|-----------------------|-------------------------------------|----------------------------|--------------------------------|----------|--------------------------|------------------------------------|--|
| Signature Sequences, | Domains and Motifs | | Na+/H+ exchanger isoform 2 (NHE2) | signature PR01086: F115-S128, K616-1627 | + TRANSPORT EXCHANGER NA | PD01672: A83-I113, I129-L177, Y178- | 1322-M355, S359-F405, Y406-F452, I489- 1231 T532-G562 P593-P640 | + PROTEIN TH | ANTIPORTER SYMPORT SODIUM EXCHANGER | GLYCOPROTEIN SODIUM/HYDROGEN PD000631: 177-A438 | 1 | DM02572 P26434 14-716: L15-L687 | Transmembrane domains: F493-F512. M554-M570 | Ankirin repeate. | L78-E108, A116-T148, F162-S194 | VANILLOID RECEPTOR SUBTYPE 1 PD101189: F115-L220 | ATP/GTP binding site (P-loop): A412-T419 | transmembrane domain: | I48-V71, V86-F104, Y172-I199, I199- | V217, F384-F402, V452-C472 | Sugar (and other) transporter: | I48-K492 | SUGAR TRANSPORT PROTEINS | DM00032 P30638 80-152:R45-K115 | VESICLE; SYNAPTIC; SV2; FORM DM08835 S34961 180-344:1119-N249 |
| Potentia1 | | tion Sites | | | | | | | | | | | N208 N358 N717 | | | | | N229 N249 | | | | | | | |
| Potential | Phosphorylation | Sites | | | | | | | | | | | S144 S155 S291 S299 | 2654 5664 | 2697 | T110 T138 T281 T379 T447 T532 | T539 | S18 S225 | S314 S373 T323 | T33 T351 T426 | | | | | |
| 0 | | Residues | | | | | | | | | | - | 729 | | | | | 492 | | | | | | | |
| Incyte | lypeptide | OI. | | | | | | | | | | | 7477852CD1 | | | | | 1471717CD1 | | | | | | | |
| SEQ | A : | <u>S</u> | 21 | | | | | | | | - | | 22 | | | | | 23 | | - | | | | | |

Table 3 (cont.)

| SEQ | Incyte | Amino | Potential | Potential | Signature Sequences, | Analytical |
|-----|------------------|----------|------------------|------------|---|---------------|
| ID | Polypeptide Acid | Acid | Phosphorylation | Glycosyla- | Glycosyla- Domains and Motifs | Methods and |
| NO: | Ωī | Residues | Sites | tion Sites | | Databases |
| 24 | 3874406CD1 | 1494 | S30 S50 S134 | N109 N130 | transmembrane domain: | HMMER |
| | | | S230 S368 S549 | N313 N421 | L204-F221, T272-L290, L735-Y753, F896- | |
| | | | 8638 8669 8686 | N453 N71 | S914, V941-1959, L975-R998, F1019-V1039 | |
| | | | S696 S792 S800 | N788 N817 | ABC transporter: | HMMER_PFAM |
| | | | S831 S912 S1004 | N84 N867 | G384-G566 G1190-G1366 | |
| - | | | S1070 S1146 | N91 N1182 | ABC transporters family proteins | BLIMPS_BLOCKS |
| | | | S1172 S1206 | | BL00211: I389-L400, L492-D523 | |
| | | | S1365 T111 T435 | | ABC transporters family signature: | PROFILESCAN |
| | | | T449 T501 T520 | | V472-D523 | |
| | | | T632 T649 T657 | | ABC TRANSPORTERS FAMILY | BLAST_DOMO |
| | | | T729 T845 T1049 | | DM00008 P41233 839-1045:I355-N565, | |
| نب | | | <# | | K1177-M1363 | |
| | | | T1247 T1295 | | DM00008 P34358 611-816:I355-N565, | |
| | | | T1318 T1339 | | A1179-M1363 | |
| | | | T1422 T1482 Y824 | | DM00008 P41233 1851-2058:K1173-S1365, | |
| | | | | | I355-N565 | |
| - | | | | | DM00008 P23703 41-246: E1162-G1366, | |
| | | | | | L377-G566 | |
| | | | | | ATP/GTP-binding site motif A (P-loop): | MOTIFS |
| - | | | | | G391-S398, G1197-2004 | |

| SEQ | Incyte | Amino | Potential | Potential | Signature Sequences, | Analytical |
|-----|-------------|----------|-----------------|------------|---|---------------|
| 요 | Polypeptide | Acid | Phosphorylation | syla- | Domains and Motifs | Methods and |
| NO: | ID | Residues | Sites | tion Sites | | Databases |
| 25 | 4599654CD1 | 774 | S356 S40 | N291 N416 | transmembrane domain: | HMMER |
| | | | S505 S552 S559 | | Y95-F118, T203-L219, L327-L353 | |
| | | | | | Transmembrane region cyclic Nucleotide | HMMER_PFAM |
| | | | S734 S736 T203 | | : | |
| | | | T418 T668 T764 | | Y168-1414 | |
| | | | V490 | | Cyclic nucleotide-binding domain: | HMMER_PFAM |
| | | | | | K443-M531 | |
| | | | | | Cyclic nucleotide-binding domain | BLIMPS_BLOCKS |
| ~ | | | | | proteins | |
| | | | | | BL00888: G452-V475, G488-L497 | |
| | | | | | cAMP-dependent protein kinase signature | BLIMPS_PRINTS |
| | | | | | PR00103: F449-R463, S489-T498 | |
| | | | | | HYPERPOLARIZATIONACTIVATED CATION | BLAST_PRODOM |
| | | | | | CHANNEL, HAC3 | |
| | | | | | PD180735: T538-M774 | |
| | | | | | CHANNEL IONIC POTASSIUM K+ SUBUNIT | BLAST_PRODOM |
| | | | | | HYPERPOLARIZATIONACTIVATED PROTEIN | |
| | | | | | PUTATIVE EAG LONG | |
| | | | | | PD001039: E74-R167 | |
| | | | - | | CAMP RECEPTOR PROTEIN CYCLIC NUCLEOTIDE- BLAST_DOMO | BLAST_DOMO |
| | | | | | BINDING DOMAIN | |
| | | | | | DM01165 A55251 333-706:H263-P561 | |
| | | | | | DM01165 P29973 311-684: H263-P561 | |
| _ | | | | | DM01165 Q03041 286-658:H263-G548 | |
| | | | | | DM01165 S52072 262-635:H263-Q595 | |

Table 3 (cont.)

| Analytical | Methods and | Databases | HMMER |), W483- | | HMMER_PFAM | BLIMPS_BLOCKS | | BLIMPS_PRINTS | .486~ | | BLIMPS_PRINTS | R519- | | MOTIFS | | res PROFILESCAN |)1 | 35 | BLAST_DOMO | 17, | |)5, | | 38, | | 17. |
|----------------------|--------------------|------------|-----------------------|---------------------------------------|----------------------------|--------------------------------|--------------------------|------------------------------|-----------------------------|-------------------------------------|-----------------|-------------------------------|--------------------------------|-----------------|--------------------|---------------------|-------------------------------------|----------------------------------|----------------------------------|------------|---------------------------------------|------------|---------------------------------------|-----------|---------------------------------------|----------------------|---------------------------------------|
| Signature Sequences, | Domains and Motifs | | transmembrane domain: | V124-I142, A168-M190, A371-V390, W483 | I511, S526-I543, F552-V570 | Sugar (and other) transporter: | Sugar transport proteins | BL00216: L174-S223, G92-S103 | Sugar transporter signature | PR00171: G92-I102, V175-I194, L486- | V507, S509-F521 | Glucose transporter signature | PR00172: V343-V364, L486-S509, | L537, W550-V570 | Sugar_Transport_1: | G138-G153 A360-A375 | Sugar transport proteins signatures | sugar_transport_1.prf: L344-S401 | sugar_transport_2.prf: A160-A225 | | DM00135 S25015 122-478:A160-D417, | L480-K574, | DM00135 P09830 101-452:G161-V405, | L481-K574 | DM00135 Q01440 101-433:R178-G388, | R178-G388, L486-G575 | DM00135 P15729 242-463: A485-S577 |
| Potential S | | tion Sites | N407 N599 t | | H | <u> </u> | S | | S | | Δ | В | | ᆸ | ß | | ß | | | ន | | <u> </u> | | <u> </u> | | <u>æ</u> | |
| Potential | horylation | | S116 S210 S290 | S577 | T267 T432 T443 | T591 | | | | | | | | | | | - | | | | | | | | | | |
| Amino | Acid | Residues | 614 | | | | | | | | | | | | | | | | | | | | | | | | |
| Incyte | Polypeptide | ជ | 5047435CD1 | | | | | | | | | | | | | | | | | | | | | | | | |
| SEQ | ΩI | No: | 26 | | | | | | | | | | | | | | - | | | | | | | | | | |

Table 3 (cont.)

| SEQ | Incyte | Amino | Potential | Potential | Signature Sequences, | Analytical |
|------------|------------|----------|------------------|------------|--|---------------|
| <u>n</u> : | lypeptide | Acid | Phosphorylation | | Domains and Motifs | Methods and |
| 02 | Ω | Residues | S | tion Sites | | Databases |
| 27 | 7475603CD1 | 2180 | S 216 | N112 N132 | transmembrane domain: | HMMER |
| | | | S260 S409 S419 | N346 N374 | F630-L648, L664-L680, V1570-V1590, | |
| | | | S842 S983 S1008 | N1100 | M1622-Q1641 | |
| | | | | N1415 | ABC transporter: | HMMER_PFAM |
| | | | | N1420 | G1854-G2035 G868-G1048 | |
| | | | S1349 S1353 | N1491 | ABC transporters family | BLIMPS_BLOCKS |
| | | | S1462 S1469 | N1552 | BL00211: F873-T884, L974-D1005 | |
| · · · · · | | | ₩. | N1695 | ABC transporters family signature: | PROFILESCAN |
| | | | -1 | N1831 | A1940-D1991, D955-D1005 | |
| | | | ന ദ | | Abc_Transporter: | MOTIFS |
| | | | _ | | L974-F988 | |
| | | | | | ATP/GTP-binding site motif A (P-loop): | MOTIFS |
| | | _ | T. FTQ.T. | | G875-T882, G1861-T1868 | |
| | | | Ξ, | | ATPBINDING TRANSPORTER CASSETTE ABC | BLAST_PRODOM |
| | | | . | | TRANSPORT PROTEIN GLYCOPROTEIN | |
| | | | - (| | TRANSMEMBRANE RIM ABCR | |
| | | | o 0 | | PD005939: L1563-N1740 | |
| | | | n (| | ATPBINDING TRANSPORTER CASSETTE ABC | BLAST_PRODOM |
| | | | - | | GLYCOPROTEIN TRANSMEMBRANE TRANSPORT | |
| _ | | | ω i | | ABCR RIM | |
| | | | TZ1Z5 Y656 Y1448 | | PD010118: R238-R514, L95-R243 | |
| | | | | | ATPBINDING TRANSPORTER CASSETTE ABC | BLAST_PRODOM |
| | | | | | GLYCOPROTEIN TRANSMEMBRANE TRANSPORT | |
| | | | | | ABCR RIM SIMILARITY | |
| | | | | | PD008845: P1307-E1560 | |
| | | | | | ATPBINDING TRANSPORTER CASSETTE ABC | BLAST_PRODOM |
| | | | | | GLYCOPROTEIN TRANSMEMBRANE TRANSPORT RIM | |
| | | | | | | |
| | | | | | PD006867: L540-S685, D515-Q541 | |

| Analytical Methods and Databases | BLAST_DOMO | нммек | HMMER_PFAM | BLIMPS_PRINTS | IDENTICAL BLAST_PRODOM T1720 | BLAST_PRODOM | BLAST_PRODOM | BLAST_PRODOM |
|--|---|---|--|---|--|--|--|---|
| Signature Sequences, Domains and Motifs | ABC TRANSPORTERS FAMILY DM00008 P41233 839-1045:V841-A1046, L1829-M2032 DM00008 P41233 1851-2058:V1826-N2034, V841-V1045 DM00008 P34358 1441-1640:L1827-M2032, V843-V1045 | transmembrane domain: M1244-A1262, V1319-F1336, I1338-F1357, A1423-I1446, W107-V126, V181-M199, S298- I321, L509-V531, V575-I598, Y879-M904, I1017-F1034, I1134-V1152 | <pre>Ion transport protein ion_trans: W32-I321 M380-I598 L884-V1155 I1206- I1446</pre> | Calcium channel signature PR00167: D535-D561 | C11D2.5 NEARLY CTED 47-S1637, E1714- | C11D2.6 PROTEIN PD178227: L1241-R1368, I1206-F1292 F585-E606 | C11D2.6 PROTEIN SIMILARITY ALONG ENTIRE GENE CALCIUM CHANNEL ALPHA PROTEINS PD041964: L599-V885, | CHANNEL CALCIUM IONIC SUBUNIT VOLTAGE GATED SODIUM ALPHA TRANSMEMBRANE L TYPE PD000032: Y887-V1120, I33-V330, K1361-F1450, I1206-F1357, I577-I598, F1337- L1356, I1134-F1159, D1416-V1443 |
| Potential Glycosyla- tion Sites | | N210 N216 N859 N1064 N1371 N1449 | | | | | | |
| Potential Phosphorylation Sites | | 154 S687 1695 S7 1766 S773 11 S1113 S1271 | 7 7 2 | ວິ | 9 9 9 | T1561 T1570 T1645 T1694 Y419 Y702 Y832 | | |
| Amino Acid Residues | | 1737 | | | | | | |
| Incyte Polypeptide ID | | 7477845CD1 | | | | | • | |
| SEQ ID NO: | 27 | . 28 | | | | | | |

Table 3 (cont.)

| SEO | Throte | Amino | Dotontial | Dotontial | Gimatiira Gamienras | Anslytical |
|------|-------------|-------|------------------------|-----------|--|--------------|
| 10.5 | Polypeptide | | Phosphorylation | 1, | Domains and Motifs | Methods and |
| NO: | ID | dues | Sites | | | Databases |
| 28 | | | | | | BLAST_DOMO |
| | *** | | | | DM00079 A55138 (1052-1268:V1020-L1227 | |
| | | | | | DM000/9 P355000 1424-1636:W1090-P1194, | |
| | | | | | IV REPEAT | BLAST DOMO |
| | | | | | DM00277 P27732 1363-1572:F1337-L1536 | |
| 20 | 168877011 | 577 | 1000 2710 | | DMU02// F13361 1364-1393:F133/-D1330 | 00000 |
| 67 | 70007 | | T075 / 075 | INTO NTO | cransmemorane domain: | HMMEK |
| | | | S336 S404 S526 T133 | N56 | F16-T35, Y180-C200, S201-V222, M410- E429, T469-Y492, L496-L514 | |
| | | | - | | Sugar (and other) transporter: | HMMER_PFAM |
| | | | T453 T58 | | L13-Q528 | 1 |
| - | | | | | ORGANIC TRANSPORTERLIKE TRANSPORT | BLAST_PRODOM |
| | | | | | PROTEIN RENAL ANION TRANSPORTER CATIONIC | |
| | | | | | KIDNEYSPECIFIC SOLUTE | |
| | | | | | PD151320: N102-L144 | |
| 30 | 7472734CD1 | 547 | S167 S201 | N102 N39 | | HMMER |
| _ | | | | N56 N62 | I18-F32, M147-Y163, Y180-C200, S201- | |
| | | | S408 S46 S526 | | V222, M410-E429, T469-Y492, L496-L514 | |
| | | | U, | | Sugar (and other) transporter: | HMMER PFAM |
| | | | T323 T432 T453 | | L18-Q528 | |
| | | | 158 | | SUGAR TRANSPORT PROTEINS | BLAST_DOMO |
| | | | , | | DM00032 P46501 280-351:V121-K173 | |
| | | | | | ORGANIC TRANSPORTERLIKE TRANSPORT | BLAST_PRODOM |
| | | | | | PROTEIN RENAL ANION TRANSPORTER CATIONIC | |
| | | | | | KIDNEYSPECIFIC SOLUTE | - |
| | | | | | PD151320: N102-K145 | |

| Polypeptide Acid Phosphorylation Glycosyla- ID Residues Sites 7473473CD1 988 S142 S237 S24 N170 N235 S252 S322 S369 N403 N466 S502 S680 S773 N663 N830 S847 S883 S925 S943 S952 S974 S981 T127 T14 T215 T442 T478 T215 T442 T478 T521 T634 T725 T73 T832 T869 T909 T929 | Signature Sequences, | Analytical |
|---|--|-------------------|
| Residues Sites 1988 142 S237 S24 N170 S252 S322 S369 N403 S502 S680 S773 S847 S883 S925 S943 S952 S974 S981 T127 T14 T215 T442 T478 T521 T634 T725 T73 T832 T869 T909 T929 T909 T929 | nd Motifs | Methods and |
| 988 S142 S237 S24 N170 S252 S322 S369 N403 S502 S680 S773 N663 S847 S883 S925 S943 S925 S943 S952 S974 S981 T127 T14 T215 T442 T478 T73 T832 T869 T909 T929 T909 T929 | α · | Databases |
| S680 S773 N663 S883 S925 S952 S974 T127 T14 T42 T478 T634 T725 T832 T869 T929 | transmembrane domain: L342-A360 | HMMER |
| 7003 7127 7127 7127 7133 71929 71929 | ne cyclic Nucleotide G: | HMMER_PFAM |
| 1127 1127 1634 1929 1929 | | 2000 |
| T442 T634 T929 T929 | Cyclic nucleotide-binding domain: H V564-A655 | HMMEK_PFAM |
| 1 T634 T832 J 9 T929 | PA: | HMMER PFAM |
| <u>ი</u> | C92-T132 | |
| | SUBUNIT | HEAG BLAST_PRODOM |
| | LONG ELECTOCARDIOGRAPHIC OT SYNDROME PD017645: K809-D984 | |
| | NIT HYPERPO- | BLAST PRODOM |
| | PUTATIVE EAG LONG | |
| | PD001039: S179-I284 | |
| | CHANNEL K+ IONIC EAG SUBUNIT | BLAST_PRODOM |
| | TRANSMEMBRANE ION TRANSPORT VOLTAGEGATED | |
| | PD011550: N658-E737 | |
| | POTASSIUM NON | BLAST_PRODOM |
| | PHOTOTROPIC HYPOCOTYL PUTATIVE SUBUNIT | |
| 5 | REPEAT EAG | |
| 5 | PD009483: M1-E89 | |
| | CAMP RECEPTOR PROTEIN CYCLIC NUCLEOTIDE-BLAST_DOMO | SLAST_DOMO |
| | BINDING DOMAIN | |
| | I48912 | |
| | DM01165 Q02280 384-776:H361-E737 | |
| | | |
| | S974-E985 | |
| | | BLAST_DOMO |
| | DM02383 148912 164-389:V162-E314, | |

Table 3 (cont.)

| ıa | Amino | Potential | Potential | Potential Signature Seguences, | Analvtical |
|-------------------|-------|-----------------|------------|--|--------------|
| Polypeptide Acid | | Phosphorylation | Glycosyla- | | Methods and |
| Residues Sites | Ø | Sites | tion Sites | | Databases |
| 32 7477725CD1 533 | | S107 S109 S143 | N102 N216 | transmembrane domain: | HMMER |
| | | S167 S282 S345 | N56 N62 | F150-D168, L380-N401, I407-V426, L486- | |
| | | S408 S469 S60 | | F504 | |
| | | T133 T289 T323 | | Sugar (and other) transporter: | HMMER_PFAM |
| | | T336 T432 T526 | | A111-K528 | |
| | | | | ORGANIC TRANSPORTER LIKE TRANSPORT | BLAST_PRODOM |
| | | | | PROTEIN RENAL ANION TRANSPORTER CATIONIC | |
| | | | | KIDNEY SPECIFIC SOLUTE | |
| | | | | PD151320: N102-K145 | |

Table 4

| | Polynucleotide SEQ ID NO: | Incyte Polynucleotide ID | Sequence Length | Selected Fragment(s) | Sequence Fragments | 5' Position | 3' Position |
|--|------------------------------|-----------------------------|--------------------|--------------------------------|---|-------------|-------------|
| | 33 | 3474673CB1 | 1775 | 1-391, 578-786, 1024-1301 | GNFL.g7798848_00000 3_004.edit | Ţ | 1156 |
| | | | | | 6724643H1 (LUNLTMT01) | 861 | 1347 |
| ··· | | | | | 3474673H1 (LUNGNOT27) | 249 | 568 |
| | | | | | 71495515V1 | 1205 | 1775 |
| | 34 | 4588877CB1 | 1545 | 261-619, 1-193, | 71495515V1) | 975 | 1545 |
| | | | | 794-1071 | FL135171_00001 71497982V1 | 539 | 1534 |
| <u>. </u> | 35 | 7472214CB1 | 1941 | 1483-1558, 1- 413, 495-616, | GBI:g8117242_000054 _edit.8639-8803 | 1171 | 1335 |
| ĺ | | | | 732-1149 | GBI:g8117242_000054 _edit.4857-4997 | 544 | 684 |
| | | | | | GBI:g8117242_000054 .edit.10305-10463 | 1441 | 1599 |
| | | | | | 6891360H1 (BRAITDR03) | 1433 | 1905 |
| 112 | | | | | GBI:g8117242_000054 _edit.50~89 | Н | 240 |
| | | | | | GBI:98117242_000054 _edit.6950-7093 | 925 | 1068 |
| | | | | | GBI:g8117242_000054 _edit.4345-4478 | 358 | 492 |
| | | | | | 60124962D2 | 1735 | 1941 |
|] | | | | | GBI:98117242_000054 _edit.8313-8414 | 1069 | 1170 |
| | | | | | GBI:g8118985_000043 _edit.12301- 12444.comp | 685 | 810 |
| | | | | | GBI:g8117242_000054 _edit.4112-4228 | 241 | 357 |
| | | | | | GBI:g8117242_000054 _edit.10957-11181 | 1717 | 1941 |
| | | | | . ' | 5500380H1 (BRABDIR01) | 907 | 1119 |
| | | | | | GBI:g8117242_000054 _edit.10616-10732 | 1600 | 1716 |

Table 4 (cont.)

| Ĺ | 101, m;; al | 1 2 2 2 2 2 | 00000000 | 20100100 | | E/ Doc: 1:00 | 21 50 11 |
|-------------|-------------|-------------------|----------|-----------------------------------|--|----------------|----------|
| S | SEQ ID NO: | Polynucleotide ID | Length | Fragment(s) | ממונים ב המשונים | | |
| 35 | | | | | GBI:g8117242_000054 _edit.8907-9011 | 1336 | 1440 |
| ! | | | | | GBI:g8117242_000054 _edit.6643-6756 | 811 | 924 |
| 36 | | 7473053CB1 | 4971 | 3312-3482, 1- 1466, 4307-4971, | 8035016H1 (SMCRUNE01) | 2315 | 2975 |
| w | | | | 2184-2221 | 6822202J1 (SINTNOR01) | 2145 | 2877 |
| | | | | | 6781747H1 (OVARDIR01) | 896 | 1449 |
| | | | | | 8035016J1 (SMCRUNE01) | 2979 | 3643 |
| | | | | | 6824230H1 (SINTNOR01) | 2867 | 3483 |
| | | | | • | 6894266H1 (BRAITDR03) | 548 | 1157 |
| | | | | | 6777836H1 (OVARDIR01) | 1601 | 2238 |
| 11 | | | | | 6908503H1 (PITUDIR01) | - 1 | 299 |
| 3 | | | | | 6908503J1 (PITUDIR01) | 1270 | 1830 |
| | | | | | 6823447H1 (SINTNOR01) | 3525 | 4260 |
| | | | | | 6823447J1 (SINTNOR01) | 4226 | 4829 |
| I | | | | | 6006310F8 (FIBRUNT02) | 4501 | 4969 |
| | | | | | 4171959T6 (SINTNOT21) | 3637 | 4287 |
| | | | | ! | 5088860F6 (UTRSTMR01) | 4461 | 4853 |
| 37 | | 7473347CB1 | 1404 | 126-633, 1013- 1404, 768-838 | GBI.lee4.edit | . | 1404 |

| Polynucleotide SEQ ID NO: | Incyte Polynucleotide ID | Sequence Length | Selected Fragment(s) | Sequence Fragments | 5' Position | 3' Position |
|------------------------------|-----------------------------|--------------------|---------------------------------|--|-------------|-------------|
| | 7474240CB1 | 4048 | | 71984804V1 | 964 | 1311 |
| | | | 1593-1658, 2614- | 71986624V1 | 1369 | 1976 |
| | | | 2908, 1138-1367 | 55055014H1 | | 130 |
| | | | | 55037111J2 | 95 | 871 |
| | | | | 71983668V1 | 1371 | 2043 |
| | | | | GBI:g5923734_edit | 2612 | 4048 |
| • | | | | 55037119J2 | 224 | 875 |
| | | | | 2502027F6 (ADRETUT05) | | 1235 |
| | 7475338CB1 | 1539 | 1412-1539, 1- 328, 495-837, | GBI:97960701_000004 _edit.549-713 | 154 | 312 |
| | | | | GBI:g7960701_000004 _edit.13381-13480 | 1015 | 1113 |
| | | | | GBI:g7960701_000004 _edit.8755-8943 | 715 | 903 |
| | | | | GBI:g7960701_000004 edit.4292-4417 | 313 | 438 |
| | | | | GBI:g7960701_000004 edit.16237-16317 | 1114 | 1194 |
| | | | | GBI:g7960701_000004 edit.20107-20325 | 1321 | 1539 |
| | | | | GBI:g7960701_000004 _edit.9989-10099 | 904 | 1014 |
| | | | | GBI:g7960701_000004 _edit.18748-18873 | 1195 | 1320 |
| | | | | GBI:97960701_000003 _edit.9783-9884 | | 153 |
| | | | | GBI:g7960701_000004 _edit.5251-5403 | i | 591 |
| | | | | GBI:g7960701_000004 _edit.8384~8506 | | 714 |
| | | - | | 71906448V1 | 627 | 1082 |
| | | | | 71753467V1 | 912 | 1539 |
| | 7476747CB1 | 3114 | 1717-1870, 1- 503, 1468-1650 | 3351512F6 (PROSNOT28) | 2185 | 2724 |
| ·· | | | | 7761783J1 (THYMNOE02) | 1943 | 2570 |
| | | | | 6934981R8 (SINTTMR02) | 78 | . 098 |

Table 4 (cont.)

| لـــــا | Polynucleotide SEQ ID NO: | Incyte Polynucleotide ID | Sequence Length | Selected Fragment(s) | Sequence Fragments | 5' Position | 3' Position |
|-------------|------------------------------|-----------------------------|--------------------|-----------------------------------|----------------------------|---------------|-------------|
| | 40 | | | | 6389368H1 (PROSTMC01) | 1782 | 2075 |
| | | | | | 70536163V1 | 2575 | 3114 |
| | | | | | 6934981F8 (SINTTMR02) | F | 643 |
| | | | | | GNN.97712065_000012 002 | 452 | 1922 |
| | | | | | 7080657H1 (STOMTMR02) | 838 | 1403 |
| | | | | | 5633289H1 (PLACFER01) | 639 | 890 |
| | | | | - | 95746200 | 1215 | 1473 |
| | 41 | 7477898CB1 | 2877 | 846-901, 1272- 1378, 2319-2877 | GBI.g2262095 | 1 | 2877 |
| - | 42 | 7472728CB1 | 2820 | 1-1399, 2207- | 55022826J1 | 1138 | 1834 |
| | | | | 2229 | 55030210H1 | 403 | 986 |
| | | | | | 4399366T6 (TESTTUT03) | 2231 | 2777 |
| _ | | | | | 55030274H1 | 1482 | 2153 |
| 11 | | | | | g565876 | 2597 | 2820 |
| 15 | | | | | 55018149J1 | 1907 | 2585 |
| | | | | | FL203597_00001 | 712 | 1807 |
| | | | | | GNN.g7263861_026.ed | -1 | 1052 |
| | 43 | 7474322CB1 | 1440 | 1-604, 714-768 | GBI.g8081632_edit | 1 | 1440 |
| ; | | | | | 71228887V1 | 1090 | 1440 |
| | | | | | 70868623V1 | 988 | 1385 |
| | 44 | 5455621CB1 | 2394 | 1483-1686, 1- 329, 838-1155, | 3696546T6 (SININOT05) | 1833 | 2394 |
| | | | | 2201-2235 | 70674954V1 | 1520 | 2091 |
| | | | | | 1426382H1 (SINTBST01) | 1224 | 1492 |
| | | | | | 3696546F6 (SININOT05) | 799 | 1381 |
| | | | | | 6828352H1 (SINTNOR01) | 530 | 1149 |
| | | | | | 3699565H1 (SININOT05) | - | 281 |
| | | | | | 7700096H1 (KIDPTDE01) | 250 | 066 |
| ال | | | | | 70678552V1 | 1419 | 2055 |

| Polynucleotide SEQ ID NO: | Incyte Polynucleotide ID | Seguence Length | Selected Fragment(s) | Sequence Fragments | 5' Position | 3' Position |
|------------------------------|-----------------------------|--------------------|-------------------------------------|----------------------------------|-------------|-------------|
| | 7477248CB1 | 2890 | 1-58, 2739-2890, 2310-2349, 329- | 2777287H1 (OVARTUT03) | 2250 | 2498 |
| | | | 1167 | 7977733H1 (LSUBDMC01) | 841 | 1427 |
| | | | | 7678168J1 (NOSETUE01) | 1271 | 1827 |
| | | | | 7611941J1 (KIDCTME01) | 2273 | 2890 |
| | | | | 6590507H1 (TLYMUNT03) | 179 | 672 |
| | | | | 2701794F6 (OVARTUT10) | 1208 | 1741 |
| | | | | 2544096F6 (UTRSNOT11) | 1732 | 2252 |
| | | | | 60117044D2 | 1 | 431 |
| | | | | 5020832H1 (QVARNON03) | 2195 | 2471 |
| | | | | 7662529H1 (UTRSTME01) | 526 | 926 |
| | 2944004CB1 | 3926 | 3338-3365, 1- 687, 1222-2267 | 4762728F6 (PLACNOT05) | 872 | 1387 |
| | | | | 92264624 | 2268 | 2446 |
| | | | | 6264977H1 (MCLDTXN03) | 1210 | 1797 |
| | | | | 2944004F6 (BRAITUT23) | 2790 | 3531 |
| | | | | 6610392H2 (MUSTTMC01) | 3306 | 3926 |
| | | | | GNN.g7328818_000024 _002.edit | 2145 | 2648 |
| | | | | 7035078H1 (SINTFER03) | | 440 |
| | | | | 7620248J1 (HEARFEE03) | 2431 | 3039 |
| | | | | 496537H1 (HNT2NOT01) | 2329 | 2487 |
| | | | | 6264427T8 (MCLDTXN03) | 453 | 1174 |
| | | | | 6264427F8 (MCLDTXN03) | 170 | 842 |

| | Polynucleotide SEQ ID NO: | Incyte Polynucleotide ID | Sequence Length | Selected Fragment(s) | Sequence Fragments | 5' Position | 3' Position |
|---------------|------------------------------|-----------------------------|--------------------|----------------------------------|---------------------------------------|-------------|-------------|
| | 46 | • | | | 7673654H1 (FIBPFEC01) | 1733 | 2239 |
| | 47 | 3046849CB1 | 2135 | 2072-2135, 596- | 8262790U1 | 1383 | 2135 |
| | | | | 1014 | 71896642V1 | 1 | 592 |
| | | | | | 71247870V1 | 1050 | 1736 |
| | | | | | FL3046849_g6815043_ 000004_g183298 | 51 | 1520 |
| | 48 | 4538363CB1 | 2637 | 1-183, 1575- 1680, 2094-2637 | FL4538363_g3126781_ g520469 | т | 1917 |
| | | | | - 1 | 71401405V1 | 1766 | 2637 |
| | 49 | 6427460CB1 | 3783 | 985-1833, 2687- | | 416 | 1035 |
| | | | | 3204 | 7727961J1 (UTRCDIE01) | 3284 | 3783 |
| 1 | | | | | 70857789V1 | 266 | 1109. |
| | | | | | g5689372_edit ' | 1092 | 3361 |
| ! | | | | | g3801917 | 1 | 452 |
| | 20 | 7474127CB1 | 2105 | 1078-2105 | GBI.g8568959_edit_3 | 1119 | 2105 |
| | | | | | g6140313 | 482 | 951 |
| īı | | | | | 5819744F7 (PROSTUS23) | 168 | 479 |
| 7 | | | | | g5920552 | 1 | 488 |
| | | | | | 55049678J1 | 862 | 1359 |
| | 51 | 7476949CB1 | 2069 | 1233-1356, 1- 117, 2047-2069. | FL7476949_g6714723_ | 7 | 2046 |
| | | | | 347~503, 1536- 1844 | 4669722H1 (SINTNOT24) | 1801 | 2069 |
| | 52 | 7477249CB1 | 4245 | 2833-3018, 1869- | 71660072V1 | 2404 | 3156 |
| | | | | 2121, 3707-4245, | 71657569V1 | 3106 | 3854 |
| | | | | 1-252, 982-1239, 289-357 | 7633968J1 (SINTDIE01) | 2579 | 3175 |
| | | | | | 6440145F8 (BRAENOT02) | 938 | 1087 |
| | | | | | 71664080V1 | 3228 | 3891 |
| | | | | | GBI.g8567478.edit | 1 | 2547 |
| | | | | | 71660176V1 | 3773 | 4245 |
| _ | | | | | 71662066V1 | 1802 | 2475 |
| | | | | | 2605539F6 (LUNGTUT07) | 433 | 939 |
| | | | | | 71659261V1 | 1690 | 2437 |
| لــــ | | | | | 3825558H1 (BRAIHCT02) | 1179 | 1270 |

| Polynucleotide SEQ ID NO: | Incyte Polynucleotide ID | Sequence Length | Selected Fragment(s) | Sequence Fragments | 5' Position | 3' Position |
|------------------------------|-----------------------------|----------------------|---|---------------------------------------|-------------|-------------|
| 52 | | | | 7765571H1 (URETTUE01) | 1 | 869 |
| | | | | 5675861H1 | 1427 | 1716 |
| 53 | 7477720CB1 | 2124 | 1-936, 1200- 1488, 1982-2124, 1562-1745 | FL747720_g5836195_ g205709 | r-l | 2124 |
| 54 | 7477852CB1 | 2195 | 1-418, 1899-2195 | | 1 | 2195 |
| 55 | 1471717CB1 | 2055 | 206-768, 881- | 1 | 492 | 994 |
| | | | 931, 1155-1323 | | 1 | 297 |
| | | | | _ 1 | 939 | 1582 |
| | | | | 8 | 772 | 1500 |
| | | | | GBI.g8039708_50_63_ 62_56.edit | 238 | 897 |
| | | | | 6540941H1 (LNODNON02) | 1571 | 2055 |
| | | | | 70466394V1 | 1035 | 1616 |
| 56 | 3874406CB1 | 4727 | 1-1299, 1576- | 71793833V1 | 4117 | 4727 |
| | | | 1632, 2550-3619, | 5505210531 | 1673 | 2128 |
| | | | 2014-2192 | 71798347V1 | 3620 | 4358 |
| | | | | 71798870V1 | 3575 | 4244 |
| | - | - | | 55058313J1 | 1380 | 2125 |
| | | | | 55051482J1 | 2475 | 3134 |
| | | | | FL3874406_g3810670_ g4240130_3_3-4 | 482 | 744 |
| | | | | 55068154H1 | 2223 | 2741 |
| | | | | 3133035F6 (SMCCNOT01) | ⊢ 1 | 605 |
| | - | | | 55058329H1 | 723 | 1528 |
| | | | | 55068182J1 | 2048 | 2685 |
| | | | | 71795307V1 | 2902 | 3593 |
| 57 | 4599654CB1 | 3852 | 1-335, 2014-3231 | 8016331J1 (BMARTXE01) | 1778 | 2424 |
| | | | | 71040001V1 | 3348 | 3852 |
| | | | | 8041905H1 | 1666 | 2352 |
| | | | | 55062505H1 | 660 | 1233 |
| | | | | g7959336_CD | 349 | 2540 |
| | | | | 6772024J1 (BRAUNOR01) | ᆏ | 623 |
| | | | | 55064208J1 | 1118 | 1718 |

| Polynucleotide SEQ ID NO: | Incyte Polynucleotide ID | Sequence Length | Selected Fragment(s) | Sequence Fragments | 5' Position | 3' Position |
|------------------------------|-----------------------------|--------------------|-------------------------|----------------------------------|-------------|-------------|
| 57 | | | | 6617183H2 (BRAXTDR14) | 2981 | 3530 |
| | | | | 6195941H1 (PITUNON01) | 2823 | 3458 |
| | | | | 71909238V1 | 1225 | 1747 |
| | | | | 2216896F6 (SINTFET03) | 2474 | 2923 |
| | | | | 71042073V1 | 2276 | 2745 |
| 58 | 5047435CB1 | 1917 | 1-238, 1162-1474 | 7431853H1 (UTRMTMR02) | 1211 | 1917 |
| | | | | GNN:g4375937_004_ed it | - -I | 1845 |
| | | | | 6426880H1 (LUNGNON07) | 814 | 1336 |
| | | | | 6781142H1 (OVARDIR01) | 224 | 941 |
| | | | | 2645767H1 (OVARNOT09) | 128 | 394 |
| 59 | 7475603CB1 | 16791 | 1-3283, 5952- | 71704421V1 | 6240 | 6791 |
| | | | 6101, 3793-4761 | 7726210H1 (THYRDIE01) | 1885 | 2602 |
| | | | | 7721710J2 (THYRDIE01) | 2696 | 3232 |
| | | | | 6340173F8 (BRANDIN01) | 5516 | 6222 |
| | | | | 71704256V1 | 3025 | 3734 |
| | | | | 7757131H1 (SPLNTUE01) | 2408 | 3093 |
| | | | | GNN.g7711543_000002 _002.edit | 198 | 2751 |
| | | | | 7464813H1 (LIVRFEE04) | 544 | 969 |
| | | | | 71703676V1 | 3250 | 3947 |
| | * | | | 7760618H1 (THYMNOE02) | 2183 | 2676 |
| | | | | 71970086V1 | 5817 | 6525 |
| | | | | 7462584H1 (LIVRFEE04) | T- | 578 |
| | | | | 7760618J1 (THYMNOE02) | 1251 · | 1983 |
| | | | | 71762287V1 | 4313 | 4879 |

| | _ | • |
|---|----------|---|
| • | Cont | |
| ` | 7 | _ |
| | <u>a</u> | 2 |
| - | 2 | 3 |
| ŀ | - | 4 |

| | Polynucleotide SEQ ID NO: | Incyte Polynucleotide ID | Sequence Length | Selected Fragment(s) | Sequence Fragments | 5' Position | 3' Position |
|-------------|------------------------------|-----------------------------|--------------------|-------------------------|--|-------------|-------------|
| | 59 | | | | 7724639H1 (THYRDIE01) | 951 | 1545 |
| | | | - | • | 55052451J1 | 4792 | 5698 |
| | | | - | | 7739867H1 (THYMNOE01) | 5131 | 5794 |
| | | | | | 6879936H1 (UTRSTMR02) | 697 | 1054 |
| | | | | | 55058371HI | 3850 | 4747 |
| | 09 | 7477845CB1 | 5214 | 2390-4599, 645- | | 1765 | 5214 |
| - | | | | 1/36 1/36 | GBI.g8052096_edit | 1132 | 1839 |
| | | | | | 8104845H1 (MIXDDIE02) | 2822 | 3367 |
| | | | | | GBI.98518014_edit | 1 | 1266 |
| <u> </u> | 61 | 168827CB1 | 1818 | 1-281, 796-912 | g1081430 | 1036 | 1525 |
| 2 2 | | | | | 168827H1 (LIVRNOT01) | 65 | 406 |
| =- | ٠ | | | | 55064792J1 | 1 | 209 |
| = | | | | | 55072770H1 | 495 | 1110 |
| 12 | | | | | GNN.g6498074_012.ed | 1321 | 1818 |
| 0 | | | | | 087510H1 (LIVRNOT01) | 314 | 574 |
| | | | | | g751568 | 1336 | 1773 |
| | 62 | 7472734CB1 | 2245 | 1223-1339, 1-710 | 55055559H1 | 16 | 669 |
| === | | | | | 55045003H2 | | 697 |
| <u>-</u> | | | - | | g5361744 | 806 | 1109 |
| | | | | | GBI.g8118965_000015 _000006_000001_0000 10_000003.edit | | 2245 |
| | | | | | g751568 | | 2200 |
| _ | 63 | 7473473CB1 | 3196 | 1-376, 460-1796 | 55049235H1 | | 1287 |
| | | | | | GBI.g8018151_000001 .edit | | 3196 |
| | | | | | GBI.g6433826_000001 .edit | 1172 | 2052 |
| | | | | | 55063069J1 | 1 | 850 |
| _ | | | | | g669271 | 1799 | 2106 |

Table 4 (cont.)

| Polynucleotide Incyte | Incyte | Sequence | Selected | Sequence Fragments 5' Position 3' Position | 5' Position | 3' Position |
|-------------------------|-------------------|----------|-------------|--|-------------|-------------|
| SEQ ID NO: | Polynucleotide ID | Length | Fragment(s) | | | i |
| 64 | 7477725CB1 | 1602 | 1072-1602 | 7455614H1 | 416 | 835 |
| | | | | (LIVRTUE01) | | |
| | | | | 4288148H1 | 112 | 257 |
| | | | | (LIVRDIR01) | | |
| | | | | GBI.g8131631_000007 0000005.edit | 7 | 1602 |
| | | | | g2656651 | 829 | 1084 |

Table 5

| Polynucleotide | Incyte | Representative Library |
|----------------|------------|------------------------|
| SEQ ID NO: | Project ID | |
| 33 | 3474673CB1 | LUNLTMT01 |
| 34 | 4588877CB1 | LUNLTWT01 |
| 35 | 7472214CB1 | BRAENOT04 |
| 36 | 7473053CB1 | SINTNOR01 |
| 38 | 7474240CB1 | ADRETUT05 |
| 39 | 7475338CB1 | SINTNOT18 |
| 40 | 7476747CB1 | SINTTMR02 |
| 42 | 7472728CB1 | TESTTUT03 |
| 43 | 7474322CB1 | SINTBST01 |
| 44 | 5455621CB1 | SININOT05 |
| 45 | 7477248CB1 | UTRSNOT11 |
| 46 | 2944004CB1 | MCLDTXN03 |
| 47 | 3046849CB1 | HNT2AGT01 |
| 48 | 4538363CB1 | PANCNOT07 |
| 49 | 6427460CB1 | BRAUNOR01 |
| 50 | 7474127CB1 | PROSTUS23 |
| 51 | 7476949CB1 | COLNTMC01 |
| 52 | 7477249CB1 | COLNPOT01 |
| 55 | 1471717CB1 | OVARDIT01 |
| 56 | 3874406CB1 | LIVRDIR01 |
| 57 | 4599654CB1 | LUNGNOT23 |
| 58 | 5047435CB1 | OVARDIR01 |
| 59 | 7475603CB1 | THYRDIE01 |
| . 60 | 7477845CB1 | MIXDDIE02 |
| 61 | 168827CB1 | LIVRNOT01 |
| 64 | 7477725CB1 | LIVRTUE01 |
| | | |

Table (

| Library | Vertor | Library Description |
|-----------|--------|---|
| ADRETUT05 | pincy | Library was constructed using RNA isolated from adrenal tumor tissue removed from a 52-year-old Caucasian female during a unilateral adrenalectomy. Pathology indicated a pheochromocytoma. |
| BRAENOT04 | pINCY | Library was constructed using RNA isolated from inferior parietal cortex tissue removed from the brain of a 35-year-old Caucasian male who died from cardiac failure. Pathology indicated moderate leptomeningeal fibrosis and multiple microinfarctions of the cerebral neocortex. Patient history included dilated cardiomyopathy, congestive heart failure, cardiomegaly and an enlarged spleen and liver. |
| BRAUNOR01 | PINCY | This random primed library was constructed using RNA isolated from striatum, globus pallidus and posterior putamen tissue removed from an 81-year-old Caucasian female who died from a hemorrhage and ruptured thoracic aorta due to atherosclerosis. Pathology indicated moderate atherosclerosis involving the internal carotids, bilaterally; microscopic infarcts of the frontal cortex and hippocampus; and scattered diffuse amyloid plaques and neurofibrillary tangles, consistent with age. Grossly, the leptomeninges showed only mild thickening and hyalinization along the superior sagittal sinus. The remainder of the leptomeninges was thin and contained some congested blood vessels. Mild atrophy was found mostly in the frontal poles and lobes, and temporal lobes, bilaterally. Microscopically, there were pairs of Alzheimer type II astrocytes within the deep layers of the neocortex. There was increased satellitosis around neurons in the deep gray matter in the middle frontal cortex. The amygdala contained rare diffuse plaques and neurofibrillary tangles. The posterior hippocampus contained a microscopic area of cystic cavitation with hemosiderin-laden macrophages surrounded by reactive gliosis. Patient history included sepsis, cholangitis, post-operative atelectasis, pneumonia of CAD, cardiomegaly due to left ventricular mphysema, CHF, hypothyroidism, and peripheral vascular disease. |
| COLNPOT01 | pincy | |
| COLNTMC01 | pincy | This large size-fractionated library was constructed using pooled cDNA from three different donors. cDNA was generated using mRNA isolated from colon epithelium tissue removed from a 13-year-old Caucasian female (donor A) who died from a motor vehicle accident; from ascending colon removed from a 29-year-old female (donor |

| Library | Vector | Library Description |
|-----------|-------------|--|
| | | tissue removed from the appendix of a 37-year-old B omectomy, dilation and curettage, right fimbrial rendectomy. Pathology for donor B indicated the proxirgins of small bowel and colon away from the mass loma. Pathology for donor C indicated an unremarkabl atched tumor tissue (donor B) indicated malignant laved (Burkitt's lymphoma, B-cell phenotype), formin of the ileocecal valve, associated with intussuscep lly. The liver and multiple (3 of 12) ileocecal regolved by lymphoma. Pathology for the associated tummultiple uterine leiomyomata. Donor C presented wi an umbilical hernia, and premenopausal menorrhagia. |
| HNT2AGT01 | PBLUESCRIPT | Library was constructed at Stratagene (STR937233), using RNA isolated from the hNT2 cell line derived from a human teratocarcinoma that exhibited properties characteristic of a committed neuronal precursor. Cells were treated with retinoic acid for 5 weeks and with mitotic inhibitors for two weeks and allowed to mature for an additional 4 weeks in conditioned medium. |
| LIVRDIR01 | pincy | The library was constructed using RNA isolated from diseased liver tissue removed from a 63-year-old Caucasian female during a liver transplant. Patient history included primary biliary cirrhosis diagnosed in 1989. Serology was positive for anti-mitochondrial antibody. |
| LIVRNOT01 | PBLUESCRIPT | Library was constructed at Stratagene, using RNA isolated from the liver tissue of a 49-year-old male. |
| LIVRTUE01 | PCDNA2.1 | 5, bias trissue cology in The pay |
| LUNGNOT23 | DINCY | Library was constructed using RNA isolated from left lobe lung tissue removed from |

| Library | Vector | Library Description |
|-----------|---------|---|
| | | a 58-year-old Caucasian male. Pathology for the associated tumor tissue indicated metastatic grade 3 (of 4) osteosarcoma. Patient history included soft tissue cancer, secondary cancer of the lung, prostate cancer, and an acute duodenal ulcer with hemorrhage. Family history included prostate cancer, breast cancer, and acute leukemia. |
| LUNLTMT01 | pINCY | The library was constructed using RNA isolated from right middle lobe lung tissue removed from a 63-year-old Caucasian female during a segmental lung resection. Pathology for the associated tumor tissue indicated grade3 adenocarcinoma in the right lower lobe and right middle lobe that infiltrated the parietal pleural surface. Metastatic grade 3 adenocarcinoma was found in the diaphragm. The lymph nodes contained metastatic grade 3 adenocarcinoma and involved the superior mediastinal and inferior mediastinal lymph nodes. Patient history included hyperlipidemia. Family history included benign hypertension, cerebrovascular disease, breast cancer, and hyperlipidemia. |
| MCLDTXN03 | pINCY | This normalized dendritic cell library was constructed from one million independent clones from a pool of two derived dendritic cell libraries. Starting libraries were constructed using RNA isolated from untreated and treated derived dendritic cells from umbilical cord blood CD34+ precursor cells removed from a male. The cells were derived with granulocyte/macrophage colony stimulating factor (GM-CSF), tumor necrosis factor alpha (TNF alpha), and stem cell factor (SCF). The GM-CSF was added at time 0 at 100 ng/ml, the TNF alpha was added at time 0 at 2.5 ng/ml. Incubation time was 13 days. The treated cells were then exposed to phorbol myristate acetate (PMA), and Ionomycin were added at 13 days for five hours. The library was normalized in two rounds using conditions adapted from Soares et al., PMAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48 hours/round) reannealing hybridization was used. |
| MIXDDIE02 | PBK-CMV | This 5' biased random primed library was constructed using pooled cDNA from seven donors. cDNA was generated using mRNA isolated from brain tissue removed from two Caucasian male fetuses who died after 23 weeks gestation from hypoplastic left heart (A) and prematurity (B); from posterior hippocampus from a 55-year-old male who died from COPD (C); from cerebellum, corpus callosum, thalmus and temporal lobe tissue from a 57-year-old Caucasian male who died from a CVA (D); from dentate nucleus and vermis from an 82-year-old Caucasian male who died from a myocardial infarction (E); from pituitary gland from a 74-year-old Caucasian female who died from a myocardial infarction (F) and vermis tissue from a 77-year-old Caucasian female who died from pneumonia (G). For donor C, pathology indicated |

| Library | Vector | Library Description |
|-----------|------------|--|
| , | | lateral ventricular enlargement. For donor F, pathology is a disease, recent multiple infarctions involving lefetal and occipital lobes (microscopic) and right cerebellus cosclerosis involving middle cerebral arteries bilaterally oid angiopathy. For donor G, pathology indicated severe Alatherosclerosis involving the middle cerebral and basilar arterophy consistent with Alzheimer's disease, For dono ided Huntington's chorea. Donor E was taking nitroglycerin taking Lopressor, heparin, ceftriaxone, captopril, Isorovil, Ecotrin and tacrine; and donor G was taking insulin. |
| OVARDIR01 | PCDNA2 . 1 | This random primed library was constructed using RNA isolated from right ovary tissue removed from a 45-year-old Caucasian female during total abdominal hysterectomy, bilateral salpingo-oophorectomy, vaginal suspension and fixation, and incidental appendectomy. Pathology indicated stromal hyperthecosis of the right and left ovaries. Pathology for the matched tumor tissue indicated a dermoid cyst (benign cystic teratoma) in the left ovary. Multiple (3) intramural leiomyomata were identified. The cervix showed squamous metaplasia. Patient history included metrorrhagia, female stress incontinence, alopecia, depressive disorder, pneumonia, normal delivery, and deficiency anemia. Family history included benign hypertension, atherosclerotic coronary artery disease, hyperlipidemia, and primary tuberculous complex. |
| OVARDIT01 | pINCY | Library was constructed using RNA isolated from diseased ovary tissue removed from a 39-year-old Caucasian female during total abdominal hysterectomy, bilateral salpingo-oophorectomy, dilation and curettage, partial colectomy, incidental appendectomy, and temporary colostomy. Pathology indicated the right and left adnexa were extensively involved by endometriosis. Endometriosis also involved the anterior and posterior serosal surfaces of the uterus and the cul-de-sac and the mesentery and muscularis propria of the sigmoid colon. Pathology for the associated tumor tissue indicated multiple (3 intramural, 1 subserosal) leiomyomata. Family history included hyperlipidemia, benign hypertension, atherosclerotic coronary artery disease, depressive disorder, brain cancer, and type II diabetes. |
| PANCNOT07 | pincy | ורא האו |
| PROSTUS23 | pincy | prostate tumor lil prostate tumor libra pith 10 million clor |

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| restarting library for subtraction was constructed by pooling equal numbers of clones from 4 prostate tumor libraries using mRNA isolated from prostate tumor removed from Caucasian males at ages 58 (A), 61 (B), 66 (C), and 68 (D) during prostatectown with lymph node excision. Pathology indicated adenocarcinoma in all donors. History included elevated PSA, induration and tobacco abuse in donor 1, a letwated PSA, induration and tobacco abuse in donor 1, a letwated PSA, induration and tobacco abuse in donor C; and elevated PSA, induration, prostate hyperplasia, remal failure, osterearthritis, remal artery stenosis, benign HTM, thrombocycopenia, hyperlipidatation probe for subtraction was constructed by pooling equal numbers of confidency from 1 prostate tissue indonor C; and elevated PSA, induration, hypercholesterolemia, and kidhey calculus in donor D. The hybridization probe for and fibroblasts from prostate stromm from 3 different donors. Subtractive hybridization conditions were based on the methodologies of Swaroop et al., NAR (1991.1954 and Bonaldo, et al. Genome Research 6 (1996):791. Library was constructed using RNA isolated from inver biopsy, incidence pependectomy, and permanent colostomy, patient history included endometricisis. Family history included hyperlipidated in anxiety, and upper lobe lung cancer, library was constructed using RNA isolated from tissue of the ileum, involving 15 cm of the small bowel. The ceum and appendix were unremarkable, and the margin by many and permanent colostomy, and observable and during bowel anastenosis. Pathology indicated crohn's disease of the ileum, involving 15 cm of the small bowel. The ceum and appendix were unremarkable, and the margin were unremarkable. Pentrasa (messlamine) and atherosclerotic connary artery disease. PCDNA2.1 Hink random primed library was constructed using RNA isolated from small intestic tissue removed from a 31-year-old caucasian female during RNA isolated from small intestic tissue constructed using RNA isolated and small moved intes | חדות דות דות | VECTOE | DIDIGITY DESCRIPCION |
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| tronal arrery stenosis, benign HTM, thrombocytopenia, hyperlipidemia, tobacco/alcohol abuse and hepatitis C (carrier) in donor B: elevated PSA, induration, and tobacco abuse in donor C; and elevated PSA, induration, hypercholesterolemia, and kidney calculus in donor D. The hybridization probe for subtraction was constructed by pooling equal numbers of coNA clones from 3 prostate tissue libraries derived from prostate tissue, prostate epithelial cell and fibroblasts from prostate stroma from 3 different donors. Subtractive hybridization conditions were based on the methodologies of Swaroop et al., NAR (1991):1554 and Bonaldo, et al. Genome Research 6 (1956):791. Library was constructed using RNA isolated from ileum tissue obtained from a 30-year-old Caucasian female during patrial colectomy, open liver biopsy, incidenta appendent cancer, liver cancer, and cirrhosis. PINCY year-old Caucasian female during patrial colectomy, open liver biopsy, incidenta appendent cancer, liver cancer, and cirrhosis. Family history included hyperlipidemia, anxiety, and upper lobe lung cancer, scometructed using RNA isolated from the ileum tissue of the line time caucasian female. The ileum tissue, along with the cecum and appendix, were removed cancer. Liver cancer, and cirrhosis. Caucasian female. The ileum tissue, along with the cecum and appendix were unremarkable, and the margins were uninvolved. The patient presented with abdominal pain and regional enterities. Patient medications included Prilosec (omeprazole), abnormal blood chemistry. Patient medications included Prilosec (omeprazole), and promary and margins were uninvolved. The patient presented with abdominal pain and regional enterities. Patient medications included prilosec (omeprazole), and promary mas constructed using RNA isolated from small intestine tissue obtained from a 19-year-old male. PCDNA2.1 This random primed library was constructed using RNA isolated from small intesti tissue removed from a 39-year-old male. PCDNA2.1 This random primed library was c | | | , |
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| tissue removed from a 31-year-old Caucasian female during Roux-en-Y gastr bypass. Patient history included clinical obesity. Library was constructed using RNA isolated from small intestine tissue ob from a 59-year-old male. PCDNA2.1 This random primed library was constructed using RNA isolated from small tissue removed from a 59-year-old male. Pathology for the matched tumor t | SINTNOR01 | PCDNA2.1 | random primed library was constructed using RNA isolated from small |
| pINCY Libra from From PCDNA2.1 This tissu | | | e removed from a 31-year-old Caucasian female during Roux-en-Y gastr |
| pINCY Libra From From PCDNA2.1 This tissu | | | s. Patient history included clinical c |
| PCDNA2.1 This | SINTNOT18 | pincy | Nry was constructed using RNA isolated from small intestine a 59-vear-old male |
| tissue removed from a 59-year-old male. Pathology for the matched tumor tissue | SINTTMR02 | PCDNA2.1 | This random primed library was constructed using RNA isolated from small intestine |
| | | | tissue removed from a 59-year-old male. Pathology for the matched tumor tissue |

| Library | Vector | Library Description |
|-----------|----------|---|
| | | indicated multiple (9) carcinoid tumors, grade 1, in the small bowel. The largest tumor was associated with a large mesenteric mass. Multiple convoluted segments of bowel were adhered to the tumor. A single (1 of 13) regional lymph node was positive for malignancy. The peritoneal biopsy indicated focal fat necrosis. |
| TESTTUT03 | pincy | Library was constructed using RNA isolated from right testicular tumor tissue removed from a 45-year-old Caucasian male during a unilateral orchiectomy. Pathology indicated seminoma. Patient history included hyperlipidemia and stomach ulcer. Family history included cerebrovascular disease, skin cancer, hyperlipidemia, acute myocardial infarction, and atherosclerotic coronary artery disease. |
| THYRDIE01 | PCDNA2.1 | This 5' biased random primed library was constructed using RNA isolated from diseased thyroid tissue removed from a 22-year-old Caucasian female during closed thyroid biopsy, partial thyroidectomy, and regional lymph node excision. Pathology indicated adenomatous hyperplasia. The patient presented with malignant neoplasm of the thyroid. Patient history included normal delivery, alcohol abuse, and tobacco abuse. Previous surgeries included myringotomy. Patient medications included an unspecified type of birth control pills. Family history included hyperlipidemia and depressive disorder in the mother; and benign hypertension, congestive heart failure, and chronic leukemia in the grandparent(s). |
| UTRSNOT11 | DINCY | Library was constructed using RNA isolated from uterine myometrial tissue removed from a 43-year-old female during a vaginal hysterectomy and removal of the fallopian tubes and ovaries. Pathology for the associated tumor tissue indicated that the myometrium contained an intramural and a submucosal leiomyoma. Family history included benign hypertension, hyperlipidemia, colon cancer, type II diabetes, and atherosclerotic coronary artery disease. |

Table 7

| Parameter Threshold | | Mismatch <50% | | ESTs: Probability value= 1.0E-8 or less Full Length sequences: Probability value= 1.0E-10 or less | ESTs: fasta E value=1.06E-6 Assembled ESTs: fasta Identity= 95% or greater and Match length=200 bases or greater; fastx E value=1.0E-8 or less Full Length sequences: fastx score=100 or greater | Probability value= 1.0E-3 or less | PFAM hits: Probability value=1.0E-3 or less Signal peptide hits: Score=0 or greater |
|---------------------|--|---|--|--|---|--|---|
| Reference | Applied Biosystems, Foster City, CA. | Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA. | Applied Biosystems, Foster City, CA. | Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410; Altschul, S.F. et al. (1997) Nucleic Acids Res. 25:3389-3402. | Pearson, W.R. and D.J. Lipman (1988) Proc. Natl. Acad Sci. USA 85:2444-2448; Pearson, W.R. (1990) Methods Enzymol. 183:63-98; and Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489. | Henikoff, S. and J.G. Henikoff (1991) Nucleic Acids Res. 19:6565-6572; Henikoff, J.G. and S. Henikoff (1996) Methods Enzymol. 266:88-105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37:417-424. | Krogh, A. et al. (1994) J. Mol. Biol. 235:1501-1531; Somhammer, B.L.L. et al. (1988) Nucleic Acids Res. 26:320-322; Durbin, R. et al. (1998) Our World View, in a Nutshell, Cambridge Univ. Press, pp. 1-350. |
| Description | A program that removes vector sequences and masks ambiguous bases in nucleic acid sequences. | A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences. | A program that assembles nucleic acid sequences. | A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic acid sequences. BLAST includes five functions: blastp, blastn, blastx, tblastn, and tblastx. | A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises as least five functions: fasta, tfasta, fastx, tfastx, and ssearch. | A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions. | An algorithm for searching a query sequence against hidden Markov model (HMM)-based databases of protein family consensus sequences, such as PFAM. |
| Program | ABI FACTURA | ABIPARACEL FDF | ABI AutoAssembler | BLAST | FASTA | BLIMPS | HMMER |

Table 7 (cont.)

| | | 7 27071 | (00110) | |
|------|-------------|---|---|---|
| ٠ | Program | Description | Reference | Parameter Threshold |
| | ProfileScan | An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite. | Gribskov, M. et al. (1988) CABIOS 4:61-66; Gribskov, M. et al. (1989) Methods Enzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221. | Normalized quality scorez GCG-specified "HIGH" value for that particular Prosite motif. Generally, score=1.4-2.1. |
| | Phred | A base-calling algorithm that examines automated sequencer traces with high sensitivity and probability. | Ewing, B. et al. (1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194. | |
| ee | Phrap | A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences. | Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489; Smith, T.F. and M.S. Waterman (1981) J. Mol. Biol. 147:195-197; and Green, P., University of Washington, Seattle, WA. | Score= 120 or greater; Match length= 56 or greater |
| | Consed | A graphical tool for viewing and editing Phrap assemblies. | Gordon, D. et al. (1998) Genome Res. 8:195-202. | |
| ·· · | SPScan | A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides. | Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audic (1997) CABIOS 12:431-439. | Score=3.5 or greater |
| - | TMAP | A program that uses weight matrices to delineate transmembrane segments on protein sequences and determine orientation. | Persson, B. and P. Argos (1994) J. Mol. Biol. 237:182-192; Persson, B. and P. Argos (1996) Protein Sci. 5:363-371. | |
| | TMHMMER | A program that uses a hidden Markov model (HMM) to delineate transmembrane segments on protein sequences and determine orientation. | Sonnharmner, B.L. et al. (1998) Proc. Sixth Inti. Conf. on Intelligent Systems for Mol. Biol., Glasgow et al., eds., The Am. Assoc. for Artificial Intelligence Press, Menlo Park, CA, pp. 175-182. | |
| | Motifs | A program that searches amino acid sequences for pattems that matched those defined in Prosite. | Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI. | 7-221; tge |

What is claimed is:

1. An isolated polypeptide selected from the group consisting of:

- a) a polypeptide comprising an amino acid sequence selected from the group consisting of
 SEQ ID NO:1-32,
 - b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32,
 - c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and
- d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.
 - An isolated polypeptide of claim 1 selected from the group consisting of SEQ ID NO:1 32.

3. An isolated polynucleotide encoding a polypeptide of claim 1.

- 4. An isolated polynucleotide encoding a polypeptide of claim 2.
- An isolated polynucleotide of claim 4 selected from the group consisting of SEQ ID NO:33-64.
 - 6. A recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide of claim 3.
 - 7. A cell transformed with a recombinant polynucleotide of claim 6.
 - 8. A transgenic organism comprising a recombinant polynucleotide of claim 6.
- 9. A method for producing a polypeptide of claim 1, the method comprising:
 - a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said
 cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide
 comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim
 1, and
- b) recovering the polypeptide so expressed.

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10. An isolated antibody which specifically binds to a polypeptide of claim 1.

- 11. An isolated polynucleotide selected from the group consisting of:
- a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64,
 - b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64,
 - c) a polynucleotide complementary to a polynucleotide of a),
 - d) a polynucleotide complementary to a polynucleotide of b), and
- e) an RNA equivalent of a)-d).
 - 12. An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 11.
- 13. A method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 11, the method comprising:
 - a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and
 - b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.
 - 14. A method of claim 13, wherein the probe comprises at least 60 contiguous nucleotides.
 - 15. A method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 11, the method comprising:
 - a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and
- b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.
 - 16. A composition comprising a polypeptide of claim 1 and a pharmaceutically acceptable excipient.

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17. A composition of claim 16, wherein the polypeptide has an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.

- 18. A method for treating a disease or condition associated with decreased expression of
 functional TRICH, comprising administering to a patient in need of such treatment the composition of claim 16.
 - 19. A method for screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
 - b) detecting agonist activity in the sample.
 - 20. A composition comprising an agonist compound identified by a method of claim 19 and a pharmaceutically acceptable excipient.
 - 21. A method for treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment a composition of claim 20.
- 22. A method for screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
 - b) detecting antagonist activity in the sample.
- 23. A composition comprising an antagonist compound identified by a method of claim 22 and a pharmaceutically acceptable excipient.
 - 24. A method for treating a disease or condition associated with overexpression of functional TRICH, comprising administering to a patient in need of such treatment a composition of claim 23.
 - 25. A method of screening for a compound that specifically binds to the polypeptide of claim 1, said method comprising the steps of:
 - a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and

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b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a compound that specifically binds to the polypeptide of claim 1.

- 26. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, said method comprising:
 - a) combining the polypeptide of claim 1 with at least one test compound under conditions permissive for the activity of the polypeptide of claim 1,
 - b) assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and
- c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.
- 27. A method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method comprising:
 - a) exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
 - b) detecting altered expression of the target polynucleotide, and
- 20 c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.
 - 28. A method for assessing toxicity of a test compound, said method comprising:
 - a) treating a biological sample containing nucleic acids with the test compound;
 - b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide of claim 11 under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim 11 or fragment thereof;
 - c) quantifying the amount of hybridization complex; and
 - d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

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- 29. A diagnostic test for a condition or disease associated with the expression of TRICH in a biological sample comprising the steps of:
- a) combining the biological sample with an antibody of claim 10, under conditions suitable for the antibody to bind the polypeptide and form an antibody; polypeptide complex; and
 - b) detecting the complex, wherein the presence of the complex correlates with the presence of the polypeptide in the biological sample.
 - 30. The antibody of claim 10, wherein the antibody is:
- a) a chimeric antibody,
 - b) a single chain antibody,
 - c) a Fab fragment,
 - d) a F(ab')₂ fragment, or
 - e) a humanized antibody.

- 31. A composition comprising an antibody of claim 10 and an acceptable excipient.
- 32. A method of diagnosing a condition or disease associated with the expression of TRICH in a subject, comprising administering to said subject an effective amount of the composition of claim
 31.
 - 33. A composition of claim 31, wherein the antibody is labeled.
- 34. A method of diagnosing a condition or disease associated with the expression of TRICH
 25 in a subject, comprising administering to said subject an effective amount of the composition of claim
 33.
 - 35. A method of preparing a polyclonal antibody with the specificity of the antibody of claim 10 comprising:
- a) immunizing an animal with a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, or an immunogenic fragment thereof, under conditions to elicit an antibody response;
 - b) isolating antibodies from said animal; and

c) screening the isolated antibodies with the polypeptide, thereby identifying a polyclonal antibody which binds specifically to a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.

- 36. An antibody produced by a method of claim 35.
 - 37. A composition comprising the antibody of claim 36 and a suitable carrier.
- 38. A method of making a monoclonal antibody with the specificity of the antibody of claim 10 comprising:
 - a) immunizing an animal with a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, or an immunogenic fragment thereof, under conditions to elicit an antibody response;
 - b) isolating antibody producing cells from the animal;
 - c) fusing the antibody producing cells with immortalized cells to form monoclonal antibodyproducing hybridoma cells;
 - d) culturing the hybridoma cells; and
 - e) isolating from the culture monoclonal antibody which binds specifically to a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.
 - 39. A monoclonal antibody produced by a method of claim 38.
 - 40. A composition comprising the antibody of claim 39 and a suitable carrier.
- 41. The antibody of claim 10, wherein the antibody is produced by screening a Fab expression library.
 - 42. The antibody of claim 10, wherein the antibody is produced by screening a recombinant immunoglobulin library.
 - 43. A method for detecting a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32 in a sample, comprising the steps of:
 - a) incubating the antibody of claim 10 with a sample under conditions to allow specific binding of the antibody and the polypeptide; and

| 1 | 36 | | _ | _ | |
|---|----|------|---|---|--|
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b) detecting specific binding, wherein specific binding indicates the presence of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32 in the sample.

- 44. A method of purifying a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32 from a sample, the method comprising:
 - a) incubating the antibody of claim 10 with a sample under conditions to allow specific binding of the antibody and the polypeptide; and
- b) separating the antibody from the sample and obtaining the purified polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.
 - 45. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:1.
 - 46. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:2.
 - 47. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:3.
 - 48. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:4.
- 49. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:5.
 - 50. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:6.
 - 51. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:7.
 - 52. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:8.
 - 53. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:9.
- 30 54. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:10.
 - 55. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:11.
 - 56. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:12.

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57. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:13. 58. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:14. 5 59. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:15. 60. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:16. 61. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:17. 10 62. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:18. 63. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:19. 64. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:20. 15 65. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:21. 66. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:22. 20 67. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:23. 68. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:24. 25 69. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:25. 70. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:26. 71. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:27. 30 72. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:28. 73. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:29. 74. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:30.

- 75. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:31.
- 76. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:32.
- 77. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:33.
- 78. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:34.
 - 79. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:35.
 - 80. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:36.
- 81. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:37.
 - 82. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:38.
- 83. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:39.
 - 84. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:40.
 - 85. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:41.

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86. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:42.

- 87. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:43.
 - 88. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:44.
- 89. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:45.
 - 90. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:46.
 - 91. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:47.
- 92. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:48.
 - 93. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:49.
- 94. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:50.
 - 95. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:51.
 - 96. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:52.

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97. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:53.

- 98. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID 5 NO:54.
 - 99. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:55.
- 100. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:56.
 - 101. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:57.
 - 102. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:58.
- 103. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID20 NO:59.
 - 104. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:60.
- 25 105. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:61.
 - 106. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:62.
 - 107. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:63.

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108. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:64.

```
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      RAUMANN, Brigitte E.
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      GREENE, Barrie D.
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Tyr Val Ile Gly Met Asn Pro Ser Gln Arg Tyr Pro Leu Trp Tyr

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Lys Asn Met Val Ser Leu Trp Ile Leu Phe Gly Met Ala Trp Leu
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Ala Leu Ile Ile Lys Leu Ile Leu Ser Gln Leu Glu Thr Pro Gly
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Arg Val Cys Ser Cys Cys His His Ser Ser Lys Glu Asp Phe Lys
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Ser Gln Ser Trp Arg Gln Gly Pro Asp Arg Glu Pro Glu Ser His
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Ser Pro Gln Gln Gly Cys Tyr Pro Glu Gly Pro Met Gly Ile
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Glu Ala Gln Arg Phe Ser His Leu Pro Lys Arg Ser Ala Val Asp
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Trp Arg Lys Arg Gly Tyr Lys Thr Leu Leu Lys Cys Leu Ser Gly
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Lys Phe Cys Arg Arg Glu Leu Ile Gly Ile Met Gly Pro Ser
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Ala Gly Lys Ser Thr Phe Met Asn Ile Leu Ala Gly Tyr Arg Glu
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Ser Gly Met Lys Gly Gln Ile Leu Val Asn Gly Arg Pro Arg Glu
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Leu Arg Thr Phe Arg Lys Met Ser Cys Tyr Ile Met Gln Asp Asp
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Val Thr Glu Ile Leu Thr Ala Leu Gly Leu Met Ser Cys Ser His
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Thr Arg Thr Ala Leu Leu Ser Gly Gly Gln Arg Lys Arg Leu Ala
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Glu Pro Thr Ser Gly Leu Asp Ser Ala Ser Cys Phe Gln Val Val
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Ser Leu Met Lys Ser Leu Ala Gln Gly Gly Arg Thr Ile Ile Cys
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Thr Ile His Gln Pro Ser Ala Lys Leu Phe Glu Met Phe Asp
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Leu Tyr Ile Leu Ser Gln Gly Gln Cys Ile Phe Lys Gly Val Val
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Thr Asn Leu Ile Pro Tyr Leu Lys Gly Leu Gly Leu His Cys Pro
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Thr Tyr His Asn Pro Ala Asp Phe Val Ile Glu Val Ala Ser Gly

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Glu Tyr Gly Asp Leu Asn Pro Met Leu Phe Arg Ala Val Gln Asn
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Gly Leu Cys Ala Met Ala Glu Lys Lys Ser Ser Pro Glu Lys Asn
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Glu Val Pro Ala Pro Cys Pro Pro Cys Pro Pro Glu Val Asp Pro
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Ile Glu Ser His Thr Phe Ala Thr Ser Thr Leu Thr Gln Phe
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Ile Leu Phe Lys Arg Thr Phe Leu Ser Ile Leu Arg Asp Thr Val
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Leu Thr His Leu Arg Phe Met Ser His Val Val Ile Gly Val Leu
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Ile Gly Leu Leu Tyr Leu His Ile Gly Asp Asp Ala Ser Lys Val
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Phe Asn Asn Thr Gly Cys Leu Phe Phe Ser Met Leu Phe Leu Met
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Phe Ala Ala Leu Met Pro Thr Val Leu Thr Val Pro Leu Glu Met
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Ala Tyr Tyr Leu Ala Lys Thr Met Aia Asp Val Pro Phe Gln Val
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Thr Ala Ile Pro Val Leu Leu Phe Ser Gly Phe Phe Val Ser Phe
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Lys Thr Ile Pro Thr Tyr Leu Gln Trp Ser Ser Tyr Leu Ser
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Val Arg Tyr Gly Phe Glu Gly Val Ile Leu Thr Ile Tyr Gly Met
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Glu Arg Gly Asp Leu Thr Cys Leu Glu Glu Arg Cys Pro Phe Arg
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Leu Tyr Met Asp Phe Leu Val Leu Gly Ile Phe Phe Leu Ala Leu
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Ser Asn Cys Ile Lys Thr Ser Lys Tyr Asn Ile Leu Thr Phe Leu

Pro Val Asn Leu Phe Glu Gln Phe Gln Glu Val Ala Asn Thr Tyr

Phe Leu Phe Leu Leu Ile Leu Gln Leu Ile Pro Gln Ile Ser Ser

4/87

| | | | | 65 | | | | | 70 | | | | | 75 |
|-----|-----|-----|-----|------------|-----|-----|-----|-----|------------|-----|-----|-----|-----|------------|
| Leu | Ser | Trp | Phe | | Thr | Ile | Val | Pro | | Val | Leu | Val | Leu | - |
| Ile | Thr | Ala | Val | Lys 95 | Asp | Ala | Thr | Asp | Asp 100 | Tyr | Phe | Arg | His | Lys 105 |
| Ser | Asp | Asn | Gln | Val 110 | Asn | Asn | Arg | Gln | Ser 115 | Gln | Val | Leu | Ile | Asn 120 |
| Gly | Ile | Leu | Gln | | Glu | Gln | Trp | Met | | Val | Cys | Val | Gly | Asp 135 |
| Ile | Ile | Lys | Leu | | Asn | Asn | Gln | Phe | | Ala | Ala | Asp | Leu | |
| Leu | Leu | Ser | Ser | | Glu | Pro | His | Gly | | Cys | Tyr | Ile | Glu | |
| Ala | Glu | Leu | Asp | | Glu | Thr | Asn | Met | | Val | Arg | Gln | Ala | |
| Pro | Val | Thr | Ser | | Leu | Gly | Asp | Ile | | Lys | Leu | Ala | Lys | |
| Asp | Gly | Glu | Val | | Cys | Glu | Pro | Pro | | Asn | Lys | Leu | Asp | |
| Phe | ser | Gly | Thr | | Tyr | Trp | Lys | Glu | | Lys | Phe | Pro | Leu | |
| Asn | Gln | Asn | Met | | Leu | Arg | Gly | Cys | | Leu | Arg | Asn | Thr | |
| Trp | Cys | Phe | Gly | Leu 245 | Val | Ile | Phe | Ala | Gly 250 | Pro | Asp | Thr | Lys | Leu 255 |
| Met | Gln | Asn | Ser | | Arg | Thr | Lys | Phe | | Arg | Thr | Ser | Ile | |
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| Cys | Met | Gly | Val | Ile 290 | Leu | Ala | Ile | Gly | Asn 295 | Ala | Ile | Trp | Glu | His 300 |
| Glu | Val | Gly | Met | Arg 305 | Phe | Gln | Val | Tyr | Leu 310 | Pro | Trp | Asp | Glu | |
| Val | Asp | Ser | Ala | Phe 320 | Phe | Ser | Gly | Phe | Leu 325 | Ser | Phe | Trp | Ser | Tyr 330 |
| Ile | Ile | Ile | Leu | Asn 335 | Thr | Val | Val | Pro | Ile 340 | Ser | Leu | Tyr | Val | Ser 345 |
| Val | Glu | Val | Ile | Arg 350 | Leu | Gly | His | Ser | Tyr 355 | Phe | Ile | Asn | Trp | Asp 360 |
| Lys | Lys | Met | Phe | Cys 365 | Met | Lys | Lys | Arg | Thr 370 | Pro | Ala | Glu | Ala | Arg 375 |
| Thr | Thr | Thr | Leu | Asn 380 | Glu | Glu | Leu | Gly | Ġln 385 | Val | Glu | Tyr | Ile | Phe 390 |
| Ser | Asp | Lys | Thr | Gly 395 | Thr | Leu | Thr | Gln | Asn 400 | Ile | Met | Val | Phe | Asn 405 |
| Lys | Cys | ser | Ile | Asn 410 | Gly | His | Ser | Tyr | Gly 415 | Asp | Val | Phe | Asp | Val 420 |
| Leu | Gly | His | Lys | Ala 425 | Glu | Leu | Gly | Glu | Arg 430 | Pro | Glu | Pro | Val | Asp 435 |
| Phe | Ser | Phe | Asn | Pro 440 | Leu | Ala | Asp | Lys | Lys 445 | Phe | Leu | Phe | Trp | Asp 450 |
| Pro | Ser | Leu | Leu | Glu 455 | Ala | Va1 | Lys | Ile | Gly 460 | Asp | Pro | His | Thr | His 465 |
| Glu | Phe | Phe | Arg | Leu 470 | Leu | Ser | Leu | Cys | His 475 | Thr | Val | Met | Ser | Glu 480 |
| Glu | ГЛЗ | Asn | Glu | Gly 485 | Glu | Leu | Tyr | Tyr | Lys 490 | Ala | Gln | Ser | Pro | |
| Glu | Gly | Ala | Leu | Val 500 | Thr | Ala | Ala | Arg | Asn 505 | Phe | Gly | Phe | Val | Phe 510 |
| Arg | Ser | Arg | Thr | | Lys | Thr | Ile | Thr | | His | Glu | Met | Gly | |
| Ala | Ile | Thr | Tyr | | Leu | Leu | Ala | Ile | | Asp | Phe | Asn | Asn | |
| Arg | Lys | Arg | Met | | Val | Ile | Val | Arg | | Pro | Glu | Gly | Lys | Ile 555 |
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| His | His | Ser | Thr | Gln | Glu | Leu | Leu | Asn | Thr | Thr | Met | Asp | His | Leu |
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| | _ | | _ | 605 | | | | | 610 | | | | | 615 |
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| Ser | Ile | Tyr | Glu | Glu 635 | Val | Glu | Asn | Asn | Met 640 | Met | Leu | Leu | Gly | Ala 645 |
| Thr | Ala | Ile | Glu | Asp 650 | Lys | Leu | Gln | Gln | Gly 655 | Val | Pro | Glu | Thr | Ile 660 |
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| Gly | Glu | Tyr | Ala | Leu 755 | Val | Ile | Asn | Gly | His 760 | Ser | Leu | Ala | His | Ala 765 |
| Leu | Glu | Ala | Asp | | Glu | Leu | Glu | Phe | | Glu | Thr | Ala | Cys | Ala 780 |
| Cys | Lys | Ala | Val | Ile | Cys | ,Cys | Arg | Val | | Pro | Leu | Gln | Lys | |
| Gln | Val | Val | Glu | | Val | Lys | Lys | Tyr | Lys | Lys | Ala | Val | Thr | Leu |
| Ala | Ile | Gly | Asp | 800 Gly | Ala | Asn | Asp | Val | | Met | Ile | Lys | Thr | |
| His | Ile | Gly | Val | 815 Gly | Ile | Ser | Gly | Gln | 820 Glu | Gly | Ile | Gln | Ala | 825 Val |
| Leu | Ala | Ser | Asp | 830 Tvr | Ser | Phe | Ser | Gln | 835 Phe | Lvs | Phe | Leu | Gln | 840 Arg |
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| | | | | 875 | | | ГÀг | | 880 | | | | | Val 885 |
| His | Phe | Trp | Phe | Gly 890 | Phe | Phe | Cys | Gly | Phe 895 | Ser | Ala | Gln | Thr | Val 900 |
| Tyr | Asp | Gln | Tyr | Phe 905 | Ile | Thr | Leu | Tyr | Asn 910 | Ile | Val | Tyr | Thr | Ser 915 |
| Leu | Pro | Val | Leu | | | Gly | Val | Phe | | Gln | Asp | Val | Pro | Glu 930 |
| Gln | Arg | Ser | Met | Glu | | Pro | Lys | Leu | Tyr | Glu | Pro | Gly | Gln | Leu 945 |
| Asn | Leu | Leu | Phe | | Lys | Arg | Glu | Phe | | Ile | Cys | Ile | Ala | Gln |
| Gly | Ile | Tyr | Thr | 950 Ser | Val | Leu | Met | Phe | 955 Phe | Ile | Pro | Tyr | Gly | 960 Val |
| Phe | Ala | Asp | Ala | 965 Thr | Arg | Asp | Asp | Gly | 970 Thr | Gln | Leu | Ala | Asp | 975 Tyr |
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| | | | | 1010 | | | | | 1015 | | | | | 1020 |
| His | Phe | Phe | Ile | Trp 1025 | | Ser | Leu | | Val 1030 | | Phe | Ala | Ile | Leu 1035 |
| Phe | Ala | Met | His | | Asn | Gly | Leu | Phe | Asp 1045 | | Phe | Pro | Asn | Gln 1050 |
| Phe | Arg | Phe | | Gly | Asn | Ala | Gln | | Thr | Leu | Ala | Gln | Pro | Thr 1065 |
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Arg Ser Gly Tyr Ala Phe Ser His Gln Glu Gly Phe Gly Glu Leu
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                                    1150
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                                                           60
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                                      70
Pro Gly Phe Gly Gly Ser Pro Val Pro Val Gly Ile Asp Val His
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                                      85
                                                           90
Val Glu Ser Ile Asp Ser Ile Ser Glu Thr Asn Met Asp Phe Thr
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                                     100
                                                          105
Met Thr Phe Tyr Leu Arg His Tyr Trp Lys Asp Glu Arg Leu Ser
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                                     115
                                                          120
Phe Pro Ser Thr Ala Asn Lys Ser Met Thr Phe Asp His Arg Leu
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                                                          135
Thr Arg Lys Ile Trp Val Pro Asp Ile Phe Phe Val His Ser Lys
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Arg Ser Phe Ile His Asp Thr Thr Met Glu Asn Ile Met Leu Arg
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                                     160
                                                          165
Val His Pro Asp Gly Asn Val Leu Leu Ser Leu Arg Ile Thr Val
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Ser Ala Met Cys Phe Met Asp Phe Ser Arg Phe Pro Leu Asp Thr
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Gln Asn Cys Ser Leu Glu Leu Glu Ser Tyr Ala Tyr Asn Glu Asp
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Asp Leu Met Leu Tyr Trp Lys His Gly Asn Lys Ser Leu Asn Thr
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Glu Glu His Met Ser Leu Ser Gln Phe Phe Ile Glu Asp Phe
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Ala Ser Ser Gly Leu Ala Phe Tyr Ser Ser Thr Gly Trp Tyr Asn
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Val Leu Pro Thr Tyr Tyr Pro Ala Ile Leu Met Val Met Leu Ser
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Trp Val Ser Phe Trp Ile Asp Arg Arg Ala Val Pro Ala Arg Val
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Ser Leu Gly Ile Thr Thr Val Leu Thr Met Ser Thr Ile Ile Thr
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Ala Val Ser Ala Ser Met Pro Gln Val Ser Tyr Leu Lys Ala Val
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Asp Val Tyr Leu Trp Val Ser Ser Leu Phe Val Phe Leu Ser Val
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Ile Glu Tyr Ala Ala Val Asn Tyr Leu Thr Thr Val Glu Glu Arg
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Lys Gln Phe Lys Lys Thr Gly Lys Ile Ser Arg Met Tyr Asn
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Asp Ala Val Gln Ala Met Ala Phe Asp Gly Cys Tyr His Asp
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Glu Ile Asp Met Asp Gln Thr Ser Leu Ser Leu Asn Ser Glu Asp
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Phe Met Arg Arg Lys
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Arg Ile Lys Arg Arg Lys Ser Leu Gly Gly His Val Gly Arg Ile
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Ile Leu Glu Asn Asn His Val Ile Asp Thr Tyr Ser Arg Ile Leu
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                                                           90
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                                                           105
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                                     115
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Glu Asn Ala Ala Thr Pro Glu Arg Val Asn Pro Ile Leu Pro Ile
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Lys Thr Val Asn Arg Lys Phe Phe Gly Phe Lys Phe Pro Gly Leu
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                                     160
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Arg Val Leu Thr Tyr Arg Lys Gln Ser Leu Pro Gln Glu Asp Pro
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                                     175
                                                           180
Asp Val Val Val Ile Asp Ser Ser Lys His Ser Asp Asp Ser Val
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                                     190
                                                           195
Ala Met Lys His Phe Lys Ser Pro Thr Lys Glu Ser Cys Ser Pro
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Ser Glu Ala Asp Asp Thr Lys Ala Leu Ile Gln Pro Ser Lys
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Ser Pro Leu Val Asn Ile Ser Gly Pro Leu Asp His Ser Ser Pro

| | | | | 230 | | | | | 235 | | | | | 240 |
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| Lys | Arg | Gln | Trp | Asp 245 | Arg | Leu | Tyr | Pro | | Met | Leu | Gln | Ser | 240 Ser 255 |
| Ser | Gln | Leu | Ser | His 260 | Ser | Arg | Ser | Arg | | Ser | Leu | Суѕ | Ser | |
| Arg | Arg | Ala | Ser | | Val | His | Asp | Ile | | Gly | Phe | Gly | Val | |
| Pro | Lys | Asn | Ile | Phe 290 | Arg | Asp | Arg | His | Ala 295 | Ser | Glu | Asp | Asn | Gly |
| Arg | Asn | Val | Lys | Gly 305 | Pro | Phe | Asn | His | Ile 310 | Lys | Ser | Ser | Leu | Leu 315 |
| Gly | Ser | Thr | Ser | Asp 320 | Ser | Asn | Leu | Asn | Lys 325 | Tyr | Ser | Thr | Ile | Asn 330 |
| Lys | Ile | Pro | Gln | Leu 335 | Thr | Leu | Asn | Phe | Ser 340 | Glu | Val | Lys | Thr | Glu 345 |
| | | | | Ser 350 | | | | | 355 | | | | | 360 |
| | | | _ | Asp 365 | _ | | | | 370 | | | _ | | 375 |
| | | | | Leu 380 | | | | | 385 | | | _ | _ | 390 |
| | | | | Ile 395 | | _ | | | 400 | ъeu | | _ | | 405 |
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| | | | | Thr 425 | | _ | | | 430 | | | | | 435 |
| | | | | Lys 440 | | | | | 445 | | | | | 450 |
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| | | | | Phe 470 | | | | | 475 | | | | | 480 |
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| _ | | | | Asp 500 | | | | | 505 | | | _ | | 510 |
| | | | | Gly 515 | | | | | 520 | | | | | 525 |
| | _ | - | | Arg 530 | | | | | 535 | _ | | | _ | 540 |
| | | | | Ser 545 | | | | | 550 | | | | | 555 |
| | | | | Ala 560 | | | | | 565 | | | | | 570 |
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| | | | | Ser 605 | | | | | 610 | | | | _ | 615 |
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| | | • | | Pro 635 | | | | | 640 | | | | | 645 |
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| | | | | Met 680 | | | | | 685 | | | | | 690 |
| | | | | Pro 695 | | | | | 700 | | | | | 705 |
| | | | | Tyr 710 | | | | | 715 | | | | | 720 |
| Lys | Gly | Phe | Pro | Glu 725 | Cys | Leu | Gln | Ala | Asp 730 | Ile | Cys | Leu | His | Leu 735 |

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Asn Gln Thr Leu Leu Gln Asn Cys Lys Ala Phe Arg Gly Ala Ser
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Lys Gly Cys Leu Arg Ala Leu Ala Met Lys Phe Lys Thr Thr His
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Ala Pro Pro Gly Asp Thr Leu Val His Cys Gly Asp Val Leu Thr
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                                     775
Ala Leu Tyr Phe Leu Ser Arg Gly Ser Ile
                                         Glu Ile Leu Lys Asp
                785
                                     790
Asp Ile Val Val Ala Ile Leu Gly Lys Asn Asp Ile Phe Gly Glu
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                                     805
Met Val His Leu Tyr
                    Ala Lys Pro Gly Lys Ser Asn Ala Asp Val
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                                                          825
                    Cys Asp Leu His Lys Ile Gln Arg Glu Asp
Arg Ala Leu Thr Tyr
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                                     835
                                                          840
Leu Leu Glu Val Leu Asp Met Tyr Pro Glu Phe Ser Asp His Phe
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Leu Thr Asn Leu Glu Leu Thr Phe Asn Leu Arg His Glu Ser Ala
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                                     865
                                                          870
Lys Ala Asp Leu Leu Arg Ser Gln Ser Met Asn Asp Ser Glu Gly
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Asp Asn Cys Lys Leu Arg Arg Lys Leu Ser Phe Glu Ser Glu
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                                     895
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Gly Glu Lys Glu Asn Ser Thr Asn Asp Pro Glu Asp Ser Ala Asp
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Thr Ile Arg His Tyr Gln Ser Ser Lys Arg His Phe Glu Glu Lys
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Lys Ser Arg Ser Ser Ser Phe Ile Ser Ser
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                                     940
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Lys Pro Leu Phe Ser Gly Ile Val Asp Ser Ser Pro Gly Ile Gly
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Lys Ala Ser Gly Leu Asp Phe Glu Glu Thr Val Pro Thr Ser Gly
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Arg Met His Ile Asp Lys Arg Ser His Ser Cys Lys Asp Ile Thr
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                                                          990
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Asp Met Arg Ser Trp Glu Arg Glu Asn Ala His Pro Gln Pro Glu
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                                    1000
Asp Ser Ser Pro Ser Ala Leu Gln Arg Ala Ala Trp Gly Ile Ser
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                                    1015
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Glu Thr Glu Ser Asp Leu Thr Tyr Gly Glu Val Glu Gln Arg Leu
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                                    1030
Asp Leu Leu Gln Glu Gln Leu Asn Arg Leu Glu Ser Gln Met Thr
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Thr Asp Ile Gln Thr Ile Leu Gln Leu Leu Gln Lys Gln Thr Thr
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Val Val Pro Pro Ala Tyr Ser Met Val Thr Ala Gly Ser Glu Tyr
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Gln Arg Pro Ile Ile Gln Leu Met Arg Thr Ser Gln Pro Glu Ala
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Ser Ile Lys Thr Asp Arg Ser Phe Ser Pro Ser Ser Gln Cys Pro
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Glu Phe Leu Asp Leu Glu Lys Ser Lys Leu Lys Ser Lys Glu Ser
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Leu Ser Ser Gly Val His Leu Asn Thr Ala Ser Glu Asp Asn Leu
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Thr Ser Leu Leu Lys Gln Asp Ser Asp Leu Ser Leu Glu Leu His
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Leu Arg Gln Arg Lys Thr Tyr Val His Pro Ile Arg His Pro Ser
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                                    1165
                                                         1170
Leu Pro Asp Ser Ser Leu Ser Thr Val Gly Ile Val Gly Leu His
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Arg His Val Ser Asp Pro Gly Leu Pro Gly Lys
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Phe Ala Gly Ile Cys Leu Leu Thr Ala Ile Tyr Ile Tyr Val Val

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Ile Pro Glu Thr Lys Gly Lys Thr Phe Val Glu Ile Asn Arg Ile
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                470
Phe Ala Lys Arg Asn Arg Val Lys Leu Pro Glu Glu Lys Glu Glu
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Ser Ser Thr Asp Val Ser Pro Glu Glu Ser Pro Ser Glu Gly Leu
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Asn Asn Leu Ser Ser Pro Gly Ser Tyr Gln Arg Phe Gly Gln Ser
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                                      70
Asn Ser Thr Trp Phe Gln Thr Leu Ile His Leu Lys Gly
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                                      85
                                                           90
Asn Ile Gly Thr Gly Leu Leu Gly Leu Pro Leu Ala Val Lys Asn
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                                     100
Ala Gly Ile Val Met Gly Pro Ile Ser Leu Leu Ile Ile Gly Ile
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                                     115
Val Ala Val His Cys Met Gly Ile Leu Val Lys Cys Ala His His
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                                     130
Phe Cys Arg Arg Leu Asn Lys Ser Phe Val Asp Tyr Gly Asp Thr
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                                     145
                                                          150
Val Met Tyr Gly Leu Glu Ser Ser Pro Cys Ser Trp Leu Arg Asn
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                                     160
                                                          165
His Ala His Trp Gly Arg Arg Val Val Asp Phe Phe Leu Ile Val
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                                     175
                                                          180
Thr Gln Leu Gly Phe Cys Cys Val Tyr Phe Val Phe Leu Ala Asp
                185
                                     190
Asn Phe Lys Gln Val Ile Glu Ala Ala Asn Gly Thr Thr Asn Asn
                200
                                     205
Cys His Asn Asn Glu Thr Val Ile Leu Thr Pro Thr Met Asp Ser
                                     220
                                                          225
                215
Arg Leu Tyr Met Leu Ser Phe Leu Pro Phe Leu Val Leu Leu Val
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                230
                                     235
Phe Ile Arg Asn Leu Arg Ala Leu Ser Ile Phe Ser Leu Leu Ala
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                                                          255
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Asn Ile Thr Met Leu Val Ser Leu Val Met Ile Tyr Gln Phe Ile
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                                     265
Val Gln Arg Ile Pro Asp Pro Ser His Leu Pro Leu Val Ala Pro
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                                     280
Trp Lys Thr Tyr Pro Leu Phe Phe Gly Thr Ala Ile Phe Ser Phe
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                                     295
                                                          300
Glu Gly Ile Gly Met Val Leu Pro Leu Glu Asn Lys Met Lys Asp
                305
                                     310
                                                          315
Pro Arg Lys Phe Pro Leu Ile Leu Tyr Leu Gly Met Val Ile Val
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                                     325
Thr Ile Leu Tyr Ile Ser Leu Gly Cys Leu Gly Tyr Leu Gln Phe
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                                     340
Gly Ala Asn Ile Gln Gly Ser Ile Thr Leu Asn Leu Pro Asn Cys
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Trp Leu Tyr Gln Ser Val Lys Leu Leu Tyr Ser Ile Gly Ile Phe
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Phe Thr Tyr Ala Leu Gln Phe Tyr Val Pro Ala Glu Ile Ile Ile
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                                     385
                                                          390
Pro Phe Phe Val Ser Arg Ala Pro Glu Pro Cys Glu Leu Val Val
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                                     400
Asp Leu Phe Val Arg Pro Val Leu Val Cys Leu Thr Ser Leu Ser
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Gly Ser Val Asp Asn Gly Trp Tyr Gly Thr Glu Ala Asp Gly Thr
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Ser Cys Gly Ser Ala Pro Leu Val Phe Val Ser Ser Ser Phe Leu
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                                     445
                                                          450
Ala His Pro Trp Leu Ser Phe Arg Cys Glu Ser Gln Trp Val Ser
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                                     460
                                                          465
Cys His Arg Asp Thr Val Val Val Trp Gly Phe Ala Arg Gly Ile
                470
                                     475
                                                          480
Leu Ala Ile Leu Ile Pro Arg Leu Asp Leu Val Ile Ser Leu Val
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                                     490
                                                          495
Gly Ser Val Ser Ser Ser Ala Leu Ala Leu Ile Ile Pro Pro Leu
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                                      505
                                                          510
Leu Glu Val Thr Thr Phe Tyr Ser Glu Gly Met Ser Pro Leu Thr
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                                     520
                                                          525
Ile Phe Lys Asp Ala Leu Ile Ser Ile Leu Gly Phe Val Gly Phe
                530
                                     535
                                                          540
Val Val Gly Thr Tyr Glu Ala Leu Tyr Glu Leu Ile Gln Pro Ser
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Asn Ala Pro Ile Phe Ile Asn Ser Thr Cys Ala Phe Ile
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Ile Ala Asn Ala Gln Met Glu Asn Cys Ala Ile Ile Tyr Cys Asn
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Asp Gly Phe Cys Glu Leu Phe Gly Tyr Ser Arg Val Glu Val Met
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Gln Gln Pro Cys Thr Cys Asp Phe Leu Thr Gly Pro Asn Thr Pro
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                                      70
Ser Ser Ala Val Ser Arg Leu Ala Gln Ala Leu Leu Gly Ala Glu
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                                      85
Glu Cys Lys Val Asp Ile Leu Tyr Tyr Arg Lys Asp Ala Ser Ser
                 95
                                      100
                                                          105
Phe Arg Cys Leu Val Asp Val Val Pro Val Lys Asn Glu Asp Gly
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                                      115
                                                          120
Ala Val Ile Met Phe Ile Leu Asn Phe Glu Asp Leu Ala Gln Leu
                125
                                      130
Leu Ala Lys Cys Ser Ser Arg Ser Leu Ser Gln Arg Leu Leu Ser
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Gln Ser Phe Leu Gly Ser Glu Gly Ser His Gly Arg Pro Gly Gly
                155
                                      160
                                                          165
Pro Gly Pro Gly Thr Gly Arg Gly Lys Tyr Arg Thr Ile Ser Gln
                170
                                      175
                                                          180
Ile Pro Gln Phe Thr Leu Asn Phe Val Glu Phe Asn Leu Glu Lys
                185
                                      190
                                                          195
His Arg Ser Ser Ser Thr Thr Glu Ile Glu Ile Ile Ala Pro His
                                      205
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| Lys | Val | Val | Glu | Arg 215 | Thr | Gln | Asn | Val | Thr 220 | Glu | Lys | Val | Thr | Gln 225 |
|-----|-----|-----|------------|------------|-----|-----|-----|-----|------------|-----|-----|-----|-----|------------|
| Val | Leu | Ser | Leu | Gly 230 | Ala | Asp | Val | Leu | | Glu | Tyr | Lys | Leu | |
| Ala | Pro | Arg | Ile | | Arg | Trp | Thr | Ile | | His | Tyr | Ser | Pro | |
| Lys | Ala | Val | Trp | Asp 260 | Trp | Leu | Ile | Leu | Leu 265 | Leu | Val | Ile | Tyr | Thr 270 |
| Ala | Va1 | Phe | Thr | Pro 275 | Tyr | Ser | Ala | Ala | Phe 280 | Leu | Leu | Ser | Asp | Gln 285 |
| Asp | Glu | Ser | Arg | Arg 290 | Gly | Ala | Cys | Ser | Tyr 295 | Thr | Суѕ | Ser | Pro | Leu 300 |
| | | | Asp | 305 | | | _ | | 310 | | | | | 315 |
| Val | Ile | Asn | Phe | Arg 320 | Thr | Thr | Tyr | Val | Asn 325 | Thr | Asn | Asp | Glu | Val 330 |
| | | | Pro | 335 | | | | | 340 | _ | | _ | _ | 345 |
| | | | Asp | 350 | | | | | 355 | | _ | | | 360 |
| | _ | | Gly | 365 | _ | | | | 370 | | | _ | | 375 |
| _ | | | Arg | 380 | | | | | 385 | | | | | 390 |
| _ | | _ | Ser | 395 | _ | _ | | | 400 | | | | | 405 |
| _ | | | Pro | 410 | | | | _ | 415 | | | | _ | 420 |
| | | _ | Asn | 425 | | _ | | _ | 430 | | | _ | | 435 |
| - | | _ | Ser | 440 | _ | | | | 445 | _ | | | | 450 |
| | _ | | Ala | 455 | _ | | | | 460 | _ | _ | _ | | 465 |
| | | _ | Phe | 470 | | | | | 475 | | | _ | | 480 |
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| | | | Met | 515 | | _ | | _ | 520 | _ | | | _ | 525 |
| | | | Pro | 530 | _ | | _ | | 535 | | | | | 540 |
| | | | | 545 | | | | | 550 | | | | | 555 |
| | | | Tyr Glu | 560 | | | | | 565 | | | | | 570 |
| | | | Leu | 575 | | | | | 580 | | | | | 585 |
| | | | Arg | 590 | | | | | 595 | | | | | 600 |
| _ | _ | | Asp | 605 | | | | _ | 610 | _ | | | | 615 |
| | | | Ile | 620 | | | | | 625 | | | | | 630 |
| | | | Ala | 635 | | _ | | | 640 | | | | | 645 |
| | | | His | 650 | | _ | _ | | 655 | | | | | 660 |
| | | | Tyr | 665 | | | | | 670 | | | | | 675 |
| | | | Leu | 680 | | | | | 685 | | | | | 690 |
| | | | Glu | 695 | | _ | | | 700 | | | | | 705 |
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710
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Leu His Ser Ser Pro Arg Gln Ala Pro Gly Ser Gln Asp His Gln
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                                     730
Gly Phe Phe Leu Ser Asp Asn Gln Ser Asp Ala Ala Pro Pro Leu
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                                      745
                                                          750
Ser Ile Ser Asp Ala Phe Trp Leu Trp Pro Glu Leu Leu Gln Glu
                 755
                                     760
                                                          765
Met Pro Pro Lys His Ser Pro Gln Ser Pro Gln Glu Asp Pro Asp
                 770
                                     775
                                                          780
Cys Trp Pro Leu Lys Leu Gly Ser Arg Leu Glu Gln Leu Gln Ala
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                                                          795
Gln Met Asn Arg Leu Glu Ser Arg Val Ser Ser Asp Leu Ser Arg
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Ile Leu Gln Leu Leu Gln Lys Pro Met Pro Gln Gly His Ala Ser
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                                                          825
Tyr Ile Leu Glu Ala Pro Ala Ser Asn Asp Leu Ala Leu Val Pro
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                                     835
                                                          840
Ile Ala Ser Glu Thr Thr Ser Pro Gly Pro Arg Leu Pro Gln Gly
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                                     850
                                                          855
Phe Leu Pro Pro Ala Gln Thr Pro Ser Tyr Gly Asp Leu Asp Asp
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                                                          870
Cys Ser Pro Lys His Arg Asn Ser Ser Pro Arg Met Pro His Leu
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                                     880
Ala Val Ala Met Asp Lys Thr Leu Ala Pro Ser Ser Glu Gln Glu
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                                                          900
Gln Pro Glu Gly Leu Trp Pro Pro Leu Ala Ser Pro Leu His Pro
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                                     910
                                                          915
Leu Glu Val Gln Gly Leu Ile Cys Gly Pro Cys Phe Ser Ser
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                                     925
                                                          930
Pro Glu His Leu Gly Ser Val Pro Lys Gln Leu Asp Phe Gln Arg
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Leu Leu Gly Ala Leu Pro Ser Val Ser Cys Gly Gly Trp Gly His
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Arg Gly Arg Gln Thr Tyr Gly Arg Ala Cys Gly Val Lys Glu Lys
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Pro Phe Ser Leu Leu Gly Pro Gln Ile Thr Val Tyr Ala Val
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                                                           75
Pro Gln Ser Glu Gly Pro Gln Glu Gly Arg Leu Arg Val Asn Ser
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                                      85
Ala Cys Leu Pro Pro Glu Arg Gly Leu Thr Asn Ala Cys Thr Asn
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                                     100
His Glu Glu Leu Ser Leu Asp Cys Leu Leu Phe Glu Asn Val Asn
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Thr Leu Thr Leu Asp Phe Cys Leu Trp Glu Lys Thr Thr Ile Val
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                                     130
                                                          135
Pro Gly Val Leu Pro Tyr Ala Gly Leu Thr Leu Gln Ser Lys Phe
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                                     145
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Leu Leu Gly Arg Ala Leu Leu Ala Gly Val His Val Ile Thr Leu
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                                     160
Thr Pro Glu Arg Val Thr His His Val His Gly Trp Tyr Met Glu
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| Asp | G1y | Phe | Lys | Gly 185 | Asp | Arg | Thr | Glu | Gly 190 | Суѕ | Arg | Ser | Asp | <i>Ser</i> 195 |
| Val | Ala | Val | Pro | Ala 200 | Ala | Ala | Pro | Val | Cys 205 | Gln | Pro | Lys | Ser | Ala 210 |
| Thr | Asn | Gly | Gln | Pro 215 | Pro | Ala | Pro | Ala | Pro 220 | Thr | Pro | Thr | Pro | Arg 225 |
| Leu | Ser | Ile | Ser | Ser 230 | Arg | Ala | Thr | Val | Val 235 | Ala | Arg | Met | Glu | Gly 240 |
| Thr | Ser | Gln | Gly | Gly 245 | Leu | Gln | Thr | Val | Met 250 | Lys | Trp | Lys | Thr | Val 255 |
| Val | Ala | Ile | Phe | Val 260 | Val | Val | Val | Val | Tyr 265 | Leu | Val | Thr | Gly | Gly 270 |
| Leu | Val | Phe | Arg | Ala 275 | Leu | Glu | Gln | Pro | Phe 280 | Glu | Ser | Ser | Gln | Lys 285 |
| | | | | Leu 290 | | | | | 295 | | | | | 300 |
| _ | | | | Gln 305 | | | | | 310 | | | | | 315 |
| | | | | Ala 320 | | | | | 325 | | | | | 330 |
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| | | | | Phe 365 | | | | | 370 | | | | | 375 |
| | | | | Leu 380 | | | | | 385 | | | | | 390 |
| | | _ | _ | Ser 395 | | | | | 400 | | | | | 405 |
| _ | | | | Gln 410 | | | | | 415 | | | | | 420 |
| | | | | Gly 425 | | | | | 430 | | Ile | | | 435 |
| | | | | Ile 440 | | | | | 445 | | | | | 450 |
| | | | | Thr 455 | | | | | 460 | | | | | 465 |
| | | | | Phe 470 | | | | | 475 | | | | | 480 |
| | | | | Leu 485 | | | | | 490 | | | | | 495 |
| _ | | | | Val 500 | | | | | 505 | | | | | Val 510 |
| | | | _ | Thr 515 | | | | | 520 | | | | | 525 |
| | | | | Lys 530 | | | | | 535 | | | | | 540 |
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| | | | | Ser 560 | | | | | 565 | | | | | 570 |
| | | | | Leu 575 | | | | | 580 | | | | | 585 |
| | | | | Asp 590 | | | | | 595 | | | | | 600 |
| | | | | Arg 605 | | | | | 610 | | | | | 615 |
| | | | | His 620 | | | | | 625 | | | | | 630 |
| | | | | Ser 635 | | | | | 640 | | | | | 645 |
| | | | | Thr 650 | | | | | 655 | | | | | 660 |
| Thr | Phe | Arg | Asn | Tyr 665 | Ser | Leu | Asp | Glu | Glu 670 | | Lys | Glu | Glu | Glu 675 |
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Thr Glu Lys Met Cys Asn Ser Asp Asn Ser Ser Thr Ala Met Leu
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Thr Asp Cys Ile Gln Gln His Ala Glu Leu Glu Asn Gly Met
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Pro Thr Asp Thr Lys Asp Arg Glu Pro Glu Asn Asn Ser Leu Leu
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Ala Leu Ala Ala Gly Thr Gly Lys Ile Gly Asn Arg His Asp Met
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Leu Leu Val Glu Pro Leu Asn Arg Leu Leu Gln Asp Lys Trp Asp
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Arg Phe Val Lys Arg Ile Phe Tyr Phe Asn Phe Leu Val Tyr
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                                      70
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Leu Tyr Met Ile Ile Phe Thr Met Ala Ala Tyr Tyr Arg Pro Val
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Asp Gly Leu Pro Pro Phe Lys Met Glu Lys Thr Gly Asp Tyr Phe
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                                     100
                                                          105
Arg Val Thr Gly Glu Ile Leu Ser Val Leu Gly Gly Val Tyr Phe
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                                     115
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Phe Phe Arg Gly Ile Gln Tyr Phe Leu Gln Arg Arg Pro Ser Met
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Lys Thr Leu Phe Val Asp Ser Tyr Ser Glu Met Leu Leu Phe Leu
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Gln Ser Leu Phe Met Leu Ala Thr Val Val Leu Tyr Phe Ser His
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Leu Lys Glu Tyr Val Ala Ser Met Val Phe Ser Leu Ala Leu Gly
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Trp Thr Asn Met Leu Tyr Tyr Thr Arg Gly Phe Gln Gln Met Gly
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                                      190
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Ile Tyr Ala Val Met Ile Glu Lys Met Ile Leu Arg Asp Leu Cys
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Arg Phe Met Phe Val Tyr Ile Val Phe Leu Phe Gly Phe Ser Thr
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Ala Val Val Thr Leu Ile Glu Asp Gly Lys Asn Asp Ser Leu Pro
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Ser Glu Ser Thr Ser His Arg Trp Arg Gly Pro Ala Xaa Arg Pro
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Asn Ser Ser Tyr Asn Ser Leu Tyr Ser Thr Cys Leu Glu Leu Phe
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Lys Phe Thr Ile Gly Met Gly Asp Leu Glu Phe Thr Glu Asn Tyr
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Asp Phe Lys Ala Val Phe Ile Ile Leu Leu Leu Ala Tyr Val Ile
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Leu Thr Tyr Ile Val Leu Leu Leu Asn Met Leu Ile Ala Leu Met
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Gly Glu Thr Val Glu Asn Val Ser Lys Glu Ser Glu Arg Ile Trp
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Arg Leu Gln Arg Ala Ile Thr Ile Leu Asp Thr Glu Lys Ser Phe
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Leu Lys Cys Met Arg Lys Ala Phe Arg Ser Gly Lys Leu Leu Gln
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Val Gly Tyr Thr Pro Asp Gly Lys Asp Asp Tyr Arg Trp Cys Phe
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                                     370
Val Asp Glu Val Asn Trp Thr Thr Trp Asn Thr Asn Val Gly Ile
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                                     385
                                                          390
Ile Asn Glu Asp Pro Gly Asn Cys Glu Gly Val Lys Arg Thr Leu
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Ser Phe Ser Leu Arg Ser Ser Arg Val Ser Gly Arg His Trp
                                                         Lys
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Asn Phe Ala Leu Val Pro Leu Leu Arg Glu Ala Ser Ala Arg Asp
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Arg Gln Ser Ala Gln Pro Glu Glu Val Tyr Leu Arg Gln Phe Ser
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Lys Glu Arg Lys Lys Ala Thr Ser Arg Glu Phe Leu Val Gly Gly
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Arg Gln Met Ser Phe Gly Pro Val Gly Leu Ser Leu Thr Ala Ser
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Phe Met Ser Ala Val Thr Val Leu Gly Thr Pro Ser Glu Val
                                                          Tyr
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                  65
Arg Phe Gly Ala Ser Phe Leu Val Phe Phe Ile Ala Tyr Leu Phe
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                                      85
Val Ile Leu Leu Thr Ser Glu Leu Phe Leu Pro Val Phe Tyr Arg
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Ser Gly Ile Thr Ser Thr Tyr Glu Tyr Leu Gln Leu Arg Phe Asn
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Lys Pro Val Arg Tyr Ala Ala Thr Val Ile Tyr Ile Val Gln Thr
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                                      130
                                                          135
Ile Leu Tyr Thr Gly Val Val Val Tyr Ala Pro Ala Leu Ala Leu
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Asn Gln Val Thr Gly Phe Asp Leu Trp Gly Ser Val Phe Ala Thr
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Gly Ile Val Cys Thr Phe Tyr Cys Thr Leu Gly Gly Leu Lys Ala
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Val Val Trp Thr Asp Ala Phe Gln Met Val Val Met Ile Val Gly
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Phe Leu Thr Val Leu Ile Gln Gly Ser Thr His Ala Gly Gly Phe
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His Asn Val Leu Glu Gln Ser Thr Asn Gly Ser Arg Leu His Ile
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Phe Asp Phe Asp Val Asp Pro Leu Arg Arg His Thr Phe Trp Thr
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Ile Leu Val Cys Ala Val Phe Ser Gly Leu Ile Met Tyr Ser His
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Phe Lys Asp Cys Asp Pro Trp Thr Ser Gly Ile Ile Ser Ala Pro
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                                                           45
Phe Arg Phe Leu His Glu Thr Gly Gly Ala Met Val Tyr Gly Leu
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| Ser | Thr | Leu | Leu | Val 95 | Asn | Ile | Thr | Asp | Gln 100 | Val | Tyr | Glu | Tyr | Lys 105 |
| Tyr | Lys | Arg | Glu | | Ser | Gln | His | Asn | | Asn | Pro | His | Gln | Gly 120 |
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| Lys | Lys | Arg | His | | Phe | Gln | Asn | Leu | | Ser | Ile | Leu | Thr | |
| Ala | Phe | Leu | Gly | | Ala | Ile | Ser | Cys | | Val | Ile | Gly | Leu | |
| Met | Tyr | Gly | Phe | | Lys | Ala | Met | Ile | | Ala | Gly | Gln | Leu | |
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| Leu | His | Val | Asp | | Asp | Leu | Tyr | Thr | | Leu | Phe | Gly | Glu | |
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| Phe | Ala | Met | Gly | | Ala | Tyr | Ala | Ile | Ile 295 | Thr | Ala | Leu | Leu | Thr |
| Lys | Phe | Thr | Lys | Leu 305 | Cys | Glu | Phe | Pro | Met 310 | Leu | Glu | Thr | Gly | Leu 315 |
| Phe | Phe | Leu | Leu | Ser 320 | Trp | Ser | Ala | Phe | Leu 325 | Ser | Ala | Glu | Ala | Ala 330 |
| Gly | Leu | Thr | Gly | Ile 335 | Val | Ala | Val | Leu | Phe 340 | Cys | Gly | Val | Thr | Gln 345 |
| Ala | His | Tyr | Thr | Tyr 350 | Asn | Asn | Leu | Ser | Ser 355 | Asp | Ser | Lys | Ile | Arg 360 |
| Thr | Lys | Gln | Leu | Phe 365 | Glu | Phe | Met | Asn | Phe 370 | Leu | Ala | Glu | Asn | Val 375 |
| Ile | Phe | Cys | Tyr | Met 380 | Gly | Leu | Ala | Leu | Phe 385 | Thr | Phe | Gln | Asn | His 390 |
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| Val | Ala | Arg | Ala | Cys 410 | Asn | Ile | Tyr | Pro | Leu 415 | ser | Phe ' | Leu | Leu | Asn 420 |
| Leu | Gly | Arg | Lys | Gln 425 | Lys | Ile | Pro | Trp | Asn 430 | Phe | Gln | His | Met | Met 435 |
| Met | Phe | Ser | Gly | Leu 440 | Arg | Gly | Ala | Ile | Ala 445 | Phe | Ala | Leu | Ala | Ile 450 |
| Arg | Asn | Thr | Glu | | Gln | Pro | Lys | Gln | | Met | Phe | Thr | Thr | Thr 465 |
| Leu | Leu | Leu | Val | | Phe | Thr | Val | Trp | | Phe | Gly | Gly | Gly | |
| Thr | Pro | Met | Leu | | Trp | Leu | Gln | Ile | | Val | Gly | Val | Asp | |
| Asp | Glu | Asn | Leu | | Glu | Asp | Pro | Ser | | Gln | His | Gln | Glu | |
| Asn | Asn | Leu | Asp | | Asn | Met | Thr | Lys | | Glu | Ser | Ala | Arg | |
| Phe | Arg | Met | Trp | | Ser | Phe | Asp | His | | Tyr | Leu | Lys | Pro | |
| Leu | Thr | His | Ser | | Pro | Pro | Leu | Thr | | Thr | Leu | Pro | Glu | |

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Glu Gln Leu Lys Glu Asp Asp Val Glu Cys Ile Val Asn Gln Asp
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Glu Leu Ala Ile Asn Tyr Gln Glu Gln Ala Ser Ser Pro Cys Ser
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Pro Pro Ala Arg Leu Gly Leu Asp Gln Lys Ala Ser Pro Gln Thr
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Pro Leu Tyr Trp Met Pro Glu Trp Arg Val Lys Ala Thr Cys Val
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Arg Val Ser Val Cys Arg Val Asn Glu Glu Ile Glu Glu Ile Phe
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Ser Thr Asp Leu Val Pro Gly Asp Val Met Val Ile Pro Leu Asn
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                                     295
Gly Thr Ile Met Pro Cys Asp Ala Val Leu Ile Asn Gly Thr Cys
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                                                          315
                                     310
Ile Val Asn Glu Ser Met Leu Thr Gly Glu Ser Val Pro Val Thr
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| Asp | Glu | Leu | Tyr | | Pro | Glu | Thr | His | | Arg | His | Thr | Leu | |
| Cys | Gly | Thr | Thr | | Ile | Gln | Thr | Arg | | Tyr | Thr | Gly | Glu | |
| Val | Lys | Ala | Ile | | Val | Arg | Thr | Gly | | Ser | Thr | Ser | Lys | Gly 390 |
| Gln | Leu | Val | Arg | _ | Ile | Leu | Tyr | Pro | Lys 400 | Pro | Thr | Asp | Phe | Lys 405 |
| Leu | Tyr | Arg | Asp | | Tyr | Leu | Phe | Leu | Leu 415 | Cys | Leu | Val | Ala | Val 420 |
| Ala | Gly | Ile | Gly | | Ile | Tyr | Thr | Ile | Ile 430 | Asn | Ser | Ile | Leu | Asn 435 |
| Glu | Val | Gln | Val | Gly 440 | Val | Ile | Ile | Ile | Glu 445 | Ser | Leu | Asp | Ile | Ile 450 |
| Thr | Ile | Thr | Val | Pro 455 | Pro | Ala | Leu | Pro | Ala 460 | Ala | Met | Thr | Ala | Gly 465 |
| Ile | Val | Tyr | Ala | Gln 470 | Arg | Arg | Leu | Lys | Lys 475 | Ile | Gly | Ile | Phe | Cys 480 |
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| Cys | Phe | Asp | ГÀг | Thr 500 | Gly | Thr | Leu | Thr | Glu 505 | Asp | Gly | Leu | Asp | Leu 510 |
| Trp | Gly | Ile | Gln | Arg 515 | Val | Glu | Asn | Ala | Arg 520 | Phe | Leu | Ser | Pro | Glu 525 |
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| Ser | Gly | Asp | Pro | Leu 560 | Asp | Leu | Lys | Met | Phe 565 | Glu | Ala | Ile | Gly | Trp 570 |
| Ile | Leu | Glu | Glu | Ala 575 | Thr | Glu | Glu | Glu | Thr 580 | Ala | Leu | His | Asn | Arg 585 |
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| | | - | | 620 | | | Val | | 625 | | | | | Ser 630 |
| | | | _ | 635 | | | Va1 | | 640 | | | _ | _ | 645 |
| _ | | _ | | 650 | | _ | Gly | | 655 | | | | | 660 |
| | _ | _ | | 665 | | | Pro | | 670 | | | | | 675 |
| | | | | 680 | | | Phe | | 685 | | | | | 690 |
| | | | | 695 | | | | | 700 | | | | | Ile 705 |
| | _ | | | 710 | | | Asn | | 715 | | | | | 720 |
| | | | | 725 | | | | | 730 | | | | | Glu 735 |
| Asp | Leu | His | Lys | Ala 740 | Asn | Ile | Arg | Thr | Val 745 | Met | Val | Thr | | 750 |
| Ser | Met | Leu | Thr | Ala 755 | Val | Ser | Val | Ala | Arg 760 | Asp | Cys | Gly | Met | Ile 765 |
| Leu | Pro | Gln | Asp | Lys 770 | Val | Ile | Ile | Ala | Glu 775 | Ala | Leu | Pro | Pro | Lys 780 |
| Asp | Gly | Lys | Val | Ala 785 | Lys | Ile | Asn | Trp | His 790 | Tyr | Ala | Asp | Ser | Leu 795 |
| | | - | | 800 | | | Ala | | 805 | | | | | 810 |
| Val | Lys | Leu | Val | His 815 | ĄsĄ | ser | Leu | Glu | Asp 820 | Leu | Gln | Met | Thr | Arg 825 |
| Tyr | His | Phe | Ala | Met | Asn | Gly | Lys | Ser | Phe | Ser | Val | Ile | Leu | Glu |

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Leu Gln Asn Val Asp Tyr Phe Val Gly Met Cys Gly Asp Gly Ala
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                                                          885
Asn Asp Cys Gly Ala Leu Lys Arg Ala His Gly Gly Ile Ser Leu
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                                     895
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Ser Glu Leu Glu Ala Ser Val Ala Ser Pro Phe Thr Ser Lys Thr
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Pro Ser Ile Ser Cys Val Pro Asn Leu Ile Arg Glu Gly Arg Ala
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Ser Ile Ile Gln Tyr
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Val Trp His Pro Lys Ser Asp Ala Cys Asn Thr Thr Gly Ser Gly
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His Asn Ile Gln Asn Tyr Glu Asn Thr Thr Val Phe Phe Ile Ser
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Ser Phe Gln Tyr Leu Ile Val Ala Ile Ala Phe Ser Lys Gly Lys
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Cys Leu Pro Trp Ala Leu Gly Cys Arg Lys Lys Thr Pro Lys Ala
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Ala Leu Tyr Thr Ile Ala Gly Gly Leu Ala Ala Val Ile Tyr
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Arg Glu Pro Pro Pro Gly Ala Glu Ala Tyr Ile Pro Gln Arg Tyr
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Pro Asp Asn Arg Ile Val Ser Ser Lys Tyr Thr Phe Trp Asn Phe
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Ile Pro Lys Asn Leu Phe Glu Gln Phe Arg Arg Val Ala Asn Phe
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Tyr Phe Leu Ile Ile Phe Leu Val Gln Leu Ile Ile Asp Thr Pro
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                                      85
Thr Ser Pro Val Thr Ser Gly Leu Pro Leu Phe Phe Val Ile Thr
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Val Thr Ala Ile Lys Gln Gly Tyr Glu Asp Trp Leu Arg His Lys
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Gly Lys Leu Val Arg Lys Gln Ser Arg Lys Leu Arg Val Gly Asp
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Ile Val Met Val Lys Glu Asp Glu Thr Phe Pro Cys Asp Leu Ile
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Phe Leu Ser Ser Asn Arg Gly Asp Gly Thr Cys His Val Thr
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Ala Ser Leu Asp Gly Glu Ser Ser His Lys Thr His Tyr Ala Val
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Gln Asp Thr Lys Gly Phe His Thr Glu Glu Asp Ile Gly Gly Leu
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                                     205
His Ala Thr Ile Glu Cys Glu Gln Pro Gln Pro Asp Leu Tyr Lys
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| Val | Arg | Pro | Leu | | Ser | Glu | Asn | Leu | | Leu | Arg | Gly | Ala | _ |
| Leu | Lys | Asn | Thr | | Lys | Ile | Phe | Gly | | Ala | Ile | Tyr | Thr | |
| Met | Glu | Thr | Lys | | Ala | Leu | Asn | Tyr | | Ser | Lys | Ser | Gln | |
| Arg | Ser | Ala | Val | | Lys | Ser | Met | Asn | Ala 295 | Phe | Leu | Ile | Val | |
| Leu | Cys | Ile | Leu | | Ser | Lys | Ala | Leu | | Asn | Thr | Val | Leu | |
| Tyr | Val | Trp | Gln | | Glu | Pro | Phe | Arg | | Glu | Pro | Trp | Tyr | |
| Gln | Lys | Thr | Glu | Ser 335 | Glu | Arg | Gln | Arg | | Leu | Phe | Leu | Lys | |
| Phe | Thr | Asp | Phe | Leu 350 | Ala | Phe | Met | Val | Leu 355 | Phe | Asn | Tyr | Ile | Ile 360 |
| Pro | Val | Ser | Met | Tyr 365 | Val | Thr | Val | Glu | Met 370 | Gln | Lys | Phe | Leu | Gly 375 |
| Ser | Tyr | Phe | Ile | Thr 380 | Trp | Asp | Glu | Asp | Met 385 | Phe | Asp | Glu | Glu | Thr 390 |
| Gly | Glu | Gly | Pro | Leu 395 | Val | Asn | Thr | Ser | Asp 400 | Leu | Asn | Glu | Glu | Leu 405 |
| Gly | Gln | Val | Glu | Tyr 410 | Ile | Phe | Thr | Asp | Lys 415 | Thr | Gly | Thr | Leu | Thr 420 |
| Glu | Asn | Asn | Met | Glu 425 | Phe | Lys | Glu | Суѕ | Cys 430 | Ile | Glu | Gly | His | Val 435 |
| Tyr | Val | Pro | His | Val 440 | Ile | Суз | Asn | Gly | Gln 445 | Val | Leu | Pro | Glu | ser 450 |
| Ser | Gly | Ile | Asp | Met 455 | Ile | Asp | Ser | Ser | Pro 460 | Ser | Val | Asn | Gly | Arg 465 |
| Glu | Arg | Glu | Glu | Leu 470 | Phe | Phe | Arg | Ala | Leu 475 | Cys | Leu | Суѕ | His | Thr 480 |
| Val | Gln | Val | Lys | Asp 485 | Asp | Asp | Ser | Val | Asp 490 | Gly | Pro | Arg | Lys | Ser 495 |
| Pro | Asp | Gly | Gly | Lys 500 | Ser | Суѕ | Val | Tyr | Ile 505 | Ser | Ser | Ser | Pro | Asp 510 |
| Glu | ۷al | Ala | Leu | Val 515 | Glu | Gly | Va1 | Gln | Arg 520 | Leu | Gly | Phe | Thr | Tyr 525 |
| Leu | Arg | Leu | Lys | Asp 530 | Asn | Tyr | Met | Glu | Ile 535 | Leu | Asn | Arg | Glu | Asn 540 |
| | Ile | | _ | 545 | | | | | 550 | | | | - | 555 |
| Val | Arg | Arg | Arg | Met 560 | Ser | Val | Ile | Val | Lys 565 | Ser | Ala | Thr | Gly | Glu 570 |
| Ile | Tyr | Leu | Phe | Cys 575 | Lys | Gly | Ala | Asp | Ser 580 | Ser | Ile | Phe | Pro | Arg 585 |
| Val | Ile | Glu | Gly | Lys 590 | Val | Asp | Gln | Ile | Arg 595 | Ala | Arg | Val | Glu | Arg 600 |
| Asn | Ala | Val | Glu | Gly 605 | Leu | Arg | Thr | Leu | Cys 610 | Val | Ala | Tyr | Lys | Arg 615 |
| Leu | Ile | Gln | Glu | Glu 620 | Tyr | Glu | Gly | Ile | Cys 625 | Lys | Leu | Leu | Gln | Ala 630 |
| Ala | Lys | Val | Ala | Leu 635 | Gln | Asp | Arg | Glu | Lys 640 | ГЛS | Leu | Ala | Glu | Ala 645 |
| Tyr | Glu | Gln | Ile | Glu 650 | ГÀЗ | Asp | Leu | Thr | Leu 655 | Leu | Gly | Ala | Thr | |
| Val | Glu | Asp | Arg | Leu 665 | Gln | Glu | Lys | Ala | Ala 670 | Asp | Thr | Ile | Glu | Ala 675 |
| Leu | Gln | Lys | Ala | | Ile | Lys | Val | Trp | | Leu | Thr | Gly | Asp | |
| Met | Glu | Thr | Ala | | Ala | Thr | Cys | Tyr | | Cys | Lys | Leu | Phe | Arg 705 |
| Arg | Asn | Thr | Gln | | Leu | Glu | Leu | Thr | | Lys | Arg | Ile | Glu | Glu 720 |
| Gln | Ser | Leu | His | | Val | Leu | Phe | Glu | _ | Ser | Lys | Thr | Val | Leu |

| Arg | His | Ser | Gly | 725 Ser 740 | Leu | Thr | Arg | Asp | 730 Asn 745 | Leu | Ser | Gly | Leu | 735 Ser 750 |
|-----|-----|-----|-----|-------------------|-----|-----|-----|-----|-------------------|-----|-----|-----|-----|-------------------|
| Ala | Asp | Met | Gln | | Tyr | Gly | Leu | Ile | | Asp | Gly | Ala | Ala | |
| Ser | Leu | Ile | Met | | Pro | Arg | Glu | Asp | | Ser | Ser | Gly | Asn | |
| Arg | Glu | Leu | Phe | | Glu | Ile | Cys | Arg | | Cys | Ser | Ala | Val | |
| Cys | Cys | Arg | Met | | Pro | Leu | Gln | Lys | | Gln | Ile | Val | Lys | |
| Ile | Lys | Phe | Ser | | Glu | His | Pro | Ile | | Leu | Ala | Ile | Gly | |
| Gly | Ala | Asn | Asp | Val 830 | Ser | Met | Ile | Leu | Glu 835 | Ala | His | Val | Gly | Ile 840 |
| Gly | Val | Ile | Gly | Lys 845 | Glu | Gly | Arg | Gln | Ala 850 | Ala | Arg | Asn | Ser | Asp 855 |
| Tyr | Ala | Ile | Pro | Lys 860 | Phe | Lys | His | Leu | Lys 865 | | Met | Leu | Leu | Val 870 |
| His | Gly | His | Phe | Tyr 875 | Tyr | Ile | Arg | Ile | Ser 880 | Glu | Leu | Val | Gln | Tyr 885 |
| Phe | Phe | Tyr | Lys | Asn 890 | Val | Cys | Phe | Ile | Phe 895 | Pro | Gln | Phe | Leu | Tyr 900 |
| Gln | Phe | Phe | Cys | Gly 905 | Phe | Ser | Gln | Gln | Thr 910 | Leu | Tyr | Asp | Thr | Ala 915 |
| Tyr | Leu | Thr | Leu | Tyr 920 | Asn | Ile | Ser | Phe | Thr 925 | Ser | Leu | Pro | Ile | Leu 930 |
| | _ | | | 935 | | | His | | 940 | | | | | 945 |
| | | | | 950 | | | Asp | | 955 | | | | | 960 |
| _ | _ | _ | | 965 | | _ | Trp | | 970 | | _ | | | 975 |
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| | | | | 995 | _ | | Ile | : | 1000 | | _ | | | 1005 |
| | | | | 1010 | | | Val | | 1015 | | | | | L020 |
| | | _ | | 1025 | _ | _ | Thr | _ : | 1030 | | | | 3 | L035 |
| | _ | | | 1040 | | _ | Val | : | 1045 | | | | | L050 |
| _ | | | | 1055 | | | Asn | | 1060 | _ | | | _ : | 1065 |
| | | | | 1070 | | | Gly | | 1075 | _ | | | - | 1080 |
| | | | | 1085 | | | Leu | : | 1090 | | | _ | | 1095 |
| | | | | 1100 | | | Thr | | 1105 | | | | | 1110 |
| | | | | 1115 | | | Asp | : | 1120 | | | | : | 1125 |
| | | | | 1130 | | | Gly | | 1135 | | | | | 1140 |
| | | | | 1145 | | | Ser | | 1150 | | | | | 1155 |
| _ | _ | | _ | Val 1160 | | | Glu | : | 1165 | | | | | 1170 |
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Met Leu Val Gly Ala Leu Cys Ala Leu Ala Gly Val Leu Thr Ile
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                                     460
Ala Met Pro Val Pro Val Ile Val Asn Asn Phe Gly Met Tyr Tyr
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                                                          480
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Ser Leu Ala Met Ala Lys Gln Lys Leu Pro Arg Lys Arg Lys
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                                     490
His Ile Pro Pro Ala Pro Gln Ala Ser Ser Pro Thr Phe Cys Lys
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Thr Glu Leu Asn Met Ala Cys Asn Ser Thr Gln Ser Asp Thr
                                                         Cys
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Leu Gly Lys Asp Asn Arg Leu Leu Glu His Asn Arg Ser Val Leu
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Ser Gly Asp Asp Ser Thr Gly Ser Glu Pro Pro Leu Ser Pro Pro
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Glu Arg Leu Pro Ile Arg Arg Ser Ser Thr Arg Asp Lys Asn Arg
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Arg Gly Glu Thr Cys Phe Leu Leu Thr Thr Gly Asp Tyr Thr Cys
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Ala Ser Asp Gly Gly Ile Arg Lys Gly Tyr Glu Lys Ser Arg
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Val Phe Val Ile Ala Val Gly Ile Trp Ser Ser Ile Arg Ala Ser
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Arg Gly Thr Ile Gly Gly Tyr Phe Leu Ala Gly Arg Ser Met Ser
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Trp Trp Pro Ile Gly Ala Ser Leu Met Ser Ser Asn Val Gly Ser
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                                      85
Gly Leu Phe Ile Gly Leu Ala Gly Thr Gly Ala Ala Gly Gly Leu
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Ala Val Gly Gly Phe Glu Trp Asn Ala Thr Trp Leu Leu Leu Ala
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Leu Gly Trp Val Phe Val Pro Val Tyr Ile Ala Ala Gly Val Val
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                                     130
Thr Met Pro Gln Tyr Leu Lys Lys Arg Phe Gly Gly Gln Arg Ile
                140
                                     145
                                                          150
Gln Val Tyr Met Ser Val Leu Ser Leu Ile Leu Tyr Ile Phe Thr
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                                     160
                                                          165
Lys Ile Ser Thr Asp Ile Phe Ser Gly Ala Leu Phe Ile Gln Met
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                                     175
Ala Leu Gly Trp Asn Leu Tyr Leu Ser Thr Gly Ile Leu Leu Val
                                                          195
                185
                                     190
Val Thr Ala Val Tyr Thr Ile Ala Gly Gly Leu Met Ala Val Ile
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                                                          210
Tyr Thr Asp Ala Leu Gln Thr Val Ile Met Val Gly Gly Ala Leu
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Val Leu Met Phe Leu Gly Phe Gln Asp Val Gly Trp Tyr Pro Gly
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Leu Glu Gln Arg Tyr Arg Gln Ala Ile Pro Asn Val Thr Val Pro
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                                     250
                                                          255
Asn Thr Thr Cys His Leu Pro Arg Pro Asp Ala Phe His Ile Leu
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                                     265
                                                          270
Arg Asp Pro Val Ser Gly Asp Ile Pro Trp Pro Gly Leu Ile Phe
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Gly Leu Thr Val Leu Ala Thr Trp Cys Trp Cys Thr Asp Gln Val
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                                     295
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Ile Val Gln Arg Ser Leu Ser Ala Lys Ser Leu Ser His Ala Lys
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                                     310
                                                          315
Gly Gly Ser Val Leu Gly Gly Tyr Leu Lys Ile Leu Pro Met Phe
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                                     325
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Phe Ile Val Met Pro Gly Met Ile Ser Arg Ala Leu Phe Pro Asp
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                                     340
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Glu Val Gly Cys Val Asp Pro Asp Val Cys Gln Arg Ile Cys Gly
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Ala Arg Val Gly Cys Ser Asn Ile Ala Tyr Pro Lys Leu Val Met
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Ala Leu Met Pro Val Gly Leu Arg Gly Leu Met Ile Ala Val
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Met Ala Ala Leu Met Ser Ser Leu Thr Ser Ile Phe Asn Ser Ser
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Ser Thr Leu Phe Thr Ile Asp Val Trp Gln Arg Phe Arg Arg Lys
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Ser Thr Glu Gln Glu Leu Met Val Val Gly Arg Val Phe Val Val
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Phe Leu Val Val Ile Ser Ile Leu Trp Ile Pro Ile Ile Gln Ser
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Ser Asn Ser Gly Gln Leu Phe Asp Tyr Ile Gln Ala Val Thr Ser
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                                     460
                                                          465
Tyr Leu Ala Pro Pro Ile Thr Ala Leu Phe Leu Leu Ala Ile Phe
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                                     475
                                                          480
Cys Lys Arg Val Thr Glu Pro Gly Ala Phe Trp Gly Leu Val Phe
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                                     490
                                                          495
Gly Leu Gly Val Gly Leu Leu Arg Met Ile Leu Glu Phe Ser
                                                          Tyr
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                                     505
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Pro Ala Pro Ala Cys Gly Glu Val Asp Arg Arg Pro Ala Val Leu
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                                     520
                                                          525
Lys Asp Phe His Tyr Leu Tyr Phe Ala Ile Leu Leu Cys Gly Leu
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Thr Ala Ile Val Ile Val Ile Val Ser Leu Cys Thr Thr Pro Ile
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Pro Glu Glu Gln Leu Thr Arg Leu Thr Trp Trp Thr Arg Asn Cys
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Pro Leu Ser Glu Leu Glu Lys Glu Ala His Glu Ser Thr Pro Glu
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Ile Ser Glu Arg Pro Ala Gly Glu Cys Pro Ala Gly Gly Ala
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Ala Glu Asn Ser Ser Leu Gly Gln Glu Gln Pro Glu Ala Pro Ser
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Arg Ser Trp Gly Lys Leu Leu Trp Ser Trp Phe Cys Gly Leu Ser
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Gly Thr Pro Glu Gln Ala Leu Ser Pro Ala Glu Lys Ala Ala Leu
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                                     640
                                                          645
Glu Gln Lys Leu Thr Ser Ile Glu Glu Glu Pro Leu Trp Arg His
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Val Cys Asn Ile Asn Ala Val Leu Leu Leu Ala Ile Asn Ile Phe
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| Lys | Ala | Leu | Val | Glu 470 | Ala | Ala | Ala | Arg | Ile 475 | Gly | Ile | Val | Phe | Ile 480 |
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| Glu | Arg | Tyr | Lys | | Leu | His | Ile | Leu | | Phe | Asp | Ser | Asp | |
| Arg | Arg | Met | Ser | _ | Ile | Val | Gln | Ala | | Ser | Gly | Glu | Lys | |
| Leu | Phe | Ala | Lys | | Ala | Glu | Ser | Ser | | Leu | Pro | Lys | Cys | |
| Gly | Gly | Glu | Ile | | Lys | Thr | Arg | Ile | | Val | Asp | Glu | Phe | |
| Leu | Lys | Gly | Leu | | Thr | Leu | Cys | Ile | | Tyr | Arg | Lys | Phe | |
| ser | Lys | Glu | Tyr | | Glu | Ile | Asp | Lys | | Ile | Phe | Glu | Ala | |
| Thr | Ala | Leu | Gln | | Arg | Glu | Glu | Lys | | Ala | Ala | Val | Phe | |
| Phe | Ile | Glu | Lys | | Leu | Ile | Leu | Leu | | Ala | Thr | Ala | Val | |
| Asp | Arg | Leu | Gln | | Lys | Val | Arg | Glu | | Ile | Glu | Ala | Leu | |
| Met | Ala | Gly | Ile | | Val | Trp | Val | Leu | | Gly | Asp | Lys | His | |
| Thr | Ala | Val | Ser | | ser | Leu | Ser | Cys | | His | Phe | His | Arg | |
| Met | Asn | Ile | Leu | | Leu | Ile | Asn | Gln | | Ser | Asp | Ser | Glu | |
| Ala | Glu | Gln | Leu | | Gln | Leu | Ala | Arg | - • - | Ile | Thr | Glu | Asp | |
| Val | Ile | Gln | His | | Leu | Val | Val | Asp | | Thr | Ser | Leu | Ser | |
| Ala | Leu | Arg | Glu | | Glu | Lys | Leu | Phe | | Glu | Val | Суз | Arg | |
| Cys | ser | Ala | Val | | Cys | Cys | Arg | Met | | Pro | Leu | Gln | Lys | |
| Lys | Val | Ile | Arg | | Ile | Lys | Ile | Ser | | Glu | Lys | Pro | Ile | |
| Leu | Ala | Val | Gly | | Gly | Ala | Asn | Asp | | Ser | Met | Ile | Gln | |
| Ala | His | Val | Gly | | Gly | Ile | Met | Gly | | Glu | Gly | Arg | Gln | |
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| Thr | Leu | Val | Gln | | Phe | Phe | Tyr | Lys | | Val | Cys | Phe | Ile | |
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| Ser | Leu | Pro | Ile | | Ile | Tyr | Ser | Leu | | Glu | Gln | His | Val | |
| Pro | His | Val | Leu | | Asn | Lys | Pro | Thr | | Tyr | Arg | Asp | Ile | |
| Lys | Asn | Arg | Leu | | Ser | Ile | Гуз | Thr | | Leu | Tyr | Trp | Thr | Ile 900 |
| Leu | Gly | Phe | Ser | | Ala | Phe | Ile | Phe | | Phe | Gly | Ser | Tyr | |
| Leu | Ile | Gly | Lys | | Thr | Ser | Leu | Leu | | Asn | Gly | Gln | Met | |
| Gly | Asn | Trp | Thr | | Gly | Thr | Leu | Val | | Thr | Val | Met | Val | |
| Thr | Val | Thr | Val | | Met | Ala | Leu | Glu | | His | Phe | Trp | Thr | |
| Ile | Asn | His | Leu | | Thr | Trp | Gly | Ser | | Ile | Phe | Tyr | Phe | |

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                                                          990
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Ala Trp Phe Ala Ile Ile Leu Met Val Val Thr Cys Leu Phe Leu
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Asp Ile Ile Lys Lys Val Phe Asp Arg His Leu His Pro Thr Ser
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Thr Glu Lys Ala Gln Leu Thr Glu Thr Asn Ala Gly Ile Lys Cys
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Leu Asp Ser Met Cys Cys Phe Pro Glu Gly Glu Ala Ala Cys Ala
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Ser Val Gly Arg Met Leu Glu Arg Val Ile Gly Arg Cys Ser Pro
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Phe Ala Ala Ala Ser Ser Glu Pro Glu Glu Gly Ile Ser Val Phe
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Glu Leu Asp Tyr Asp Tyr Val Gln Ile Pro Tyr Glu Val Thr Leu
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Trp Ile Leu Leu Ala Ser Leu Ala Lys Ile Gly Phe His Leu Tyr
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His Arg Leu Pro Gly Leu Met Pro Glu Ser Cys Leu Leu Ile Leu
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Val Gly Ala Leu Val Gly Gly Ile Ile Phe Gly Thr Asp His Lys
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Ser Pro Pro Val Met Asp Ser Ser Ile Tyr Phe Leu Tyr Leu Leu
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Pro Pro Ile Val Leu Glu Gly Gly Tyr Phe Met Pro Thr Arg Pro
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Phe Phe Glu Asn Ile Gly Ser Ile Leu Trp Trp Ala Val Leu Gly
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                                     160
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Ala Leu Ile Asn Ala Leu Gly Ile Gly Leu Ser Leu Tyr Leu Ile
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                                     175
Cys Gln Val Lys Ala Phe Gly Leu Gly Asp Val Asn Leu Leu Gln
                185
                                     190
Asn Leu Leu Phe Gly Ser Leu Ile Ser Ala Val Asp Pro Val Ala
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                                     205
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Val Leu Ala Val Phe Glu Glu Ala Arg Val Asn Glu Gln Leu
                                                          Tyr
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                                     220
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Met Met Ile Phe Gly Glu Ala Leu Leu Asn Asp Gly Ile Thr Val
                230
                                     235
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Val Leu Tyr Asn Met Leu Ile Ala Phe Thr Lys Met His Lys Phe
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Glu Asp Ile Glu Thr Val Asp Ile Leu Ala Gly Cys Ala Arg Phe
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Ile Val Val Gly Leu Gly Gly Val Leu Phe Gly Ile Val Phe Gly

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Phe Ile Ser Ala Phe Ile Thr Arg Phe Thr Gln Asn Ile Ser Ala
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Ile Glu Pro Leu Ile Val Phe Met Phe Ser Tyr Leu Ser Tyr Leu
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                305
                                     310
Ala Ala Glu Thr Leu Tyr Leu Ser Gly Ile Leu Ala Ile Thr Ala
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                                                          330
Cys Ala Val Thr Met Lys Lys Tyr Val Glu Glu Asn Val Ser Gln
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                                     340
                                                          345
Thr Ser Tyr Thr Thr Ile Lys Tyr Phe Met Lys Met Leu Ser Ser
                350
                                     355
                                                          360
Val Ser Glu Thr Leu Ile Phe Ile Phe Met Gly Val Ser Thr Val
                                     370
                365
Gly Lys Asn His Glu Trp Asn Trp Ala Phe Ile Cys Phe Thr Leu
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                                                          390
                380
Ala Phe Cys Gln Ile Trp Arg Ala Ile Ser Val Phe Ala Leu Phe
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                                     400
Tyr Ile Ser Asn Gln Phe Arg Thr Phe Pro Phe Ser Ile Lys Asp
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Gln Cys Ile Ile Phe Tyr Ser Gly Val Arg Gly Ala Gly Ser Phe
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Ser Leu Ala Phe Leu Leu Pro Leu Ser Leu Phe Pro Arg Lys Lys
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Met Phe Val Thr Ala Thr Leu Val Val Ile Tyr Phe Thr Val Phe
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Ile Gln Gly Ile Thr Val Gly Pro Leu Val Arg Tyr Leu Asp Val
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                                     475
                                                          480
Lys Lys Thr Asn Lys Lys Glu Ser Ile Asn Glu Glu Leu His Ile
                                     490
                485
                                                          495
Arg Leu Met Asp His Leu Lys Ala Gly Ile Glu Asp Val Cys Gly
                500
                                     505
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His Trp Ser His Tyr Gln Val Arg Asp Lys Phe Lys Lys Phe Asp
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                                     520
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His Arg Tyr Leu Arg Lys Ile Leu Ile Arg Lys Asn Leu Pro Lys
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                                     535
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Ser Ser Ile Val Ser Leu Tyr Lys Lys Leu Glu Met Lys Gln Ala
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                                     550
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Ile Glu Met Val Glu Thr Gly Ile Leu Ser Ser Thr Ala Phe Ser
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Ile Pro His Gln Ala Gln Arg Ile Gln Gly Ile Lys Arg Leu Ser
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                                     580
Pro Glu Asp Val Glu Ser Ile Arg Asp Ile Leu Thr Ser Asn Met
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                                     595
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Tyr Gln Val Arg Gln Arg Thr Leu Ser Tyr Asn Lys Tyr Asn Leu
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Lys Pro Gln Thr Ser Glu Lys Gln Ala Lys Glu Ile Leu Ile Arg
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Arg Gln Asn Thr Leu Arg Glu Ser Met Arg Lys Gly His Ser Leu
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Pro Trp Gly Lys Pro Ala Gly Thr Lys Asn Ile Arg Tyr Leu Ser
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                                     655
                                                          660
Tyr Pro Tyr Gly Asn Pro Gln Ser Ala Gly Arg Asp Thr Arg Ala
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Ala Gly Phe Ser Gly Lys Leu Pro Thr Trp Leu Leu Cys Cys Phe
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Met Ile Gln Lys Met Ile Phe Gly Asp Leu Met Arg Phe Cys Trp
                                     490
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Leu Met Ala Val Val Ile Leu Gly Phe Ala Ser Ala Phe Tyr Ile
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Ile Phe Gln Thr Glu Asp Pro Thr Ser Leu Gly Gln Phe Tyr Asp
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Tyr Pro Met Ala Leu Phe Thr Thr Phe Glu Leu Phe Leu Thr Val
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Ile Asp Ala Pro Ala Asn Tyr Asp Val Asp Leu Pro Phe Met Phe
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                                     550
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Ser Ile Val Asn Phe Ala Phe Ala Ile Ile Ala Thr Leu Leu Met
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Leu Asn Leu Phe Ile Ala Met Met Gly Asp Thr His Trp Arg Val
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                                     580
                                                          585
Ala Gln Glu Arg Asp Glu Leu Trp Arg Ala Gln Val Val Ala Thr
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Thr Val Met Leu Glu Arg Lys Leu Pro Arg Cys Leu Trp Pro Arg
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Ser Gly Ile Cys Gly Cys Glu Phe Gly Leu Gly Asp Arg Trp Phe
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                                     625
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Leu Arg Val Glu Asn His Asn Asp Gln Asn Pro Leu Arg Val Leu
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Arg Tyr Val Glu Val Phe Lys Asn Ser Asp Lys Glu Asp Asp Gln
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Glu His Pro Ser Glu Lys Gln Pro Ser Gly Ala Glu Ser Gly
                                                          Thr
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                                                          675
Leu Ala Arg Ala Ser Leu Ala Leu Pro Thr Ser Ser Leu Ser Arg
                680
                                     685
                                                          690
Thr Ala Ser Gln Ser Ser His Arg Gly Trp Glu Ile Leu Arg
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Thr Phe Thr Val Glu Asp Ala Val Glu Thr Ile Gly Phe Gly Arg
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                                      40
Phe His Ile Ala Leu Phe Leu Ile Met Gly Ser Thr Gly Val Val
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                                                           60
Glu Ala Met Glu Ile Met Leu Ile Ala Val Val Ser Pro Val Ile
                                      70
                  65
Arg Cys Glu Trp Gln Leu Glu Asn Trp Gln Val Ala Leu Val Thr
                 80
                                      85
                                                           90
Thr Met Val Phe Phe Gly Tyr Met Val Phe Ser Ile Leu Phe Gly
                  95
                                     100
                                                          105
Leu Leu Ala Asp Arg Tyr Gly Arg Trp Lys Ile Leu Leu Ile Ser
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                                     115
                                                          120
Phe Leu Trp Gly Ala Tyr Phe Ser Leu Leu Thr Ser Phe Ala Pro
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                                     130
                                                          135
Ser Tyr Ile Trp Phe Val Phe Leu Arg Thr Met Val Gly Cys Gly
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Val Ser Gly His Ser Gln Gly Leu Ile Ile Lys Thr Glu Phe Leu
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Pro Thr Lys Tyr Arg Gly Tyr Met Leu Pro Leu Ser Gln Val Phe
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Trp Leu Ala Gly Ser Leu Leu Ile Ile Gly Leu Ala Ser Val Ile
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Ile Pro Thr Ile Gly Trp Arg Trp Leu Ile Arg Val Ala Ser Ile
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                                     205
Pro Gly Ile Ile Leu Ile Val Ala Phe Lys Phe Ile Pro Glu Ser
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                                     220
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Ala Arg Phe Asn Val Ser Thr Gly Asn Thr Arg Ala Ala Leu Ala
                230
                                     235
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Thr Leu Glu Arg Val Ala Lys Met Asn Arg Ser Val Met Pro Glu
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Gly Lys Leu Val Glu Pro Val Leu Glu Lys Arg Gly Arg Phe Ala
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Asp Leu Leu Asp Ala Lys Tyr Leu Arg Thr Thr Leu Gln Ile Trp
                275
                                     280
Val Ile Trp Leu Gly Ile Ser Phe Ala Tyr Tyr Gly Val Ile Leu
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                                     295
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Ala Ser Ala Glu Leu Leu Glu Arg Asp Leu Val Cys Gly Ser
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Ser Asp Ser Ala Val Val Thr Gly Gly Asp Ser Gly Glu Ser
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                                     325
                                                          330
Gln Ser Pro Cys Tyr
                    Cys His Met Phe Ala Pro Ser Asp Tyr Arg
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Thr Met Ile Ile Ser
                    Thr Ile Gly Glu Ile Ala Leu Asn Pro Leu
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Asn Ile Leu Gly Ile Asn Phe Leu Gly Arg Arg Leu Ser Leu Ser
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                                     370
                                                          375
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Ile Thr Met Gly Cys
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                                                          390
                                     385
Cys Thr Ser Ser Ala Gly Leu Ile Gly Phe Leu Phe Met Leu Arg
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                                     400
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Ala Leu Val Ala Ala Asn Phe Asn Thr Val Tyr Ile Tyr Thr Ala
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Glu Val Tyr Pro Thr
                    Thr Met Arg Ala Leu Gly Met Gly Thr Ser
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Gly Ser Leu Cys Arg Ile Gly Ala Met Val Ala Pro Phe Ile Ser
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                                                          450
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Gln Val Leu Met Ser Ala Ser Ile Leu Gly Ala Leu Cys Leu Phe
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Leu Leu Glu Trp Gly
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                                      40
Leu Phe Ser Ser Ser Met Arg Asn Val Gln Phe Pro Gly Met Ala
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                                      55
                                                           60
Pro Gln Asn Leu Gly Arg Val Asp Lys Phe Asn Ser Ser Ser Leu
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Met Val Val Tyr Thr Pro Ile Ser Asn Leu Thr Gln Gln Ile Met

70

65

Asn Lys Thr Ala Leu Ala Pro Leu Leu Lys Gly Thr Ser Val Ile Gly Ala Pro Asn Lys Thr His Met Asp Glu Ile Leu Leu Glu Asn Leu Pro Tyr Ala Met Gly Ile Ile Phe Asn Glu Thr Phe Ser Tvr Lys Leu Ile Phe Phe Gln Gly Tyr Asn Ser Pro Leu Trp Lys Glu Asp Phe Ser Ala His Cys Trp Asp Gly Tyr Gly Glu Phe Ser Cys Thr Leu Thr Lys Tyr Trp Asn Arg Gly Phe Val Ala Leu Gln Thr Ala Ile Asn Thr Ala Ile Ile Glu Val Ala Leu Val Phe Leu Met Ser Val Leu Leu Lys Lys Ala Val Leu Thr Asn Leu Val Val Phe Leu Leu Thr Leu Phe Trp Gly Cys Leu Gly Phe Thr Val Phe Tyr Glu Gln Leu Pro Ser Ser Leu Glu Trp Ile Leu Asn Ile Cys Ser Pro Phe Ala Phe Thr Thr Gly Met Ile Gln Ile Ile Lys Leu Asp Tyr Asn Leu Asn Gly Val Ile Phe Pro Asp Pro Ser Gly Asp Ser Tyr Thr Met Ile Ala Thr Phe Ser Met Leu Leu Leu Asp Gly Leu Ile Tyr Leu Leu Ala Leu Tyr Phe Asp Lys Ile Leu Pro Tyr Gly Asp Glu Arg His Tyr Ser Pro Leu Phe Phe Leu Asn Ser Ser Ser Cys Phe Gln His Gln Arg Thr Asn Ala Lys Val Ile Glu Lys Glu Ile Asp Ala Glu His Pro Ser Asp Asp Tyr Phe Glu Pro Val Ala Pro Glu Phe Gln Gly Lys Glu Ala Ile Arg Ile Arg Asn Val Lys Lys Glu Tyr Lys Gly Lys Ser Gly Lys Val Glu Ala Leu Lys Gly Leu Leu Phe Asp Ile Tyr Glu Gly Gln Ile Thr Ala Ile Leu Gly His Ser Gly Ala Gly Lys Ser Ser Leu Leu Asn Ile Leu Asn Gly Leu Ser Val Pro Thr Glu Gly Ser Val Thr Ile Tyr Asn Lys Asn Leu Ser Glu Met Gln Asp Leu Glu Glu Ile Arg Lys Ile Thr Gly Val Cys Pro Gln Phe Asn Val Gln Phe Asp Ile Leu Thr Val Lys Glu Asn Leu Ser Leu Phe Ala Lys Ile Lys Gly Ile His Leu Lys Glu Val Glu Gln Glu Val Gln Arg Ile Leu Leu Glu Leu Asp Met Gln Asn Ile Gln Asp Asn Leu Ala Lys His Leu Ser Glu Gly Gln Lys Arg Lys Leu Thr Phe Gly Ile Thr Ile Leu Gly Asp Pro Gln Ile Leu Leu Asp Glu Pro Thr Thr Gly Leu Asp Pro Phe Ser Arg Asp Gln Val Trp Ser Leu Leu Arg Glu Arg Arg Ala Asp His Val Ile Leu Phe Ser Thr Gln Ser Met Asp Glu Ala Asp Ile Leu Ala Asp Arg Lys Val Ile Met Ser Asn Gly Arg Leu Lys Cys Ala Gly Ser Ser Ile Phe Leu Lys Arg Arg Trp Gly Leu Gly Tyr His Leu Ser Leu His Arg Asn Glu Ile Cys Asn Pro Glu Gln Ile

| | | | | 590 | | | | | 595 | | | | | 600 |
|-----|-----|-----|-----|-------------|-----|-----|-----|-----|-------------|-----|-----|-----|-----|-------------|
| Thr | Ser | Phe | Ile | | His | His | Ile | Pro | | Ala | Lys | Leu | Lys | |
| Glu | Asn | Lys | Glu | Lys 620 | Leu | Val | Tyr | Thr | | Pro | Leu | Glu | Arg | Thr 630 |
| Asn | Thr | Phe | Pro | Asp 635 | Leu | Phe | Ser | Asp | Leu 640 | Asp | Lys | Cys | Ser | Asp 645 |
| Gln | Gly | Val | Thr | Gly 650 | Tyr | Asp | Ile | Ser | Met 655 | Ser | Thr | Leu | Asn | Glu 660 |
| Val | Phe | Met | Lys | Leu 665 | Glu | Gly | Gln | Ser | Thr 670 | Ile | Glu | Gln | Asp | Phe 675 |
| Glu | Gln | Val | Glu | Met 680 | Ile | Arg | Asp | Ser | Glu 685 | Ser | Leu | Asn | Glu | Met 690 |
| Glu | Leu | Ala | His | Ser 695 | Ser | Phe | Ser | Glu | Met 700 | Gln | Thr | Ala | Val | Ser 705 |
| _ | | _ | Leu | 710 | | | | | 715 | | | | | 720 |
| Arg | Phe | Leu | Lys | Leu 725 | Lys | Arg | Gln | Thr | Lys 730 | Val | Leu | Leu | Thr | Leu 735 |
| Leu | Leu | Val | Phe | Gly 740 | Ile | Ala | Ile | Phe | Pro 745 | Leu | Ile | Val | Glu | Asn 750 |
| Ile | Ile | Tyr | Ala | Met 755 | Leu | Asn | Glu | Lys | Ile 760 | Asp | Trp | Glu | Phe | Lys 765 |
| | | | Tyr | 770 | | | | | 775 | | | | | 780 |
| | | | Leu | 785 | | | | | 790 | | | | | 795 |
| Asp | Phe | Ile | Lys | Ser 800 | Leu | ГÀЗ | His | Gln | Asn 805 | Ile | Leu | Leu | Glu | Val 810 |
| - | - | | Glu | 815 | _ | | | | 820 | _ | | | - | 825 |
| | | | Ile | 830 | | | | | 835 | | | | | 840 |
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| | | | Ser | 860 | | | | | 865 | | | | | 870 |
| | | | Ile | 875 | | | | | 880 | | | | | 885 |
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| | | | Ile | 905 | | | | | 910 | | | | | 915 |
| | | | Asn | 920 | | | | | 925 | | | | | 930 |
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| Asn | Met | Gln | Tyr | Leu 965 | Leu | Ile | Thr | ser | Gln 970 | Ile | Val | Phe | Ala | Leu 975 |
| | | | Thr | 980 | _ | | | Ala | 985 | | | | | 990 |
| | | | Ser | 995 | | | | | 1000 | | | | | 1005 |
| Leu | Trp | Ser | Phe | Tyr 1010 | Phe | Phe | Phe | | Ser 1015 | Thr | Ile | Met | | Ser 1020 |
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| Leu | Glu | Val | Arg | Asp 1055 | Gln | Glu | His | | Arg 1060 | Glu | Phe | Pro | | Ala 1065 |
| Asn | Phe | Glu | Leu | | Ala | Thr | Asp | Phe | | Val | Суз | Phe | Ile | |
| Tyr | Phe | Gln | Thr | | | Phe | Val | Phe | | Leu | Arg | Суѕ | Met | |
| | | | | | | | | | | | | | | |

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               1115
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Ile Asp Glu Asp Glu Asp Ile Gln Thr Glu Arg Ile Arg Thr Val
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Thr Ala Leu Thr Thr Ser Ile Leu Asp Glu Lys Pro Val Ile Ile
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                                   1150
Ala Ser Cys Leu His Lys Glu Tyr Ala Gly Gln Lys Lys Ser Cys
               1160
                                    1165
                                                        1170
Phe Ser Lys Arg Lys Lys Ile Ala Ala Arg Asn Ile Ser Phe
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                                   1180
                                                        1185
Cys Val Gln Glu Gly Glu Ile Leu Gly Leu Leu Gly Pro Ser Gly
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Ala Gly Lys Ser Ser Ser Ile Arg Met Ile Ser Gly Ile Thr Lys
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Pro Thr Ala Gly Glu Val Glu Leu Lys Gly Cys Ser Ser Val Leu
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Gly His Leu Gly Tyr Cys Pro Gln Glu Asn Val Leu Trp Pro Met
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Leu Thr Leu Arg Glu His Leu Glu Val Tyr Ala Ala Val Lys Gly
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Ala Phe Lys Leu His Glu Gln Leu Asn Val Pro Val Gln Lys Leu
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Thr Ala Gly Ile Thr Arg Lys Leu Cys Phe Val Leu Ser Leu Leu
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               1295
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Gly Asn Ser Pro Val Leu Leu Asp Glu Pro Ser Thr Gly Ile
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Asp Pro Thr Gly Gln Gln Met Trp Gln Ala Ile Gln Ala Val
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Val Lys Asn Thr Glu Arg Gly Val Leu Leu Thr Thr His Asn Leu
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Gly Arg Leu Arg Cys Ile Gly Ser Ile Gln His Leu Lys Asn Lys
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Leu Gly Lys Asp Tyr
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Gln Val Thr Leu Val His Thr Glu Ile Leu Lys Leu Phe Pro Gln
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Ala Ala Gly Gln Gln Arg Tyr Ser Ser Leu Leu Thr Tyr Lys Leu
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Pro Val Ala Asp Val Tyr Pro Leu Ser Gln Thr Phe His Lys Leu
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Gln Cys Thr Leu Glu Lys Val Phe Leu Glu Leu Ser Lys Glu Gln
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Leu Glu Glu Phe Pro Met Met Arg Arg Ala Phe Glu Thr Val Ala
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Met Asp Arg Leu Leu Arg Ile Gly Lys Lys Asn Ser Ile Leu Gln
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Arg Lys Arg Ser Glu Pro Ser Pro Gly Ser Ser Gly Gly Ile Met
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Glu Gln His Leu Val Gln His Asp Arg Asp Met Ala Arg Gly Val
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                                     580
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Arg Gly Arg Ala Pro Ser Thr Gly Ala Gln Leu Ser Gly Lys Pro
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Val Leu Trp Glu Pro Leu Val His Ala Pro Leu Gln Ala Ala Ala
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Val Thr Ser Asn Val Ala Ile Ala Leu Thr His Gln Arg Gly Pro
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PCT/US01/21448 WO 02/04520

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| Leu | Tvr | Glu | Ala | 200 Glv | Ile | Thr | Val | Gly | 205 Ile | Leu | Leu | Ser | Tyr | 210 Ala |
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| | | | | 260 | | | | | 265 | | | | | 270 |
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| Met | Arg | Gly | Arg | Thr 305 | Thr | Val | Gly | Leu | Gly 310 | Leu | Val | Leu | Phe | Gln 315 |
| Gln | Leu | Thr | Gly | Gln 320 | Pro | Asn | Val | Leu | Cys 325 | Tyr | Ala | Ser | Thr | Ile 330 |
| Phe | Ser | Ser | Val | Gly 335 | Phe | His | Gly | Gly | Ser | Ser | Ala | Val | Leu | Ala 345 |
| Ser | Val | Gly | Leu | | Ala | Val | Lys | Val | | Ala | Thr | Leu | Thr | |
| Met | Gly | Leu | Val | | Arg | Ala | Gly | Arg | | Ala | Leu | Leu | Leu | |
| Gly | Cys | Ala | Leu | | Ala | Leu | Ser | Val | | Gly | Ile | Gly | Leu | |
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| Leu | Gln | Asp | Ser | | Leu | Pro | Pro | Ile | | Arg | Thr | Asn | Glu | |
| Gln | Arg | Glu | Pro | | Leu | Ser | Thr | Ala | | Lys | Thr | Lys | Pro | |
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| Ser | Ala | Leu | Pro | | Pro | Pro | Leu | Pro | | Arg | Gly | His | Ala | |
| Leu | Arg | Trp | Thr | | Leu | Leu | Cys | Leu | | Val | Phe | Val | Ser | |
| Phe | Ser | Phe | Gly | | Gly | Pro | Val | Thr | | Leu | Val | Leu | Ser | |
| Ile | Tyr | Pro | Val | | Ile | Arg | Gly | Arg | | Phe | Ala | Phe | Cys | |
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| Pro | Glu | Thr | Lys | | Gln | Ser | Leu | Ala | | Ile | Asp | Gln | Gln | |
| Gln | Lys | Arg | Arg | Phe | Thr | Leu | Ser | Phe | Gly | His | Arg | Gln | Asn | Ser |
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| Val | Arg | Ile | Lys | | Arg | Met | Asp | Ile | | Val | Val | Thr | Arg | |
| Asn | ГЛЗ | Ile | Arg | Asp 515 | Arg | Phe | Trp | Asp | Pro 520 | Gly | Pro | Ala | Ala | Asp 525 |
| Pro | Leu | Thr | Asp | Leu 530 | Arg | Tyr | Val | Trp | Gly 535 | Gly | Phe | Val | Tyr | Leu 540 |
| Gln | Asp | Leu | Val | Glu 545 | Arg | Ala | Ala | Val | Arg 550 | Val | Leu | Ser | Gly | Ala 555 |
| Asn | Pro | Arg | Ala | Gly 560 | Leu | Tyr | Leu | Gln | Gln 565 | Met | Pro | Tyr | Pro | Cys 570 |
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| | | | | 635 | | | | | 640 | | | | Leu | 645 |
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Gly Gly Met Gln Arg Lys Leu Ser Val Ala Ile Ala Phe Val Gly Gly Ser Gln Val Val Ile Leu Asp Glu Pro Thr Ala Gly Val Asp Pro Ala Ser Arg Arg Gly Ile Trp Glu Leu Leu Lys Tyr Arg Glu Gly Arg Thr Leu Ile Leu Ser Thr His His Leu Asp Glu Ala Glu Leu Leu Gly Asp Arg Val Ala Val Val Ala Gly Gly Arg Leu Cys Cys Cys Gly Ser Pro Leu Phe Leu Arg Arg His Leu Gly Ser Gly Tyr Tyr Leu Thr Leu Val Lys Ala Arg Leu Pro Leu Thr Thr Thr Asp Met Glu Gly Ser Val Asp Thr Arg Asn Glu Lys Ala Asp Gln Glu Lys Lys Asn Gly Ser Gln Gly Ser Arg Val Gly Thr Pro Gln Leu Leu Ala Leu Val Gln His Trp Val Pro Gly Ala Arg Leu Val Glu Glu Leu Pro His Glu Leu Val Leu Pro Tyr Thr Gly Ala His Asp Gly Ser Phe Ala Thr Leu Phe Arg Glu Leu Asp Thr Arg Leu Ala Glu Leu Arg Leu Thr Gly Tyr Gly Ile Ser Asp Thr Ser Leu Glu Glu Ile Phe Leu Lys Val Val Glu Glu Cys Ala Ala Asp Thr Asp Met Glu Asp Gly Ser Cys Gly Gln His Leu Cys Thr Gly Ile Ala Gly Leu Asp Val Thr Leu Arg Leu Lys Met Pro Pro Gln Glu Thr Ala Leu Glu Asn Gly Glu Pro Ala Gly Ser Ala Pro Glu Thr Asp Gln Gly Ser Gly Pro Asp Ala Val Gly Arg Val Gln Gly Trp Ala Leu Thr Arg Gln Gln Leu Gln Ala Leu Leu Leu Lys Arg Phe Leu Leu Ala Arg Arg Ser Arg Arg Gly Leu Phe Ala Gln Ile Val Leu Pro Ala Leu Phe Val Gly Leu Ala Leu Val Phe Ser Leu Ile Val Pro Pro Phe Gly His Tyr Pro Ala Leu Arg Leu Ser Pro Thr Met Tyr Gly Ala Gln Val Ser Phe Phe Ser Glu Asp Ala Pro Gly Asp Pro Gly Arg Ala Arg Leu Leu Glu Ala Leu Leu Gln Glu Ala Gly Leu Glu Glu Pro Pro Val Gln His Ser Ser His Arg Phe Ser Ala Pro Glu Val Pro Ala Glu Val Ala Lys Val Leu Ala Ser Gly Asn Trp Thr Pro Glu Ser Pro Ser Pro Ala Cys Gln Cys Ser Arg Pro Gly Ala Arg Arg Leu Leu Pro Asp Cys Pro Ala Ala Ala Gly Gly Pro Pro Pro Gln Ala Val Thr Gly Ser Gly Glu Val Val Gln Asn Gln Thr Gly Arg Asn Leu Ser Asp Phe Leu Val Lys Thr Tyr Pro Arg Leu Val Arg Gln Gly Leu Lys Thr Lys Lys Trp Val Asn Glu Val Arg Tyr Gly Phe Ser Leu Gly Gly Arg Asp Pro Gly Leu Pro Ser Gly Gln Glu Leu Gly Arg Ser Val

Glu Glu Leu Trp Ala Leu Leu Ser Pro Leu Pro Gly Gly Ala Leu 1480 1475 Asn Leu Thr Ala Trp Ala His Ser Leu Asp Asp Arg Val Leu Lys 1495 1500 1490 Ala Gln Asp Ser Leu Lys Ile Trp Phe Asn Asn Lys Gly Trp His 1510 1505 1515 Ser Met Val Ala Phe Val Asn Arg Ala Ser Asn Ala Ile Leu Arg 1525 1520 1530 Ala His Leu Pro Pro Gly Pro Ala Arg His Ala His Ser Ile Thr 1535 1540 1545 Thr Leu Asn His Pro Leu Asn Leu Thr Lys Glu Gln Leu Ser Glu 1550 1555 1560 Ala Ala Leu Met Ala Ser Ser Val Asp Val Leu Val Ser Ile Cys 1565 1570 1575 Val Val Phe Ala Met Ser Phe Val Pro Ala Ser Phe Thr Leu Val 1585 1590 1580 Leu Ile Glu Glu Arg Val Thr Arg Ala Lys His Leu Gln Leu Met 1595 1600 1605 Thr Leu Tyr Trp Leu Gly Asn Phe Leu Trp Gly Gly Leu Ser Pro 1610 1615 1620 Asp Met Cys Asn Tyr Leu Val Pro Ala Cys Ile Val Val Leu Ile 1625 1630 1635 Phe Leu Ala Phe Gln Gln Arg Ala Tyr Val Ala Pro Ala Asn Leu 1640 1645 1650 Pro Ala Leu Leu Leu Leu Leu Leu Tyr Gly Trp Ser Ile Thr 1655 1660 1665 Ala Ser Phe Phe Phe Ser Val Pro Ser Thr Pro Leu Met Tyr Pro 1670 1675 1680 Ala Tyr Val Val Leu Thr Cys Ile Asn Leu Phe Ile Gly Ile Asn 1685 1690 1695 Phe Val Leu Glu Leu Phe Ser Asp Gln Lys Gly Ser Met Ala Thr 1700 1705 1710 Arg Ile Leu Lys Gln Val Phe Leu Ile Phe Leu Gln Glu Val Ser 1715 1720 Gly Arg Gly Leu Ile Asp Met Val Arg Asn Pro His Phe Cys Leu 1730 1735 1740 Gln Ala Met Ala Asp Ala Phe Glu Arg Leu Gly Asp Arg Gln Phe 1745 1750 1755 Trp Glu Val Val Gly Lys Asn Leu Leu Ala Gln Ser Pro Leu Arg 1765 1770 1760 Pro Leu Phe Leu Leu Phe Thr Leu Leu Leu Met Val Ile Gln Gly 1775 1780 1785 Gln His Arg Ser Gln Leu Leu Pro Gln Pro Arg Val Arg Ser Leu 1790 1795 Glu Asp Glu Asp Val Ala Arg Glu Arg Glu Pro Leu Leu Gly Glu 1805 1810 1815 Ala Thr Gln Gly Asp Val Leu Val Leu Arg Arg Val Val Gln Gly 1820 1825 1830 Tyr Arg Gly Gln Arg Met Pro Ala Val Asp Asn Leu Thr Lys Val 1835 1840 1845 Arg Leu Cys Leu Gly Ile Pro Pro Gly Glu Cys Phe Gly Leu Leu 1850 1855 1860 Gly Lys Thr Ser Thr Phe Arg Met Val Thr Gly Val Asn Gly Ala 1865 1870 1875 Gly Asp Thr Leu Ala Ser Arg Gly Glu Ala Val Leu Ala Gly His 1880 1885 1890 Pro Ser Ala Ala His Leu Ser Met Gly Tyr Ser Val Ala Arg Glu 1895 1900 1905 Cvs Pro Gln Ser Asp Ala Ile Phe Glu Leu Leu Thr Gly Arg Glu 1910 1915 1920 Val Pro Glu Ala Gln His Leu Glu Leu Leu Ala Arg Leu Arg Gly 1925 1930 1935 Val Ala Gln Thr Ala Gly Ser Gly Leu Ala Arg Leu Gly Leu Ser 1940 1945 1950 Trp Tyr Ala Asp Arg Pro Ala Gly Thr Tyr Ser Gly Gly Asn Lys 1965 1955 1960 Arg Lys Leu Ala Thr Ala Leu Ala Leu Val Gly Asp Pro Ala Val

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                                    2005
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| Pro | Gly | Phe | Lys | | Met | Asp | Leu | Glu | Asp 250 | Leu | Gly | Leu | Ser | Arg 255 |
| Gln | Glu | Leu | Gly | Tyr 260 | Ser | Gly | Phe | Asn | Glu 265 | Ile | Gly | Thr | Ser | Ile 270 |
| Phe | Thr | Val | Tyr | _ | Ala | Ala | Ser | Gln | | Gly | Trp | Val | Phe | Leu 285 |
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| Val | Phe | Ile | Ala | Val 320 | Ile | Ile | Glu | Thr | Phe 325 | Ala | Glu | Ile | Arg | Val 330 |
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| Leu | Val | Ala | Va1 | Asp 365 | Val | Asn | Lys | Pro | Gln 370 | Gly | Arg | Ala | Pro | Ala 375 |
| Cys | Leu | Gln | Lys | Met 380 | Met | Arg | Ser | Ser | Val 385 | Phe | His | Met | Phe | Ile 390 |
| Leu | Ser | Met | Val | Thr 395 | Val | Asp | Val | Ile | Val 400 | Ala | Ala | Ser | Asn | Tyr 405 |
| Tyr | Lys | Gly | Glu | Asn 410 | Phe | Arg | Arg | Gln | Tyr 415 | Asp | Glu | Phe | Tyr | Leu 420 |
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| Ser | Phe | Met | Lys | | Phe | Ile | Asp | Arg | | Gln | Gln | Asp | Thr | |
| Cys | Leu | Leu | Arg | | Leu | Pro | Thr | Thr | | Ser | Ser | Ser | Cys | |
| His | Ser | Lys | Arg | | Ala | Ile | Glu | Asp | | Lys | Tyr | Ile | Asp | |

Lys Leu Arg Lys Ser Val Phe Ser Ile Arg Ala Arg Asn Leu Leu Glu Lys Glu Thr Ala Val Thr Lys Ile Leu Arg Ala Cys Thr Arg Gln Arg Met Leu Ser Gly Ser Phe Glu Gly Gln Pro Ala Lys Glu Arg Ser Ile Leu Ser Val Gln His His Ile Arg Gln Glu Arg Arg Ser Leu Arg His Gly Ser Asn Ser Gln Arg Ile Ser Arg Gly Lys Ser Leu Glu Thr Leu Thr Gln Asp His Cys Asn Thr Val Ile Tyr Arg Asn Ala Gln Arg Glu Val Ser Glu Ile Lys Met Ile Gln Glu Lys Lys Glu Leu Ala Glu Met Leu Gln Gly Lys Cys Lys Lys Glu Leu Arg Glu Ser His Pro Tyr Phe Asp Lys Pro Leu Phe Ile Val Gly Arg Glu His Arg Phe Arg Asn Phe Cys Arg Val Val Val Arg Ala Arg Phe Asn Ala Ser Lys Thr Asp Pro Val Thr Gly Ala Val Lys Asn Thr Lys Tyr His Leu Leu Tyr Asp Leu Leu Gly Leu Val Thr Tyr Leu Asp Trp Val Met Ile Ile Val Thr Ser Asp Ser Cys Ile Ser Met Met Phe Glu Ser Pro Phe Arg Arg Val Met His Ala Pro Thr Leu Gln Ile Ala Glu Tyr Val Phe Val Ile Phe Met Ser Ile Glu Leu Asn Leu Lys Ile Met Ala Asp Gly Leu Phe Phe Thr Pro Thr Ala Val Ile Arg Asp Phe Gly Gly Val Met Asp Ile Phe Ile Tyr Leu Val Ser Leu Ile Phe Leu Cys Trp Met Pro Gln Asn Val Pro Ala Glu Ser Gly Ala Gln Leu Leu Met Val Leu Arg Cys Leu Arg Pro Leu Arg Ile Phe Lys Leu Val Pro Gln Met Arg Lys Val Val Arg Glu Leu Phe Ser Gly Phe Lys Glu Ile Phe Leu Val Ser Ile Leu Leu Thr Leu Met Leu Val Phe Ala Ser Phe Gly Val Gln Leu Phe Ala Gly Lys Leu Ala Lys Cys Asn Asp Pro Asn Ile Ile Arg Arg Glu Asp Cys Asn Gly Ile Phe Arg Ile Asn Val Ser Val Ser Lys Asn Leu Asn Leu Lys Leu Arg Pro Gly Glu Lys Lys Pro Gly Phe Trp Val Pro Arg Val Trp Ala Asn Pro Arg Asn Phe Asn Phe Asp Asn Val Gly Asn Ala Met Leu Ala Leu Phe Glu Val Leu Ser Leu Lys Gly Trp Val Glu Val Arg Asp Val Ile Ile His Arg Val Gly Pro Ile His Gly Ile Tyr Ile His Val Phe Val Phe Leu Gly Cys Met Ile Gly Leu Thr Leu Phe Val Gly Val Val Ile Ala Asn Phe Asn Glu Asn Lys Gly Thr Ala Leu Leu Thr Val Asp Gln Arg Arg Trp Glu Asp Leu Lys Ser Arg Leu Lys Ile Ala Gln Pro Leu His Leu Pro Pro Arg Pro Asp Asn Asp Gly Phe Arg Ala Lys Met Tyr Asp Ile Thr Gln His Pro Phe Phe Lys Arg Thr

| | | | | | | | | | | | 4045 |
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Trp Phe Tyr Met Gln Ile Ala Pro Ile Arg Asn Glu His Glu Lys
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| Leu | Thr | Pro | Met | Asn 170 | Lys | Thr | Glu | Val | Val 175 | His | Lys | His | Ser | Arg 180 |
| Leu | Ala | Glu | Val | Leu 185 | Gln | Leu | Gly | Ser | Asp 190 | Ile | Leu | Pro | Gln | Tyr 195 |
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| | | | Lys | 215 | | | | | 220 | | | | | 225 |
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| | | | Asn | 245 | | | | | 250 | | | | | 255 |
| | | | Leu | 260 | | | | | 265 | | | | | 270 |
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| Ser | Thr | Trp | Ser | | Ser | Lys | G1y | Ile | | Thr | Glu | Lys | Val | 510 Leu 525 |
| Ser | Ile | Cys | Pro | | qaA | Met | Arg | Ala | | Ile | Суѕ | Val | His | |
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| Cys | Ala | Pro | Gly | Asp 575 | Leu | Ile | Tyr | His | | Gly | Glu | Ser | Val | |
| | | | Phe | 590 | | | | | Leu 595 | | | | | Asp 600 |
| | | | Val | 605 | | | | | Gly 610 | | | | | Asp 615 |
| Ile | Phe | Trp | Lys | Glu | Thr | Thr | Leu | Ala | His | Ala | Суз | Ala | Asn | |

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57/87

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| His | Ser | Thr | Ser | Glu 110 | Ala | Asp | Thr | Glu | Pro 115 | Cys | Val | Asp | Gly | Trp 120 |
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| Asp | Leu | Val | Cys | Asp 140 | Tyr | Gln | Ser | Leu | Lys 145 | Ser | Val | Val | Gln | Phe 150 |
| Leu | Leu | Leu | Thr | Gly 155 | Met | Leu | Val | Gly | Gly 160 | Ile | Ile | Gly | Gly | His 165 |
| Val | Ser | Asp | Arg | Phe 170 | Gly | Arg | Arg | Phe | Ile 175 | Leu | Arg | Trp | Cys | Leu 180 |
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[Continued on next page]

(54) Title: TRANSPORTERS AND ION CHANNELS

(57) Abstract: The invention provides human transporters and ion channels (TRICH) and polynucleotides which identify and encode TRICH. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with aberrant expression of TRICH.





before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

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A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12N15/12 C07K14/47 C12Q1/68 G01N33/68 C07K16/18 According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system followed by classification symbols) C12N C07K C12Q G01N Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) EMBL, WPI Data, EPO-Internal C. DOCUMENTS CONSIDERED TO BE RELEVANT Relevant to daim No. Citation of document, with indication, where appropriate, of the relevant passages 1,3,4, 11,12 DATABASE EMBL [Online] Х 7 January 2000 (2000-01-07) "Human DNA sequence from clone RP1-137F1 on chromosome 6p21.1-21.2 Contains two genes for novel members of the potassium channel subfamily K (KCNK).Contains ESTs, STSs, GSSs and a CpG island." Database accession no. AL136087 XP002212498 see nts 51861-67180 and complement thereof;/product="dJ137F1.1 (novel member of the potassium channel subfamily K)";/translation="MYRPRARA....;/db_xref=" SPTREMBL:Q9H592" -/--Further documents are listed in the continuation of box C. Patent family members are listed in annex. Special categories of cited documents: "T" later document published after the international filing date or priority date and not in conflict with the application but "A" document defining the general state of the art which is not considered to be of particular relevance cited to understand the principle or theory underlying the invention earlier document but published on or after the international "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such docudocument referring to an oral disclosure, use, exhibition or ments, such combination being obvious to a person skilled other means document published prior to the international filing date but later than the priority date claimed "&" document member of the same patent family Date of mailing of the international search report Date of the actual completion of the international search 0 9, 01, 2003 6 September 2002 Authorized officer Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016 MADDOX, A

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international application No. PCT/US 01/21448

INTERNATIONAL SEARCH REPORT

| Box I | Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet) |
|-----------|--|
| This Inte | emational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons: |
| 1. X | Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: see FURTHER INFORMATION sheet PCT/ISA/210 |
| 2. X | Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically: see FURTHER INFORMATION sheet PCT/ISA/210 |
| з. 🗌 | . Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a). |
| Box II | Observations where unity of invention is lacking (Continuation of item 2 of first sheet) |
| This Inte | rnational Searching Authority found multiple inventions in this international application, as follows: |
| | see additional sheet |
| 1. | As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims. |
| 2. | As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee. |
| 3. | As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.: |
| 4. X | No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 45 and 77 both completely, 1-44 all partially |
| Remark | The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees. |

Form PCT/ISA/210 (continuation of first sheet (1)) (July 1998)

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

Invention 1: 45 and 77 both completely, 1-44 all partially

Polypeptide and correpsonding nucleotide as defined by SEQ ID NOS:1 and 33,methods,hosts,compositions and antibodies based on said sequences

Inventions 2-32: Claims 1-108 in so far as is applicable for the sequences as defined below

Polypeptide and corresponding polynucleotide defined by SEQ ID NOS:2-32 and 34-64, each individual polypeptide sequence of 2 through 32 representing an individual invention in combination with the correpsonding polynucleotide sequence of 34 through 64, where invention 2 is represented by SEQ ID NOS:2 and 34 and each subsequent sequential pair representing another invention through to invention 32 represented by SEQ ID NOS:32 and 64;methods,hosts,compositions and antibodies based on said sequences.

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FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.1

Although claim 18 is directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.

Although claim(s)32 and 34 are directed to a diagnostic method practised on the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.

Continuation of Box I.2

Present claims 20,21,23, and 24 relate to a product/compound/method defined by reference to a desirable characteristic or property, namely agonists and antagonists of the polypeptide of claim 1. The claims cover all products/compounds/methods having this characteristic or property, whereas the application does not provide support within the meaning of Article 6 PCT and/or disclosure within the meaning of Article 5 PCT. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Independent of the above reasoning, the claims also lack clarity (Article 6 PCT). An attempt is made to define the product/compound/method by reference to a result to be achieved. Again, this lack of clarity in the present case is such as to render a meaningful search over the whole of the claimed scope impossible. Consequently, the search has not been carried out for claims 20,21,23, and 24

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

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